



UNIVERSIDADE FEDERAL DO MARANHÃO
PROGRAMA DE PÓS-GRADUAÇÃO EM BIODIVERSIDADE E
BIOTECNOLOGIA - REDE BIONORTE



**ÓLEOS ESSENCIAIS E TERPENOS COMO ESTRATÉGIA DE
CONTROLE DE CARRAPATOS E NEMATOIDES**

ANILDES IRAN PEREIRA SOUSA

São Luís - MA

2022

ANILDES IRAN PEREIRA SOUSA

**ÓLEOS ESSENCIAIS E TERPENOS COMO ESTRATÉGIA DE
CONTROLE DE CARRAPATOS E NEMATOIDES**

Tese de doutorado apresentada ao Curso de Doutorado do Programa de Pós-Graduação em Biodiversidade e Biotecnologia da Rede BIONORTE, na Universidade Federal do Maranhão, como requisito parcial para a obtenção do Título de Doutora em Biodiversidade e Biotecnologia.

Orientadora: Profa. Dra. Alexandra Martins dos Santos Soares

Coorientador: Prof. Dr. Lívio Martins Costa Junior

São Luís - MA

2022

Ficha gerada por meio do SIGAA/Biblioteca com dados fornecidos pelo(a) autor(a).
Diretoria Integrada de Bibliotecas/UFMA

IRAN PEREIRA SOUSA, ANILDES.

ÓLEOS ESSENCIAIS E TERPENOS COMO ESTRATÉGIA DE CONTROLE DE CARRAPATOS E NEMATOIDES / ANILDES IRAN PEREIRA SOUSA. -2022.

80 p.

Coorientador(a): Livio Martins Costa Junior. Orientador(a): Alexandra Martins dos Santos Soares. Tese (Doutorado) - Programa de Pós-graduação em Rede - Rede de Biodiversidade e Biotecnologia da Amazônia Legal/ccbs, Universidade Federal do Maranhão, São Luís,2022.

1. Haemonchus. 2. Parasiticidas. 3. Produtos naturais. 4. Rhipicephalus. I. Martins Costa Junior,Livio. II. Martins dos Santos Soares, Alexandra. III.Título.

ANILDES IRAN PEREIRA SOUSA

ÓLEOS ESSENCIAIS E TERPENOS COMO ESTRATÉGIA DE CONTROLE DE CARRAPATOS E NEMATOIDES

Tese de doutorado apresentada ao Curso de Doutorado do Programa de Pós-Graduação em Biodiversidade e Biotecnologia da Rede BIONORTE, na Universidade Federal do Maranhão, como requisito parcial para a obtenção do Título de Doutor em Biodiversidade e Biotecnologia.

Aprovada em 15/12/2022

Banca examinadora

 Documento assinado digitalmente
ALEXANDRA MARTINS DOS SANTOS SOARES
Data: 12/01/2023 12:06:44-0300
Verifique em <https://verificador.iti.br>

**Prof^a. Dr^a. Alexandra Martins dos Santos Soares
Presidente da banca / Docente - UFMA**

 Documento assinado digitalmente
MAYARA CRISTINA PINTO DA SILVA
Data: 13/01/2023 10:01:04-0300
Verifique em <https://verificador.iti.br>

**Prof^a. Dr^a Mayara Cristina Pinto da Silva
Docente - UFMA**

 Documento assinado digitalmente
RACHEL MELO RIBEIRO
Data: 13/01/2023 10:58:58-0300
Verifique em <https://verificador.iti.br>

**Prof^a. Dr^a Rachel Melo Ribeiro
Docente - UFMA**

 Documento assinado digitalmente
MARIA DO SOCORRO DE SOUSA CARTAGEN
Data: 13/01/2023 14:19:07-0300
Verifique em <https://verificador.iti.br>

**Prof^a. Dr^a Maria do Socorro de Sousa Cartagenes
Docente - UFMA**

 Documento assinado digitalmente
DANILO RODRIGUES BARROS BRITO
Data: 13/01/2023 14:54:48-0300
Verifique em <https://verificador.iti.br>

**Prof. Dr. Danilo Rodrigues Barros Brito
Docente - IFMA**

São Luís - MA

2022

À vida!

A Deus, por sua eterna generosidade

AGRADECIMENTOS

A Deus, por tudo vivido!

Às políticas públicas, pelo acesso ao ensino e pesquisa.

Ao Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq), pelo incentivo à pesquisa.

À Fundação de Amparo à Pesquisa e ao Desenvolvimento Científico e Tecnológico do Maranhão (FAPEMA), pela possibilidade de realização da pesquisa.

À Universidade Federal do Maranhão (UFMA), por ter sido sede de desenvolvimento deste trabalho e demais segmentos da minha formação.

Ao BIONORTE pela formação disponibilizada.

À Profa. Dra. Alexandra Martins dos Santos Soares, pelos valiosos ensinamentos e norteamento na orientação.

Ao Prof. Dr. Lívio Martins Costa Júnior pela coorientação.

Ao Biotério Central da UFMA pela manutenção dos animais.

Ao Laboratório de Bioquímica Vegetal (LBV), pela disponibilização da estrutura para realização dos testes de formulação, conhecimento científico e parceria consolidada.

Ao Laboratório de Controle de Parasitos (LCP), pela disponibilização da estrutura para realização dos testes “*in vitro*”.

Ao BIONORTE / Maranhão, e ao docente Dr. Glécio Machado de Siqueira, pela atenção recebida neste polo.

Às alunas de Iniciação Científica, Glayane Castro e Juliane Moreira, pela colaboração na execução do II capítulo.

Aos colegas do Laboratório de Bioquímica Vegetal (LBV), Amanda, Ana Carolina, Cecília, Daniela, Eduarda, Glayane, Marcelo e Wallyson, e os colegas de sala de aula, pelo convívio.

Aos docentes membros das minhas bancas de qualificação e de defesa de doutorado por disponibilizarem o seu tempo para avaliação da minha tese.

A todos que contribuíram, em algum momento, para desenvolvimento deste trabalho.

**“Deem graças ao Senhor porque Ele é bom.
O Seu amor dura para sempre!”**

Salmo 136,1.

SOUZA, Anildes Iran Pereira. Óleos essenciais e terpenos como estratégia de controle de carrapatos e nematoides. 2022. 80 f. Tese (Doutorado em Biodiversidade e Biotecnologia) - Universidade Federal do Maranhão, São Luís, 2022.

RESUMO

Carrapatos e nematoides são parasitos amplamente distribuídos de importância veterinária. O uso contínuo de acaricidas sintéticos e anti-helmínticos tem resultado em aumento da resistência, motivo pelo qual alternativas terapêuticas têm sido consideradas, como produtos naturais. A atividade parasiticida de óleos essenciais de plantas e seus componentes têm sido investigados contra carrapatos e nematoides. O objetivo deste estudo foi avaliar a ação de óleos essenciais e seus constituintes sobre o nematoide *Haemonchus contortus* e a ação de um shampoo contendo o terpeno carvacrol contra os carrapatos *Rhipicephalus (Boophilus) microplus* e *Rhipicephalus. sanguineus*. A ação de óleos essenciais obtidos de 16 cultivares de *Ocimum basilicum*, assim como a ação do linalol, metil chavicol, citral e eugenol, foi avaliada, *in vitro*, através do ensaio de inibição da eclosão de ovos do nematoide *H. contortus*. Adicionalmente, avaliou-se a ação de três cultivares, que foram simuladas usando uma combinação dos dois principais compostos de cada uma. Para os estudos com carrapatos, um shampoo contendo carvacrol foi formulado com lauril éter sulfato de sódio, cocoamidopropil betáína, lauril glucosídeo, carboximetilcelulose, metilparabeno e ácido cítrico. As características sensoriais do shampoo e o pH foram avaliados a 37, 25 e 5 °C. A eficácia do shampoo contendo carvacrol contra *R. microplus* e *R. sanguineus* foi avaliada pelo teste de imersão larval. Com relação aos ensaios sobre nematoides, os óleos essenciais de diferentes cultivares de *O. basilicum* apresentaram concentração para inibir 50% da eclosibilidade dos ovos (IC_{50}) de *H. contortus*, variando de 0,56 a 2,22 mg/mL. A cultivar com maior inibição de eclosão de ovos, napoletano, é constituída principalmente de linalol e metil chavicol. Entre os compostos individuais testados, o citral foi o mais eficaz (IC_{50} 0,30 mg/mL). A melhor combinação de compostos foi obtida com 11% de eugenol e 64% de linalol (IC_{50} 0,44 mg/mL). Como resultados dos ensaios com carrapatos, a mortalidade de larvas de *R. microplus* e *R. sanguineus* foi de 100% após o tratamento com 0,15% de shampoo contendo carvacrol (diluição 1:19 do shampoo em água). O shampoo com carvacrol apresentou-se estável nas condições analisadas. Concluímos que cultivares de *O. basilicum* apresentam diferentes eficácia sobre *H. contortus*, sendo as cultivares contendo linalol e metil chavicol as mais promissoras e, também, concluímos que o shampoo contendo carvacrol apresenta eficácia acaricida, sendo ser um potencial agente de controle de carrapatos.

Palavras-chave: Produtos naturais; Parasiticidas; *Haemonchus*; *Rhipicephalus*.

SOUSA, Anildes Iran Pereira. Essential oils and terpenes as a tick and nematode control strategy. 2022. 83 f. Tesis (PhD in Biodiversity and Biotechnology) - Federal University of Maranhão, São Luís, MA-Brazil, 2022.

ABSTRACT

Ticks and nematodes are widely distributed parasites of veterinary importance. The continuous use of synthetic acaricides and anthelmintics has resulted in increased resistance. Thus, therapeutic alternatives such as natural products have been considered. The parasitic activity of plant essential oils (EOs) and their components have been investigated against ticks and nematodes. This study aimed to evaluate essential oils and terpenes as a strategy to control nematodes and ticks. The action of essential oils (EOs) obtained from 16 cultivars of *Ocimum basilicum*, linalool, methyl chavicol, citral and eugenol was evaluated *in vitro* in the test of inhibition of the hatching of eggs of the nematode *Haemonchus contortus*. Additionally, the action of three cultivars was evaluated, which were simulated using a combination of the two main compounds of each one. For the tick studies, a shampoo containing carvacrol was formulated with sodium lauryl ether sulfate, cocoamidopropyl betaine, lauryl glucoside, carboxymethyl cellulose, methyl paraben and citric acid. The shampoo's sensory characteristics and pH were evaluated at 37, 25 and 5 °C. The effectiveness of shampoo containing carvacrol against *R. microplus* and *R. sanguineus* was evaluated by the larval immersion test. Regarding the tests on nematodes, the EOs from different cultivars of *O. basilicum* showed a concentration to inhibit 50% of the hatchability of eggs (IC_{50}) of *H. contortus*, ranging from 0.56 to 2.22 mg/mL. The cultivar with the most significant inhibition of egg hatching, napoletan, consists mainly of linalool and methyl chavicol. Among the individual compounds tested, citral was the most effective (IC_{50} 0.30 mg/mL). The best combination of compounds was obtained with 11% eugenol and 64% linalool (IC_{50} 0.44 mg/mL). As a result of the assessments on ticks, the mortality of *R. microplus* and *R. sanguineus* was 100% after treatment with 0.15% shampoo containing carvacrol (1:19 dilution of shampoo in water). The shampoo with carvacrol was stable under the conditions analyzed. We conclude that different cultivars of *O. basilicum* have different efficacies against *H. contortus*, with the cultivars containing linalool and methyl chavicol being the most promising; we also conclude that the shampoo containing carvacrol has acaricidal efficacy and is a potential tick control agent.

Keywords: Natural Products; Antiparasitic; *Haemonchus*; *Rhipicephalus*.

LISTA DE FIGURAS

- Figura 1.** Ciclo de vida do nematoide gastrointestinal *Haemonchus contortus*. L: estágios larvais..... 15
- Figura 2.** Ciclo de vida do carrapato (A) *Rhipicephalus microplus*; e do carrapato (B) *Rhipicephalus sanguineus*. 18
- Figura 3.** *Ocimum basilicum L* (manjericão). 22

LISTA DE TABELAS

Artigo 1: Essential oils from *Ocimum basilicum* cultivars: analysis of their composition and determination of the effect of the major compounds on *Haemonchus contortus* eggs

Table 1. Major compounds (%) from essential oils of cultivars and hybrid of *Ocimum basilicum* and concentrations required for achieving 50% inhibition of egg hatching in *Haemonchus contortus* (IC_{50}) with respective 95% confidence intervals (95% CI).....30

Table 2. Inhibition concentrations required for achieving 50% of egg hatching in *Haemonchus contortus* (IC_{50}) with respective 95% confidence intervals (95% CI) from major compounds and their combinations simulating cultivars of *Ocimum basilicum*.....30

Artigo 2: In vitro assessment of the acaricidal activity of a carvacrol shampoo on tick larvae

Table 1. Mortality (mean \pm SD) of *Rhipicephalus sanguineus* and *Rhipicephalus microplus* larvae, treated with different amounts of the carvacrol shampoo and with the experimental controls (Capítulo 2).....44

LISTA DE ABREVIATURAS

OEs	Óleos Essenciais
L1	Larvas de primieiro estágio
L2	Larvas de segundo estágio
L3	Larvas de terceiro estágio infectante
GINs	Gastrointestinal nematodes
IC ₅₀	Inhibition Concentration 50 %

SUMÁRIO

1. INTRODUÇÃO	12
2. REVISÃO BIBLIOGRÁFICA.....	14
2.1 <i>Haemonchus contortus</i>	14
2.2 <i>Rhipicephalus (Boophilus) microplus</i> e <i>Rhipicephalus sanguineus</i>	16
2.3. Controle de helmintos e carapatos e resistência aos antiparasitários	19
2.4 Produtos de Origem Vegetal	21
3. OBJETIVOS	23
3.1 Objetivos Gerais	23
3.2 Objetivos específicos.....	23
CAPÍTULO 1.....	24
CAPÍTULO 2.....	39
5. CONSIDERAÇÕES FINAIS	49
6. REFERÊNCIAS GERAIS.....	50

1. INTRODUÇÃO

As infecções parasitárias em ruminantes como bovinos, ovinos e caprinos, em áreas tropicais e subtropicais do mundo, podem diminuir a produtividade devido aos danos severos, incluindo a morte do animal, levando a perdas econômicas importantes (EHSAN et al., 2020).

Dentre esses parasitos, os endoparasitos, como nematoides gastrointestinais, causam efeitos negativos nesse panorama econômico (VANDE VELDE et al., 2018). *Haemonchus contortus*, por exemplo, é um dos parasitos mais relevantes no que se refere a infecção de pequenos ruminantes em todo o mundo (RODRÍGUEZ et al., 2015; EL-ASHRAM et al., 2017).

A infecção por *H. contortus* no animal é caracterizada clinicamente por anemia grave e hipoproteinemia (perda de 200-600 mL de sangue/dia), diminuição da produção de lã, má qualidade da carcaça, edema submandibular e até morte de animais infectados (EHSAN et al., 2020). Estima-se que as perdas econômicas anuais causadas pelo *H. contortus* para a indústria pecuária representaram US\$ 30-300 milhões globalmente (ROEBER et al., 2013; EMERY et al., 2016).

Além dos endoparasitos, os ectoparasitos como os carapatos representam um grande problema para a indústria pecuária. Dentre eles, destaca-se o carapato *Rhipicephalus (Boophilus) microplus*, que é encontrado em todo mundo, sendo vetor de protozoários, como *Babesia bovis* e *B. bigemina*, e da bactéria *Anaplasma marginale*, que são os principais patógenos da babesiose e anaplasmosse bovina (FUTSE, 2003; GIGLIOTTI et al., 2018). *R. microplus* leva a diferentes condições mórbidas aos seus hospedeiros, que em geral são bovinos levando a altas perdas econômicas (GRISI et al., 2014).

O carapato *R. microplus* pode infestar também burros, ovelhas, cães, cabras, dentre outros animais (BRITO; SANTOS, GUERRA 2005; POUND et al., 2010; SILVA et al., 2018). Em cães, o carapato *Rhipicephalus sanguineus* é o mais prevalente, apesar de infestar humanos ocasionalmente (RENÉ-MARTELLET et al., 2015). *R. sanguineus* é considerado um importante vetor, devido à sua capacidade de se replicar e transmitir muitos agentes bacterianos e parasitários, que incluem *Ehrlichia canis*, *Mycoplasma haemocanis*, *Babesia vogeli*, *Hepatozoon canis* e *Rickettsia conorii* (DANTAS-TORRES, 2008; SALANO-GALLEGOS, BANETH, 2011).

Diante da ameaça que estes parasitos representam para a saúde humana, animal e para a economia, é altamente relevante que haja estratégias de controle dos mesmos, o que em geral é feito com produtos químicos (OLIVEIRA et al., 2015). Entretanto, esse método tornou-se vulnerável principalmente devido ao alto nível de resistência a esses produtos convencionalmente relatados (ALBUQUERQUE et al., 2017; LOVIS et al., 2013; CASTRO JANER et al., 2015). Além disso, o efeito negativo de muitos produtos químicos no ambiente não é desejável no contexto da produção animal sustentável (ABOSHADY et al., 2020).

Neste cenário, compostos bioativos de plantas, como os óleos essenciais (OEs), surgem como alternativas potenciais de controle de carrapatos e nematoides (ANDRE et al., 2016; GARCIA-BUSTOS et al., 2019). Os OEs de plantas e seus principais constituintes (terpenóides e fenilpropanóides) apresentam atividade sobre nematoides e carrapatos (ANDRÉ et al., 2018; SANTOS et al., 2021; FERREIRA et al., 2019) e apresentam vantagens em relação aos produtos sintéticos, como baixa toxicidade ao ambiente, ao homem e animais, e possivelmente exercem baixa pressão de seleção desses parasitos (KATIKI et al., 2012; ANDRÉ et al., 2018; CASTILHO et al., 2017; FERREIRA et al., 2018; BORGES; SOUSA; BARBOSA, 2011; GOVINDARAJAN; SIVAKUMAR, 2011).

O óleo essencial (OE) obtido de diferentes cultivares de manjericão (*Ocimum basilicum* L.) é constituído principalmente por linalol, metil chavicol, citral e eugenol (VIEIRA; SIMON, 2000; PASCUAL-VILLALOBOS; BALLESTA-ACOSTA, 2003; SAJJADI, 2006; MARTINS; et al., 2010; OTTAI; AHMED; EL DIN., 2012). O OE de *O. basilicum* tem comprovada ação parasitícola (SANTOS; VOGEL; MONTEIRO, 2012; TIWARI et al., 2017).

Assim, o objetivo deste estudo foi avaliar a ação de óleos essenciais de diferentes cultivares de *Ocimum basilicum* e seus constituintes sobre o nematoide *H. contortus*, assim como desenvolver um shampoo carrapaticida, efetivo contra *R. microplus* e *R. sanguineus*, contendo o terpeno carvacrol,

2. REVISÃO BIBLIOGRÁFICA

Endoparasitos e ectoparasitos são responsáveis por perdas econômicas significativas na produção de ruminantes. As doenças infecciosas e parasitárias levam a reduções no desempenho produtivo, reprodutivo e também morte de animais (STOTZER et al., 2014).

Os helmintos gastrintestinais, em especial o nematoide *Haemonchus contortus*, são responsáveis por uma das maiores causas de perdas econômicas, principalmente em caprinos e ovinos (STEPEK et al., 2004; BESIER, 2016). O carapato *Rhipicephalus sanguineus* é um dos principais ectoparasitos de cães, sendo alvo para desenvolvimento de drogas de diversas indústrias farmacêuticas atuantes no seguimento de medicina veterinária (BECHARA et al., 2000). Como outro exemplo de carapato, o *Rhipicephalus (Boophilus) microplus*, causa grandes prejuízos e desconforto aos bovinos (CARVALHO FILHO et al., 2003).

Exemplificando, no Brasil, as perdas anuais do parasitismo sobre a produtividade do gado foram de 7,11 bilhões de dólares para nematoides gastrintestinais; 3,24 bilhões de dólares para *Rhipicephalus (Boophilus) microplus*; 2,56 bilhões de dólares para *Haematobia irritans*; 0,38 bilhões de dólares para *Dermatobia hominis*, 0,34 bilhões de dólares para *Cochliomyia hominivorax* e 0,34 bilhões de dólares para *Stomoxys calcitrans*, totalizando cerca de US\$ 13,96 bilhões (GRISI et al., 2014).

2.1 *Haemonchus contortus*

Dentre as doenças causadas por endoparasitos, destacam-se as parasitoses gastrintestinais, que causam prejuízos extremamente significativos à criação de ruminantes (COSTA et al., 2009, SILVA et al., 2018), em decorrência do crescimento retardado, perda de peso, redução no consumo de alimentos, queda da produção de leite, baixa fertilidade e até mortalidade (PAIVA; NEVES, 2009). As infecções parasitárias normalmente são mistas e compreendem diversas famílias e gêneros, sendo que as mais representativas para a produção de ruminantes, pertencem à família *Trichostrongylidae*, com destaque para os gêneros *Haemonchus* spp., *Ostertagia* spp., *Trichostrongylus* spp., *Cooperia* spp. e família *Strongylidae* representada pelos gêneros *Chabertia* pp. e *Oesophagostomum* spp. (VIVEIROS, 2009; BRITO et al., 2005).

Haemonchus contortus é um nematoide considerado o principal parasito de pequenos ruminantes, devido à alta taxa de mortalidade (STEPEK et al., 2004; BESIER, 2016; PERRY; MOENS, 2011; TSUKAHARA et al., 2021). Este parasito, de alta prevalência em zonas

climáticas tropicais, pertencente ao filo nematelmintos, classe nematoda, ordem Strongylida, família trichostrongylidae e gênero *Haemonchus* (MELO et al., 2003; CARVALHO, 2012).

O ciclo de *H. contortus* (Figura 1) inicia-se com a eclosão dos ovos na massa fecal do hospedeiro, que ocorre através de estímulos ambientais e pelas enzimas da eclosão, liberadas pelo embrião (ENGSTRÖM et al., 2016). As larvas rompem os ovos e se alimentam de bactérias no estágio (L1). No estágio infectante (L3), as larvas obtêm motilidade, e alcançam as folhagens dos pastos, em que seus hospedeiros se alimentam e ingerem estas larvas. No interior de seu hospedeiro, ocorre o processo de desembainhamento, proveniente dos estímulos a secreção de fluido rico em enzimas promovendo a digestão da bainha e a liberação das larvas no abomaso, onde passam por mais duas fases. Ao atingir a fase adulta os organismos podem chegar a tamanhos entre 1 a 3 cm de comprimento com uma cavidade bucal especializado com uma lanceta que permite fixar-se no intestino do animal (JACQUIET et al., 1998; AMARANTE et al., 2005).

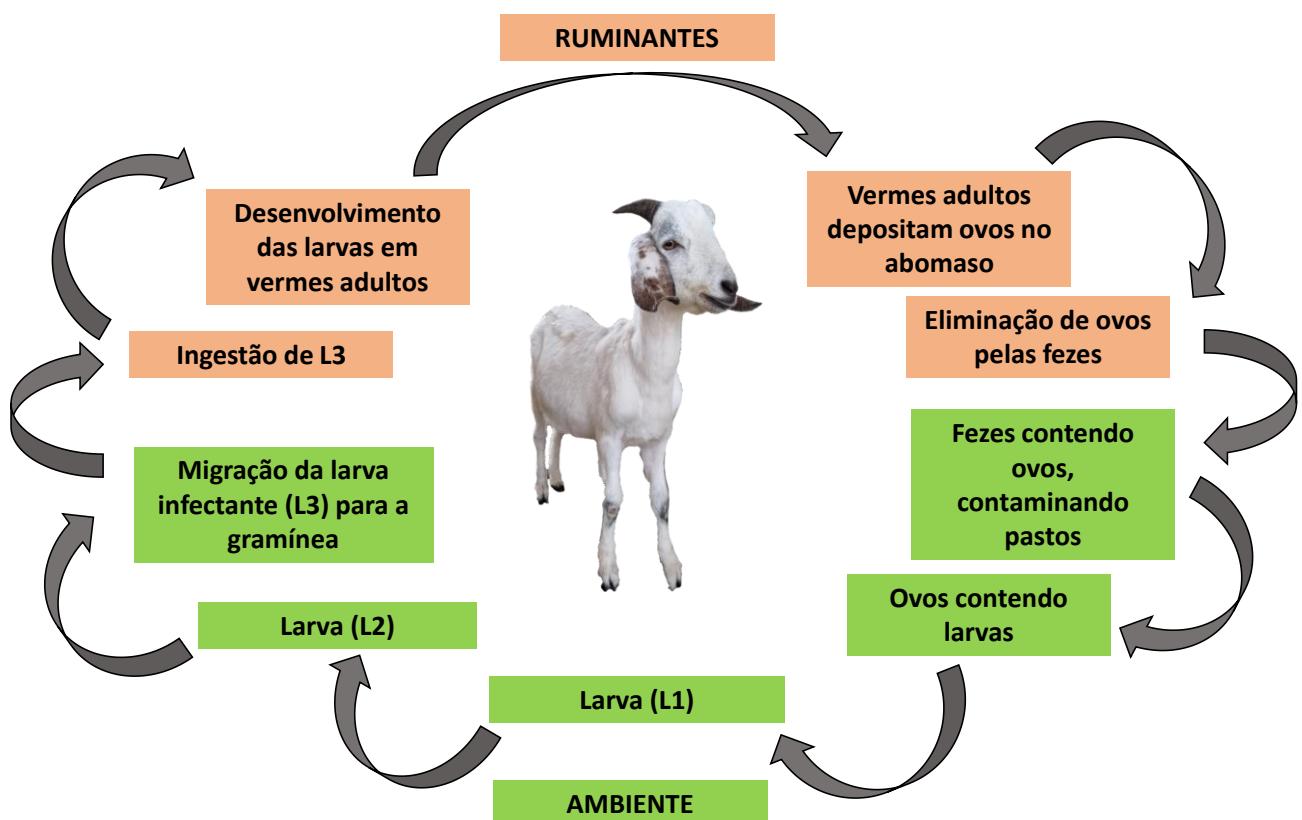


Figura 1. Ciclo de vida do nematoide gastrointestinal *Haemonchus contortus*. L1, L2 e L3: estágios larvais. Em verde, fase de vida livre e em laranja, fase parasitária.

Fonte: adaptado pela autora

2.2 *Rhipicephalus (Boophilus) microplus* e *Rhipicephalus sanguineus*

Dentre os diversos ectoparasitos que impactam a produção animal, os carapatos (subordem *Ixodidae*) são o grupo mais importante de vetores de patógenos dentro do filo Arthropoda, sendo comparáveis apenas aos mosquitos (família *Culicidae*) (KLAFKE, 2008). Eles são responsáveis pela manutenção e transmissão de muitos patógenos que afetam animais domésticos e humanos (JONGEJAN, UILENBERG, 2004).

O carapato *Rhipicephalus (Boophilus) microplus* (CANESTRINI, 1887) (Acari: Ixodidae) representa o principal ectoparasito de bovinos nas regiões tropicais e subtropicais. Foi identificado parasitando bovinos, caprinos e equinos, em estudos sobre ixodofauna de mamíferos domésticos da ilha de São Luís, Estado do Maranhão, no Brasil (BRITO; GUERRA, 2005; COSTA et al., 2008). Pertence ao reino metazoa, filo artropoda, classe aracnida, subclasse acari, superordem parasitiformes, ordem ixodida, superfamília ixodoidea, família ixodidae, subfamília rhipicephalinae, gênero *Rhipicephalus*, subgênero *boophilus*, espécie *Rhipicephalus (Boophilus) microplus*. Recentemente, este carapato teve o nome da espécie alterado, de *Bophilus microplus* para *Rhipicephalus (Boophilus) microplus*, devido a estudos utilizando metodologias taxonômicas moleculares, que demonstraram a proximidade filogenética do gênero *Boophilus* com o *Rhipicephalus* (MURREL et al., 2001; BEATI, KEIRANS, 2001; SILVEIRA; CARVALHO; PECONICK, 2014).

Por serem ectoparasitos hematófagos, se tornam os principais vetores dos três agentes causadores da tristeza parasitária bovina, um dos problemas sanitários de maior prejuízo econômico na pecuária bovina (REGITANO, PRAYAGA, 2010). Estudos comprovam ainda que altas infestações por *R. microplus* favorecem o aparecimento de outras doenças, como a miíase nos bovinos (RECK et al., 2014).

R. microplus é um carapato de um hospedeiro, preferencialmente nos bovinos (FURLONG, 2005), apresentando um estágio de vida livre e um estágio parasitário que vive no corpo do hospedeiro. O ciclo de vida deste carapato está representado na figura 2 A. O ectoparasito se alimenta e muda de larva para ninfa e de ninfa para adultos no mesmo hospedeiro em um período que dura cerca de três semanas. Após o acasalamento, a fêmea adulta inicia a fase de alimentação lenta, que dura cerca de 5 dias, quando o carapato ingere quantidades moderadas de sangue. A fase de ingurgitamento rápido leva cerca de 2 dias, em que uma grande quantidade de sangue é ingerida (ROBERTS, 1968). A fêmea totalmente ingurgitada se desprende do hospedeiro bovino e realiza a oviposição no solo. Terminada a

postura dos ovos, a fêmea morre e a larva eclode. Após a eclosão as larvas parasitam um novo hospedeiro, o mesmo é localizado pelo odor, vibrações, sombreamento, estímulo visual e pela concentração de CO₂ (SONENSHINE, 1991).

O carapato *Rhipicephalus sanguineus* (LATREILLE, 1806), descrito originalmente como *Ixodes sanguineus*, foi reclassificado como pertencente ao gênero *Rhipicephalus* por Koch em 1844, sendo a espécie-tipo desse gênero (Figura 2B). *R. sanguineus*, conhecido também como “carapato marrom”, é o carapato mais difundido no mundo. O cão doméstico é o principal hospedeiro do *R. sanguineus* em áreas urbanas e rurais (BECHARA et al., 2000). Ocionalmente, esses carrapatos podem infestar uma ampla gama de hospedeiros domésticos e selvagens, incluindo gatos, roedores, pássaros e humanos (SAXENA, 1985; DANTAS-TORRES et al., 2010). Segundo Dantas-Torres (2010), esta é uma das espécies mais estudadas devido a sua relevância do ponto de vista da saúde pública e veterinária.

Este carapato de mais ampla distribuição no mundo, encontra-se presente em todos os continentes habitados por humanos e cães domésticos (WALKER et al., 2000). É um vetor de muitos agentes de doenças, alguns deles (*Coxiellaburnetii*, *Ehrlichia canis*, *Rickettsia conorii* e *Rickettsia rickettsii*) sendo de preocupação zoonótica (DANTAS-TORRES, 2008; GRAY et al., 2013; DANTAS-TORRES et al., 2013).

Em condições favoráveis de temperatura e umidade, o ciclo biológico do *Rhipicephalus sanguineus* (Figura 2 B) se completa entre 63–91 dias (LOULY et al., 2007). Entretanto, a duração do ciclo de vida pode variar conforme sua região. No Brasil, onde as condições ambientais são bastante favoráveis, *R. sanguineus* pode completar até quatro gerações por ano (DANTAS-TORRES et al., 2006; DANTAS-TORRES, 2010).

As fêmeas adultas de *R. sanguineus* se alimentam do sangue do seu hospedeiro. Uma vez ingurgitadas, se desprendem do hospedeiro para realizar a digestão sanguínea, maturação e postura dos ovos. As fêmeas põem em média 4.000 ovos. Após a postura dos ovos, a fêmea sucumbe e os ovos são depositados em locais estratégicos, como frestas e buracos, normalmente acima do nível do solo (DANTAS-TORRES, 2008).

Após incubação, pequenas larvas eclodem e, após o enrijecimento da cutícula, passam imediatamente a procurar um hospedeiro para realização do repasto sanguíneo. Larvas se alimentam por três a 10 dias, antes de se desprenderem do hospedeiro e mudarem para ninfas. O período de muda de larva para ninfa varia de 3 a 14 dias. As ninfas se alimentam, e então se desprendem do hospedeiro (BARROS-BATTESTI, 2006; JITTAPALAPONG et al.,

2000). O período de muda de ninfa para adultos varia de 9 a 47 dias. Um estudo demonstrou que a temperatura parece interferir na especificidade de hospedeiro do *R. sanguineus*, aumentando a probabilidade desse carrapato se alimentar em seres humanos (PAROLA et al., 2008; ENCINOSA GUZMAN et al., 2016).

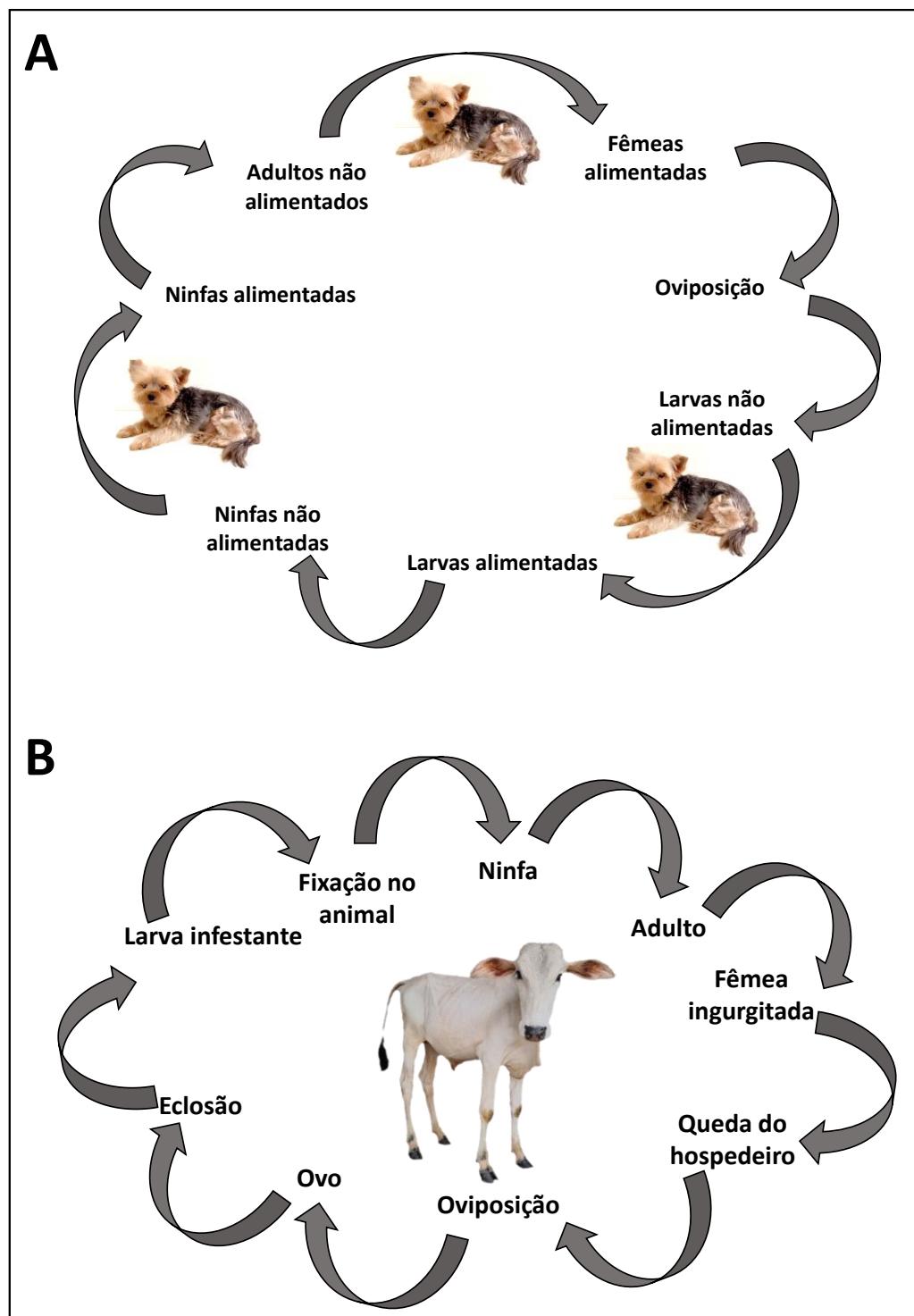


Figura 2. Ciclo de vida do carrapato (A) *Rhipicephalus microplus*; e do carrapato (B) *Rhipicephalus sanguineus*. **Fonte:** Adaptado pela autora.

2.3. Controle de helmintos e carrapatos e resistência aos antiparasitários

O controle de parasitos deve ser estudado de acordo com a dinâmica populacional destes nos rebanhos e pastagens. Desta forma, é possível desenvolver as estratégias de controle para eliminar os parasitos dos animais e prevenir a contaminação do ambiente.

É considerado um anti-helmíntico ideal, uma substância nematicida que causa a expulsão do parasito, sem causar danos significativos ao hospedeiro, com tratamento multivalente, não tóxico e rapidamente eliminado, fácil de administrar e economicamente viável (MARTIN; ROBERTSON; BJORN, 1997). Existem oito classes químicas de anti-helmínticos de amplo espectro utilizadas em ruminantes (AMARANTE; RAGOZO; SILVA, 2014). Esses anti-helmínticos convencionais, por muitos anos, demonstraram ser eficazes. No entanto, devido à resistência adquirida pelos parasitos, essas substâncias atingiram seus limites de eficácia (AMARANTE; RAGOZO; SILVA, 2014; COSTA JUNIOR et al., 2016).

Haemonchus contortus desenvolveu resistência a todas as classes de anti-helmínticos utilizados para seu controle (Lactonas macrocíclicas, benzimidazol, organofosfatos, salicilanilidas, imidazotiazol e amino-acetonitrila) (WOOSTER et al., 2001; VAN DEN BROM et al., 2015). No início de 2000, relatou-se os primeiros indícios acerca da ineficiência desses produtos sobre este parasito, pelo surgimento de isolados mostrando resistência a três ou mais anti-helmínticos de diferentes classes de medicamentos (CEZAR et al., 2008; ECHEVARRIA et al., 1996; EDDI et al., 1996; MACIEL et al., 2006), tornando o controle de *H. contortus* cada vez mais difícil (KOTZE et al., 2016).

O controle de carrapatos é fator indispensável para assegurar produtividade, benefícios econômicos e tornar a pecuária bovina uma atividade competitiva. Consiste na redução da exposição dos animais aos carrapatos em uma área específica durante um período determinado (FURLONG, 1993; BRITO et al., 2006). Os métodos de controle podem ser aplicados tanto na fase de vida livre, visto que a grande maioria dos carrapatos (95%) se encontram no ambiente, como na fase parasitária por meio do controle nos bovinos (BITTENCOURT, 1992; FURLONG, 1993). Existem vários métodos que são utilizados para controle de ectoparasitos, dentre eles estão o controle químico, biológico e orgânico, a fitoterapia, homeopatia, manejo de pastagem, rotação de pastagem, tipo da pastagem, rotação com lavouras, adubação de pastagem, aplicação de carrapaticidas no pasto, controle imunológico e vacinas (CAMPOS et al., 2012; SILVEIRA et al., 2014).

A utilização dos compostos químicos sintéticos representa o principal meio para o controle de carapatos (LOPES, 2015), no entanto o uso indiscriminado dos carrapaticidas tem determinado um grave quadro de resistência, de ordem genética, dos carapatos em relação as drogas (FURLONG, 1993; TAYLOR, 2001). Os carrapaticidas que atuam por contato direto pertencem a cinco famílias: organofosforados, amidínicos, piretróides sintéticos, fenilpirazóis e spinosad. Neste grupo ocorre a penetração do produto através da cutícula para que haja intoxicação e morte do carapato. Os carrapaticidas sistêmicos são representados por dois grupos, as lactonas macrocíclicas e as benzofenilureas que são metabolizados, distribuídos pela circulação a todo corpo do animal, atingindo os carapatos (FURLONG; SALES, 2007).

Ainda como forma de controle para carapatos, shampoos podem ser utilizados, principalmente para *R. sanguineus*. Entretanto, a maioria dos shampoos comercializados têm como princípio ativo, piretroides e devido a seleção de populações resistentes de carapatos à piretroides, a eficácia destes produtos vem reduzindo no decorrer do tempo (SUNKARA et al., 2022).

Geralmente, a ação dos compostos carrapaticidas ocorre no sistema nervoso do carapato, afetando os mecanismos de crescimento, neuropeptídeos e o sistema neuroendócrino (TAYLOR, 2001). Entretanto, alguns destes carrapaticidas não podem ser utilizados durante a lactação, nem nos bovinos de corte antes de seis semanas do abate (GEORGE et al., 2004; FURLONG; SALES, 2007; VALENTE, 2014).

A resistência aos princípios ativos químicos disponíveis comercialmente pelo *Rhipicephalus sp* vem sendo relatada em alguns estudos (FURLONG; MARTINS, 2000; CAMILO et al., 2017; RAYNAL et al., 2013). O surgimento de populações resistentes aos acaricidas vem sendo crescente em todas as regiões onde o parasito encontra condições favoráveis ao seu desenvolvimento, como na maioria dos países da América do Sul e da América Central, África do Sul e Austrália. No Brasil, a resistência das populações de carapatos aos carrapaticidas é generalizada (FARIAS CUNHA FILHO; VAZ JUNIOR, 2008; GRAF et al., 2004; HIGA et al., 2016).

Diante deste cenário, métodos alternativos para o controle dos ectoparasitos e endoparasitos são alvos de pesquisas científicas (BURKE; MILLER, 2006). A fitoterapia no controle parasitário é uma alternativa amplamente recomendado em fazendas orgânicas. Pode reduzir custos com tratamentos (VIEIRA et al., 2008), além de não apresentar efeito nocivo

ao meio ambiente (FAJIMI; TAIWO, 2005). O uso empírico de plantas ou de extratos de plantas é bastante comum e de grande importância em algumas regiões (ATHANASIADOU; KYRIAZAKIS, 2004).

2.4 Produtos de Origem Vegetal

O metabolismo vegetal divide-se em primário e secundário. O metabolismo primário origina substâncias ou metabólitos essenciais para a vida do vegetal, que são de ampla ocorrência e comuns a todas as espécies, como açúcares, proteínas, nucleotídeos e lipídeos. O metabolismo secundário origina substâncias, denominadas de metabólitos secundários, a partir dos compostos primários e são de ocorrência específica, restrito a algumas espécies e que desempenham função de adaptação, proteção, polinização, atrativa como exemplos compostos fenólicos, alcaloides, flavonoides, terpenos e outros (DEWICK, 2002).

Estima-se que mais de 30% dos fármacos são direta ou indiretamente baseados em estruturas químicas de produtos naturais (BEN-ERIK; WINK, 2017). Assim, os produtos de origem vegetal podem ser uma alternativa de grande importância em substituição as drogas sintéticas ou serem utilizadas conjuntamente (BROGLIO-MICHELETTI et al., 2009).

As plantas e seus produtos, podem apresentar atividades antimicrobianas, antivirais, fungicidas e inseticidas (CHAGAS et al., 2012), possuindo vantagens como suprimento sustentável, baixo custo, e biodegradabilidade (HEIMERDINGER et al., 2006). Acredita-se que o uso de extratos vegetais de uma forma isolada ou associada pode dificultar o processo de resistência de microrganismos (ROEL, 2001; PEREIRA et al., 2015).

Os terpenos compõem um grupo de produtos naturais, com mais de 35 mil substâncias identificadas (GERSHENZON; DUDAREVA, 2007). O crescente interesse destes compostos é atribuído à gama de propriedades biológicas, tais como efeito antitumoral, antimicrobiano, antifúngico, antiviral, anti-hiperglicêmico, analgésico, anti-inflamatório e atividades antiparasitárias (PADUCH et al., 2007). Dentre os terpenos que podem estar presentes na composição dos óleos essenciais, o timol e o carvacrol são monoterpenos aromáticos com diversas atividades biológicas. Carvacrol e timol são os constituintes principais dos OEs de várias plantas, tais como *Thymus vulgaris L.* (Lamiaceae), *Origanum compactum* (Labiatae), *Acalypha phleoides* (Euphorbiaceae), *Ocimum basilicum L.* e *Lippia sidoides* (Verbenaceae) (PEIXOTO-NEVES et al., 2010; ALMEIDA, 2015).

A atividade anti-helmíntica e carrapaticida de terpenos, principalmente carvacrol e timol, já fora relatada em diversos estudos (MONTEIRO et al., 2009; CHAGAS et al., 2012;

CRUZ, 2013; LOPES, 2015; ANDRE, 2016; ANDRE, 2017; FERREIRA, 2016), e capazes de reduzir a quantidade de ovos de nematoides nas fezes de animais (ANDRE, 2016; ANDRE, 2017); BORTOLUZZI et al., 2021; ANDRE et al., 2017; CETIN et al., 2010; DOLAN et al., 2009; KOC et al., 2013; COSTA-JÚNIOR et al., 2016; SENRA et al., 2013).

O gênero *Ocimum* comprehende mais de 50 espécies, das quais *Ocimum basilicum*, pertencente à família Lamiaceae (Figura 3), popularmente conhecido alfavaca ou manjericão de folha larga, possui elevado valor econômico. Esta planta é encontrada em locais como Ásia tropical, África, América central e América do sul. Trata-se de uma erva aromática e terapêutica amplamente utilizada na medicina popular para combater dores de cabeça, tosse, distúrbios renais e como agente antiespasmódico (ZAKARIA et al., 2008).

Adicionalmente diversas outras atividades já foram relatadas para *O. basilicum*, como atividade antisséptica, antibacteriana, anti-inflamatória, antimicrobiana, antioxidante e parasiticida (ALMEIDA et al. 2007; GÜLÇİN et al., 2007; MARTINEZ-VELAZQUEZ et al., 2011; CASTRO et al., 2017; BRUIN et al., 2017). Diferentes cultivares de *O. basilicum* foram identificados e sua classificação é feita de acordo com a concentração de seus diferentes componentes químicos. As cultivares de *O. basilicum* apresentam óleos essenciais com linalol, metil chavicol, citral e eugenol como constituintes principais, em concentrações variáveis (VIEIRA; SIMON, 2000; PASCUAL-VILLALOBOS; BALLESTA-ACOSTA, 2003; SAJJADI, 2006; MARTINS et al., 2010; OTTAI et al., 2012).



Figura 3 - *Ocimum basilicum L* (manjericão).

Fonte: UFRJ- Vanessa Ribeiro Affonso

Diante do cenário apresentado sobre a necessidade de desenvolvimento de novos produtos contra nematoides e carrapatos e levando-se em consideração que a composição de produtos naturais varia entre diferentes cultivares, este estudo foi desenvolvido para responder se diferentes cultivares de *O. basilicum* teriam diferentes potenciais anti-helmínticos, em decorrência de sua variada composição, e se, um shampoo contendo o terpeno carvacrol seria efetivo contra larvas de *Rhipicephalus microplus* e *R. sanguineus*.

3. OBJETIVOS

3.1 Objetivo Geral

Estudar a ação de óleos essenciais e terpenos como estratégia de controle de carrapatos e nematoides.

3.2 Objetivos específicos

Avaliar a ação de óleos essenciais de diferentes cultivares de *Ocimum basilicum* L. e das combinações entre seus componentes majoritários sobre *Haemonchus contortus*.

Desenvolver um shampoo contendo o terpeno carvacrol efetivo sobre larvas de *Rhipicephalus microplus* e *R. sanguineus*.

CAPÍTULO 1

Essential oils from *Ocimum basilicum* cultivars: analysis of their composition and determination of the effect of the major compounds on *Haemonchus contortus* eggs

Journal of Helminthology, 2021

Qualis A3, Fator de impacto: 1,866

doi: 10.1017/S0022149X21000080

PMID: 33745470

Essential oils from *Ocimum basilicum* cultivars: analysis of their composition and determination of the effect of the major compounds on *Haemonchus contortus* eggs

A.I.P. Sousa¹, C.R. Silva², H.N. Costa-Júnior², N.C.S. Silva², J.A.O. Pinto³,
A.F. Blank³, A.M.S. Soares¹ and L.M. Costa-Júnior²

¹ Laboratory of Plant Biochemistry, Federal University of Maranhão, São Luís, MA, Brazil.

² Laboratory of Parasite Control, Federal University of Maranhão, São Luís, MA, Brazil.

³ Agronomy Department, Federal University of Sergipe, São Cristóvão, SE, Brazil.

Abstract

The continuous use of synthetic anthelmintics against gastrointestinal nematodes (GINs) has resulted in the increased resistance, which is why alternative methods are being sought, such as the use of natural products. Plant essential oils (EOs) have been considered as potential products for the control of GINs. However, the chemical composition and, consequently, the biological activity of EOs vary in different plant cultivars. The aim of this study was to evaluate the anthelmintic activity of EOs from cultivars of *Ocimum basilicum* L. and that of their major constituents against *Haemonchus contortus*. The EOs from 16 cultivars as well the pure compound linalool, methyl chavicol, citral and eugenol were used in the assessment of the inhibition of *H. contortus* egg hatch. In addition, the composition of three cultivars was simulated using a combination of the two major compounds from each. The EOs from different cultivars showed mean Inhibition Concentration (IC_{50}) varying from 0.56 to 2.22 mg/mL. The cultivar with the highest egg-hatch inhibition, napoletano, is constituted mainly of linalool and methyl chavicol. Among the individual compounds tested, citral was the most effective (IC_{50} 0.30 mg/mL). The best combination of compounds was obtained with 11% eugenol plus 64% linalool (IC_{50} 0.44 mg/mL), simulating the Italian Large Leaf (Richters) cultivar. We conclude that different cultivars of *O. basilicum* show different anthelmintic potential, with cultivars containing linalool and methyl chavicol being the most promising; and that citral or methyl chavicol isolated should also be considered for the development of new anthelmintic formulations.

1. Introduction

Gastrointestinal nematodes (GINs) have very important economically negative effects on several animal production systems (Nieuwhof & Bishop, 2005; Lane et al., 2015). The nematode *Haemonchus contortus* is one of the most relevant GINs that infects small ruminants around the world (Rodríguez et al., 2015). The control of this nematode is performed mainly with synthetic anthelmintics. However, the increased resistance of this parasite to anthelmintics has major economic impacts on livestock worldwide (Kotze et al., 2014; Albuquerque et al., 2017). The development of natural-based formulations is being considered as an alternative. Natural products can be used to control strains of *H. contortus* that are resistant to synthetic compounds (Andre et al., 2016; Garcia-Bustos et al., 2019). Among these products, plant essential oils (EOs) and their major compounds, terpenoid and phenylpropanoid, have shown promising anthelmintic effects (Katiki et al., 2012; Castilho et al., 2017; Ferreira et al., 2018). However, the yield and composition in terms of bioactive volatile compounds depend on genetic, environmental and agronomic factors (Yang et al., 2018).

The plant *Ocimum basilicum* L., popularly known as basil, is native to Asia and grows spontaneously in tropical and sub-tropical regions (Khair et al., 2012). The *O. basilicum* EOs present compounds of interest to the food, cosmetic and also pharmaceutical industries, with a production higher than 40 tons annually (Lawrence, 1992; Telci et al., 2006). The *O. basilicum* EOs have been shown to exhibit several biological activities (Govindarajan et al., 2013; El-Soud et al., 2015; Silva et al., 2015; Güez et al., 2017), including action against *H. contortus* (Castro et al., 2017).

The distinction among numerous basil varieties is largely based on their EO composition, which is of the utmost importance to biological activities and consumers' preference (Kiferle et al., 2019). Several cultivars of *O. basilicum* present EOs with linalool, methyl chavicol (estra-gol), citral and eugenol as its main constituents, in variable concentrations (Vieira & Simon, 2000; Pascual-Villalobos & Ballesta-Acosta, 2003; Sajjadi, 2006; Martins et al., 2010; Ottai et al., 2012). These compounds have been shown to have anthelmintic activity, isolated or in a mixture, and they are also present, in different concentrations, in several other EOs (Katiki et al., 2017; Ferreira et al., 2018; Macedo et al., 2019). The standardization of efficient cultivars or the combination of natural compounds is extremely important to human and veterinary pharmaceutical industries.

Considering that different cultivars of the same plant species may have different EO

compositions, with different bioactivity, the objective of this study was to evaluate the action of EOs obtained from different cultivars of *O. basilicum*, as well as combinations of their major constituents, on *H. contortus*.

2. Materials and methods

2.1. Plant material and EOs

EOS from 15 commercial cultivars and one experimental hybrid from the Basil Genetic Breeding Program of Universidade Federal de Sergipe were evaluated. The following 15 commercial cultivars were used: Anise, Napoletano, Genovese, Ararat, Edwina, Dark Opal, Italian Large Leaf (Richters), Magical Michael, Mrs Burns, Nufar F1, Purple Ruffles (Richters Herbs, Goodwood, ON, Canada), Italian Large Leaf (Isla), Italian Large Red Leaf, Limoncino (Isla Sementes, Porto Alegre, RS, Brazil) and Maria Bonita (Blank et al., 2007), and the experimental hybrid Genovese × Maria Bonita. All EOS used were obtained from the study of Pinto et al. (2019). The cultivars were planted and collected simultaneously during the rainy season (April–June 2016), and EOS were extracted and analysed according to Pinto et al. (2019).

2.2. Parasitological procedures

The *H. contortus* strain used in the present study was isolated from a goat naturally infected, as described in Silva et al. (2021). Third larvae stage (L3) of *Haemonchus contortus* ($n = 2000$ L3/animal) was used to experimentally infect a donor sheep confirmed to be parasite-free, with five successive negative faecal egg counts (Robert & O'sullivan, 1950) performed in three- day intervals. After 30 days, the infection was confirmed by faecal egg count, faecal culture and L3 identification (Robert & O'sullivan, 1950; Van Wyk & Mayhew, 2013). Through previous in vitro tests, the *H. contortus* strain used was confirmed to be resistant to benzimidazoles and susceptible to levamisole.

The nematode eggs were recovered from faeces, according to Silva et al. (2021), and stored in a 15 mL conical tube (eggs primary solution). The total number of eggs collected was estimated in three samples of 20 mL of the primary solution, and then a solution of 1000 eggs/mL was prepared. The experimental procedures were performed according to the guidelines of the Animal Ethics Committee (CEUA) of the Federal University of Maranhão, and were approved under the protocol number 23115018061/2011-01.

2.3. Egg-hatching assay

The eggs were added to a saturated sodium chloride solution and centrifuged (1350 g) for three minutes. The floating eggs were collected (Coles et al., 1992), washed three times and re-suspended in distilled water. A suspension of 100 eggs/well was placed in a 96-well sterile plate. The EOs from all cultivars and commercial samples of their major constituents linalool, methyl chavicol, citral and eugenol purchased from Sigma-Aldrich (St Louis, MO, US), were individually diluted in 3% Tween in different concentrations (7.0, 4.9, 3.4, 2.4, 1.7, 1.2, 0.8, 0.6, 0.4 and 0.3 mg/mL). Each sample's test was performed in quadruplicate ($n = 4$), using at least six concentrations. The negative control was performed with 3% Tween. The eggs were incubated for 48 h at 27°C. Eggs and first-stage larvae were counted under an inverted microscope at 40× magnification.

2.4. Compound combinations

Linalool, methyl chavicol, citral and eugenol (Sigma-Aldrich) were used to simulate the composition of three cultivars using the two major compounds of each. Cultivars with low and intermediate IC₅₀ (concentration required to inhibit 50% of hatching) and different major compounds, were selected. The efficacy of compounds in combination to simulate Genovese (57% linalool and 27% methyl chavicol), Mrs Burns (38% linalool and 49% citral) and Italian Large Leaf (Richters) cultivars (64% linalool and 11% eugenol) was assessed in an egg-hatching assay. To complete each mixture to 100% of composition, olive oil was used. The isolated compounds and their mixtures were diluted in 3% Tween in decreasing concentrations (3.4, 2.4, 1.7, 1.2, 0.8, 0.6, 0.4 and 0.3 mg/mL). The tests of each compound were performed in quadruplicate using at least six of the above-described concentrations. The negative control was performed with 3% Tween in olive oil, at 25 mg/mL. The egg-hatching assays were performed as described above.

2.5. Statistical analysis

The results were used to determine the IC₅₀ with respective 95% confidence intervals using GraphPad Prism 8.0 software (GraphPad Inc, San Diego, CA, US). The data were initially transformed into Log (X), normalized and then non-linear regression was applied to obtain the IC₅₀ values. The differences among the IC₅₀ were assessed using the F test (GraphPad Inc). Linear regression was applied to compare the IC₅₀ values from isolated compounds, their combinations and cultivars, for which the percentages of the four major constituents are listed (GraphPad Inc).

3. Results

The EOs from different cultivars showed differences in the IC₅₀ (table 1). This difference reached up to 3.96-fold, between the Napoletano cultivar, which presented the highest efficacy (IC₅₀ 0.56 mg/mL), and the cultivars with the lowest efficacy such as Purple Ruffles and Italian Large Red Leaf (IC₅₀ 2.22 mg/mL) (table 1).

The anthelmintic activity of the major EO constituents – linalool, methyl chavicol, eugenol and citral – was also assessed. Citral was the most effective compound (IC₅₀ 0.30 mg/mL) (table 2). Two of the three assessed combinations – eugenol + linalool and methyl chavicol + linalool – showed higher efficacy than their isolated compounds. However, the combination of citral + linalool is less effective than citral alone, and more effective than only linalool.

The best result was obtained with the combination of 11% eugenol plus 64% linalool (IC₅₀ 0.30 mg/mL), simulating the Italian Large Leaf (Richters) cultivar (table 2). This Compound combination was 2.7 times more effective than EOs from the cultivar. On the other hand, the combination of 38% linalool and 49% citral was 1.4 times more effective than EOs from the cultivar Mrs Burns. The other combination used in the present study – 57% linalool and 27% methyl chavicol – did not differ statistically from the EO of the Genovese cultivar.

A negative correlation was observed at increase the concentration of citral in cultivars, compounds isolated and its combinations decreasing the IC₅₀ value (P = 0.03). No other correlation was found.

Table 1. Major compounds (%) from essential oils of cultivars and hybrid of *Ocimum basilicum* and concentrations required for achieving 50% inhibition of egg hatching in *Haemonchus contortus* (IC₅₀) with respective 95% confidence intervals (95% CI).

Cultivar	Linalool 1	Major compound Methyl chavicol	(%)	Eugenol	Citra 1	IC ₅₀	95% CI	R ²
Napoletano	26	54	0	0	0.569	0.49–0.63	0.86	
Genovese	57	27	0	0	0.62 ^b	0.60–0.63	0.98	
Ararat	16	68	0	0	0.86 ^c	0.82–0.92	0.96	
Mrs Burns	38	0	0	49	0.97 ^d	0.91–1.05	0.95	
Dark Opal	55	0	0	0	1.09 ^e	1.05–1.14	0.97	
Limoncino	9	0	0	50	1.18 ^f	1.13–1.22	0.95	
Italian Large Leaf (Richters)	64	0	11	0	1.19 ^g	1.13–1.26	0.96	
Magical Michael	64	0	20	0	1.54 ^h	1.48–1.68	0.95	
Edwina	73	0	6	0	1.66 ^{h,i}	1.57–1.75	0.97	
Italian Large Leaf (Isla)	61	0	8	0	1.69 ⁱ	1.56–1.85	0.92	
Genovese x Maria Bonita	68	0	0	20	1.72 ^j	1.69–1.77	0.99	
Anise	0	81	0	0	1.77 ⁱ	1.65–1.89	0.94	
Nufar F1	66	12	0	0	2.11 ^k	2.02–2.22	0.97	
Maria Bonita	78	0	0	0	2.13 ^k	2.05–2.22	0.98	
Purple Ruffles	18	57	0	0	2.22 ^k	2.08–2.36	0.95	
Italian Large Red Leaf	64	0	13	0	2.22 ^k	2.11–2.34	0.97	

Chemical composition from Pinto et al. (2019).

R², regression coefficient. The R² value quantifies goodness-of-fit at the non-linear regression curve performed to estimate the IC₅₀. Different superscript letters in the IC₅₀ column indicate significant differences (P < 0.05).

Table 2. Inhibition concentrations required for achieving 50% of egg hatching in *Haemonchus contortus* (IC₅₀) with respective 95% confidence intervals (95% CI) from major compounds and their combinations simulating cultivars of *Ocimum basilicum*.

Compounds	IC ₅₀	95% CI	R ²
Citral	0.30 ^a	0.29–0.32	0.98
Methyl chavicol	0.66 ^c	0.65–0.68	0.98
Eugenol	1.39 ^e	1.33–1.45	0.97
Linalool	1.75 ^e	1.65–1.86	0.94
Eugenol + linalool ¹	0.44 ^b	0.37–0.54	0.71
Methyl chavicol + linalool ²	0.65 ^d	0.63–0.68	0.97
Citral + linalool ³	0.69 ^d	0.65–0.75	0.94

R², regression coefficient. The R² value quantifies goodness-of-fit at the non-linear regression curve

performed to estimate the IC₅₀; all combinations were used with olive oil to complete 100% composition; among the different treatments, IC₅₀ values with the same superscript letter are statistically equivalent ($P < 0.05$).

¹ 11% eugenol and 64% linalool, simulating the EO from Italian Large Leaf (Richters) cultivar.

² 27% methyl chavicol and 57% linalool, simulating the EO from Genovese cultivar.

³ 49% citral and 38% linalool, simulating the EO from Mrs Burns cultivar.

4. Discussion

The EO of *O. basilicum* has several biological activities, such as antifungal (El-Soud et al., 2015), antimicrobial (Lang & Buchbauer, 2012), antiprotozoal (Almeida et al., 2007; Santoro et al., 2007), insecticidal (Rodríguez-González et al., 2019), acaricidal (Martinez-Velazquez et al., 2011) and anthelmintic (Castro et al., 2017). However, there are several basil cultivars with considerably different EO composition (Sharopov et al., 2016). This is the first study to show a statistical difference in the inhibition of *H. contortus* egg hatch – up to 3.96 times – among EOs from cultivars of the same plant species (table 1).

The egg-hatch test used in the present study has been developed as a phenotypic diagnostic of resistant nematodes for the benzimidazoles, looking at the eggs that fail to hatch (Lacey et al., 1987; FAO, 2004). The benzimidazoles inhibit embryonation and hatching by interfering with microtubules' formation (Mandelkow & Mandelkow, 1990; Coles et al., 1992). Additionally, natural compounds altered the egg's surface and increased benzimidazole activity (Silva et al., 2021). Therefore, the rationale for using egg-hatch assay in the present study was to use it as a model to search for new compounds against nematode infection, and not to target specific use in nematode eggs.

Inhibition of *H. contortus* egg hatch was previously demonstrated by the EO of one *O. basilicum* cultivar and associated with methyl chavicol and linalool as major compounds of the EO tested (Castro et al., 2017). In the present study, the EOs from Napoletano, Genovese and Ararat cultivars showed the highest anthelmintic activity, and they also contain methyl chavicol and linalool, as major compounds (table 1). However, the EOs from Nufar F1 and Purple Ruffles cultivars exhibited low efficacy against *H. contortus* while having a similar chemical composition with methyl chavicol and linalool as major compounds. Despite methyl chavicol showing a relatively good efficacy in inhibiting *H. contortus* egg hatch, Anise cultivar, which possesses 81% methyl chavicol, does not present good efficacy when compared to other cultivars with low amounts of this compound. Interestingly, the hybrid cultivar Genovese + Maria Bonita presented an intermediate anthelmintic effect when

compared with separate Genovese and Maria Bonita cultivars.

Citral, a natural combination of the isomers neral and geranial, has been shown to be effective against several nematodes, including *H. contortus*, both isolated and as the major compound of EO (Hierro et al., 2006; Macedo et al., 2019). A negative correlation between the citral concentration and efficacy was found when the results of all EO cultivars were analysed ($P = 0.03$), whereas isolated citral showed the best activity when tested alone (table 2). The composition of EOs extracted from basil varies considerably. It can be classified into four, five or seven chemical groups or chemotypes according to the main components and the statistical analysis performed (Martins et al., 2010; Liber et al., 2011; Giachino et al., 2014; Pinto et al., 2019). The variability of chemical composition from different chemotypes has been found in diverse regions of the world (Hassanpouraghdam et al., 2010). Differences in EO efficacy from the same vegetal species with different chemical compositions against parasites have been reported (Peixoto et al., 2015; Costa-Júnior et al., 2016; Lima et al., 2016). However, the efficacy could not be correlated with the chemotype or the EO's main compound, and seems to be associated with a blend of compounds (Cruz et al., 2013; Soares et al., 2016).

The combinations of the components eugenol + linalool and methyl chavicol + linalool showed more efficacy than the isolated compounds (table 2), demonstrating that the combined compounds potentialized egg-hatch inhibition. Linalool represents the main component of many species of *Ocimum*, and is considered responsible for biological activities, representing reasons for its relevance (Ravid et al., 1997).

Despite the benefits of using *O. basilicum* EOs in human and animal health, the present study has considerable importance for the bioprospection of pure or combinations of natural compounds to control ruminant nematodes. Our results clearly show differences in the bioactivity of EOs from different *O. basilicum* cultivars, related to the citral concentration. Additionally, the combinations using linalool and other compounds showed higher inhibition of *H. contortus* eggs than linalool alone, demonstrating the potential use of these compounds for the development of products for nematode control.

Acknowledgement. We thank Dr Paul Michels (The University of Edinburgh, UK) for his valuable suggestions, assistance and critical review of this manuscript.

Financial support. We thank the FAPEMA (Maranhão State Research Foundation) for financial support and for awarding a fellowship to H.N. Costa-Junior. We thank the FINEP (Funding Authority for Studies and Projects) and FAPEMA for supporting the IECT (Science and Technology Institute of Maranhão) Biotechnology. We thank CAPES (Higher Education Personnel Improvement Coordination) for awarding a fellowship to N.C.S. Silva and CNPq (Brazilian National Council for Scientific and Technological Development) for awarding a fellowship to L.M. Costa-Junior. This study was financed, in part, by CAPES (finance code 001).

Conflicts of interest. None.

Ethical standards. The authors assert that all procedures contributing to this work comply with the ethical standards of the relevant national and institutional guides on the care and use of laboratory animals.

5. References

- Albuquerque ACA, Bassetto CC, Almeida FA and Amarante AFT (2017) Development of *Haemonchus contortus* resistance in sheep under suppressive or targeted selective treatment with monepantel. *Veterinary Parasitology* 246, 112–117.
- Almeida I, Alviano DS, Vieira DP, Alves PB, Blank AF, Lopes AHCS, Alviano CS and Rosa MDSS (2007) Antigiardial activity of *Ocimum basilicum* essential oil. *Parasitology Research* 101, 443–452.
- Andre WPP, Ribeiro WLC, Cavalcante GS, et al. (2016) Comparative efficacy and toxic effects of carvacryl acetate and carvacrol on sheep gastrointestinal nematodes and mice. *Veterinary Parasitology* 218, 52–58.
- Blank AF, Souza EM, Arrigoni-Blank MDF, Paula JWA and Alves PB (2007) Maria Bonita: a linalool type basil cultivar. *Pesquisa Agropecuária Brasileira* 42, 1811–1813.
- Castilho CVV, Fantatto RR, Gaínza YA, Bizzo HR, Barbi NS, Leitão SG and Chagas ACS (2017) *In vitro* activity of the essential oil from *Hesperozygis myrtoides* on *Rhipicephalus (Boophilus) microplus* and *Haemonchus contortus*. *Brazilian Journal of Pharmacognosy* 27, 70–76.
- Castro LM, Pinto NB, Mota TO, Moura MQ, Castro LLD, Madrid IM, Freitag RA and Berne

MEA (2017) Atividade ovicida do óleo essencial e do extrato hidroalcoólico de *Ocimum basilicum* sobre nematoides gastrintestinais de ovinos. *Science and Animal Health* 5, 138–150.

Coles GC, Bauer C, Borgsteede FHM, Geerts S, Klei TR, Taylor MA and Waller PJ (1992) World association for the advancement of veterinary parasitology (W.A.A.V.P.) methods for the detection of anthelmintic resistance in nematodes of veterinary importance. *Veterinary Parasitology* 44, 35–44.

Costa-Júnior LM, Miller RJ, Alves PB, Blank AF, Li AY and Pérez de León AA (2016) Acaricidal efficacies of *Lippia gracilis* essential oil and its phytochemicals against organophosphate-resistant and susceptible strains of *Rhipicephalus (Boophilus) microplus*. *Veterinary Parasitology* 228, 60–64.

Cruz EM, Costa-Junior LM, Pinto JAO, et al. (2013) Acaricidal activity of *Lippia gracilis* essential oil and its major constituents on the tick *Rhipicephalus (Boophilus) microplus*. *Veterinary Parasitology* 195, 198–202.

El-Soud NHA, Deabes M, El-Kassem LA and Khalil M (2015) Chemical composition and antifungal activity of *Ocimum basilicum* L. essential oil. *Open Access Macedonian Journal of Medical Sciences* 3, 374–379.

FAO (2004) Resistance management and integrated parasite control in ruminants, Guidelines. Rome, Animal Production and Health Division, FAO, pp. 78–118.

Ferreira LE, Benincasa BI, Fachin AL, Contini SHT, França SC, Chagas ACS and Beleboni RO (2018) Essential oils of *Citrus aurantifolia*, *Anthemis nobile* and *Lavandula officinalis*: *in vitro* anthelmintic activities against *Haemonchus contortus*. *Parasites & Vectors* 11, 269.

Garcia-Bustos JF, Sleebs BE and Gasser RB (2019) An appraisal of natural products active against parasitic nematodes of animals. *Parasites & Vectors* 12, 1–22.

Giachino RRA, Tonk CSFA, Bayram E, Yuce S, Telci I and Furan MA (2014) RAPD and essential oil characterization of *Turkish basil* (*Ocimum basilicum* L.). *Plant Systematics and Evolution* 300, 1779–1791.

Govindarajan M, Sivakumar R, Rajeswary M and Yogalakshmi K (2013) Chemical composition and larvicidal activity of essential oil from *Ocimum basilicum* (L.) against *Culex tritaeniorhynchus*, *Aedes albopictus* and *Anopheles subpictus* (Diptera: Culicidae).

Experimental Parasitology 134, 7–11.

Güez CM, de Souza RO, Fischer P, et al. (2017) Evaluation of basil extract (*Ocimum basilicum* L.) on oxidative, anti-genotoxic and anti-inflammatory effects in human leukocytes cell cultures exposed to challenging agents. Brazilian Journal of Pharmaceutical Sciences 53, e15098.

Hassanpouraghdam MB, Gohari GR, Tabatabaei SJ and Dadpour MR (2010) Inflorescence and leaves essential oil composition of hydroponically grown *Ocimum basilicum* L. Journal of the Serbian Chemical Society 75, 1361–1368.

Hierro I, Valero A and Navarro MC (2006) In vivo larvicidal activity of monoterpenic derivatives from aromatic plants against L3 larvae of *Anisakis simplex* s.l. Phytomedicine 13, 527–531.

Katiki LM, Chagas ACS, Takahira RK, Juliani HR, Ferreira JFS and Amarante AFT (2012) Evaluation of *Cymbopogon schoenanthus* essential oil in lambs experimentally infected with *Haemonchus contortus*. Veterinary Parasitology 186, 312–318.

Katiki LM, Barbieri AME, Araujo RC, Veríssimo CJ, Louvandini H and Ferreira JFS (2017) Synergistic interaction of ten essential oils against *Haemonchus contortus* *in vitro*. Veterinary Parasitology 243, 47–51.

Khair S, Bariyah U, Khair-Ul-Bariyah S, Ahmed D and Ikram M (2012) *Ocimum basilicum*: a review on phytochemical and pharmacological studies. Pakistan Journal of Chemistry 2, 78–85.

Kiferle C, Ascrizzi R, Martinelli M, Gonzali S, Mariotti L, Pistelli L, Flamini G and Perata P (2019) Effect of iodine treatments on *Ocimum basilicum* L.: biofortification, phenolics production and essential oil composition. PLoS One 14, e0226559.

Kotze AC, Hunt PW, Skuce P, et al. (2014) Recent advances in candidate-gene and whole-genome approaches to the discovery of anthelmintic resistance markers and the description of drug/receptor interactions. International Journal for Parasitology: Drugs Drug Resistance 4, 164–184.

Lacey E, Brady RL, Prichard RK and Watson TR (1987) Comparison of inhibition of polymerisation of mammalian tubulin and helminth ovicidal activity by benzimidazole carbamates. Veterinary Parasitology 23, 105–119.

Lane J, Jubb T, Shephard R, Webb-Ware J and Fordyce GLA (2015) Final report: priority list of endemic diseases for the red meat industries. 282 pp. Sydney, Australia, Meat and Livestock Australia.

Lang G and Buchbauer G (2012) A review on recent research results (2008-2010) on essential oils as antimicrobials and antifungals. A review. Flavour and Fragrance Journal 27, 13–39.

Lawrence BM (1992) Chemical components of labiate oils and their exploitation. pp. 399–436 in Harley RM and Reynolds T (Eds) Advances in labiate science. Kew, Royal Botanic Gardens.

Liber Z, Stanko KJ, Politeoc O, Strikic F, Kolakb I, Milosc M and Satovicb Z (2011) Chemical characterization and genetic relationships among *Ocimum basilicum* L. cultivars. Chemistry & Biodiversity 8, 1978–1989.

Lima A, Carvalho JF, Peixoto MG, Blank AF, Borges LM and Costa Junior LM (2016) Assessment of the repellent effect of *Lippia alba* essential oil and major monoterpenes on the cattle tick *Rhipicephalus microplus*. Medical and Veterinary Entomology 30, 73–77.

Macedo ITF, Oliveira LMB, André WPP, et al. (2019) Anthelmintic effect of *Cymbopogon citratus* essential oil and its nanoemulsion on sheep gastrointestinal nematodes. Revista Brasileira de Parasitologia Veterinária 28, 522–527.

Mandelkow E and Mandelkow EM (1990) Microtubular structure and tubulin polymerization. Current Opinion in Cell Biology 2, 3–9.

Martinez-Velazquez M, Castillo-Herrera GA, Rosario-Cruz R, Flores-Fernandez JM, Lopez-Ramirez J, Hernandez-Gutierrez R and Del Carmen Lugo-Cervantes E (2011) Acaricidal effect and chemical composition of essential oils extracted from *Cuminum cyminum*, *Pimenta dioica* and *Ocimum basilicum* against the cattle tick *Rhipicephalus (Boophilus) microplus* (Acari: Ixodidae). Parasitology Research 108, 481–487.

Martins AGLA, Nascimento AR, Filho JEM, Filho NEM, Souza AG, Aragão NE and Silva DSV (2010) Atividade antibacteriana do óleo essencial do manjericão frente a sorogrupos de *Escherichia coli* enteropatogênica isolados de alfaces. Ciencia Rural 40, 1791–1796.

Nieuwhof GJ and Bishop SC (2005) Costs of the major endemic diseases of sheep in Great Britain and the potential benefits of reduction in disease impact. Animal Science 81, 23–29.

Ottai MES, Ahmed SS and El Din MM (2012) Genetic variability among some quantitative characters, insecticidal activity and essential oil composition of two Egyptian and French sweet basil varieties. Australian Journal of Basic and Applied Sciences 6, 185–192.

Pascual-Villalobos MJ and Ballesta-Acosta MC (2003) Chemical variation in an *Ocimum basilicum* germplasm collection and activity of the essential oils on *Callosobruchus maculatus*. Biochemical Systematics and Ecology 31, 673–679.

Peixoto MG, Costa-Júnior LM, Blank AF, et al. (2015) Acaricidal activity of essential oils from *Lippia alba* genotypes and its major components carvone, limonene, and citral against *Rhipicephalus microplus*. Veterinary Parasitology 210, 118–122.

Pinto JAO, Blank AF, Nogueira PCL, Arrigoni-Blank MF, Andrade TM, Sampaio TS and Pereira KLG (2019) Chemical characterization of the essential oil leaves of basil genotypes cultivated in different seasons. Boletín Latinoamericano y del Caribe de Plantas Medicinais y Aromaticas 18, 58–70.

Ravid U, Putievsky E, Katzir I and Lewinsohn E (1997) Enantiomeric composition of linalool in the essential oils of *Ocimum* species and in commercial basil oils. Flavour and Fragrance Journal 12, 293–296.

Robert FHS and O'Sullivan PJ (1950) Methods for egg counts and larvae cultures for strongyles infecting. Australian Journal of Agricultural Research 1, 2–99.

Rodríguez-González Á, Álvarez-García S, González-López Ó, Silva F and Casquero PA (2019) Insecticidal properties of *Ocimum basilicum* and *Cymbopogon winterianus* against *Acanthoscelides obtectus*, insect pest of the common bean (*Phaseolus vulgaris*, L.). Insects 10, 151.

Rodríguez AV, Goldberg V, Viotti H and Ciappesoni G (2015) Early detection of *Haemonchus contortus* infection in sheep using three different faecal occult blood tests. Open Veterinary Journal 5, 90–97.

Sajjadi SE (2006) Analysis of the essential oils of two cultivated basil (*Ocimum basilicum* L.) from Iran. Daru – Journal of Faculty of Pharmacy 14, 128–130.

Santoro GF, Cardoso MG, Guimarães LGL, Mendonça LZ and Soares MJ (2007) *Trypanosoma cruzi*: activity of essential oils from *Achillea millefolium* L., *Syzygium aromaticum* L. and *Ocimum basilicum* L. on epimastigotes and trypomastigotes. Experimental

Parasitology 116, 283–290.

Sharopov FS, Satyal P, Ali NAA, Pokharel S, Zhang H, Wink M, Kukaniev MA and Setzer WN (2016) The essential oil compositions of *Ocimum basilicum* from three different regions: Nepal, Tajikistan, and Yemen. *Chemistry & Biodiversity* 13, 241–248.

Silva VA, Sousa JP, Guerra FQS, Pessôa HLF, Freitas AFR, Alves LBN and Lima EO (2015) Antibacterial activity of *Ocimum basilicum* essential oil and linalool on bacterial isolates of clinical importance. *International Journal of Pharmacognosy and Phytochemical Research* 7, 1066–1071.

Silva CR, Lifschitz AL, Macedo S, Campos N, Viana-Filho M, Alcântara A, Araújo JG, Alencar L and Costa-Junior LM (2021) Combination of synthetic anthelmintics and monoterpenes: assessment of efficacy, and ultrastructural and biophysical properties of *Haemonchus contortus* using atomic force microscopy. *Veterinary Parasitology* 290, 109345.

Soares AMS, Penha TA, Araújo SA, Cruz EMO, Blank AF and Costa-Júnior LM (2016) Assessment of different *Lippia sidoides* genotypes regarding their acaricidal activity against *Rhipicephalus (Boophilus) microplus*. *Revista Brasileira de Parasitologia Veterinária* 25, 401–406.

Telci I, Bayram E, Yilmaz G and Avci B (2006) Variability in essential oil composition of Turkish basil (*Ocimum basilicum* L.). *Biochemical Systematics and Ecology* 34, 489–497.

Van Wyk JA and Mayhew E (2013) Morphological identification of parasitic nematode infective larvae of small ruminants and cattle: a practical lab guide. *Onderstepoort Journal of Veterinary Research* 80, 539.

Vieira RF and Simon JE (2000) Chemical characterization of basil (*Ocimum spp.*) found in the markets and used in traditional medicine in Brazil. *Economic Botany* 54, 207–216.

Yang L, Wen KS, Ruan X, Zhao YX, Wei F and Wang Q (2018) Response of plant secondary metabolites to environmental factors. *Molecules* 23, 762.

CAPÍTULO 2

In vitro assessment of the acaricidal activity of a carvacrol shampoo on tick larvae

Experimental Parasitology, 2022

Qualis A4, Fator de Impacto: 2.132

doi.org/10.1016/j.exppara.2022.108364

PMID: 36027929

In vitro assessment of the acaricidal activity of a carvacrol shampoo on tick larvae

Anildes I P Sousa¹, Glayane J S Castro¹, Caio P Tavares², Tássia L do Vale², Livio M Costa-Junior^{3*}, Alexandra M S Soares^{4**}

¹Laboratory of Plant Biochemistry, Federal University of Maranhão, São Luís, MA, Brazil.

²Laboratory of Parasite Control, Federal University of Maranhão, São Luís, MA, Brazil.

³Laboratory of Parasite Control, Federal University of Maranhão, São Luís, MA, Brazil. Electronic address: livio.martins@ufma.br.

⁴Laboratory of Plant Biochemistry, Federal University of Maranhão, São Luís, MA, Brazil. Electronic address: alexandra.ufma@gmail.com.

Abstract

Ticks are a widely distributed arthropod of veterinary importance. Resistance of ticks to synthetic acaricides has become widespread, warranting the development of new drugs for tick management. Carvacrol is a volatile monoterpenone, with promising results against various species of ticks; however, to be used for therapeutic purposes, carvacrol must be included in a formulation that makes its application feasible. This study aims to develop a formulation of a carvacrol-containing shampoo that is effective against two species of ticks: *Rhipicephalus sanguineus* and *R. microplus*. Shampoo sensory characteristics and pH were evaluated at 37, 25 and 5 °C, for a maximum of 15 days. The shampoo remained stable at 25 and 5 °C. The efficacy of the carvacrol-containing formulation against two species of ticks was assessed by the larval immersion test. Mortality of both tick species was significantly higher for the carvacrol shampoo than for a carvacrol hydroalcoholic solution. In conclusion, the carvacrol-containing shampoo showed larvicidal efficacy on ticks.

1. Introduction

The brown dog tick, *Rhipicephalus sanguineus* sensu lato, is one of the most widely distributed tick species globally and preferentially parasitizes dogs, but may also be found feeding on other hosts, including humans (Dantas-Torres, 2008). The management of *R. sanguineus* is mostly based on the use of synthetic chemical acaricides. However, the intensive use of these compounds has led to the selection of acaricide-resistant tick strains (Borges et al., 2007; Rodriguez-Vivas et al., 2017; Becker et al., 2019). Consequently, there is a need for the development of alternative approaches for tick management, and some plant-based compounds have been evaluated (Ellse and Wall, 2013).

Carvacrol is a volatile plant monoterpene which is currently classified as GRAS (Generally Recognized As Safe) and approved for use in human food (EAFUS, 2006; Hyldgaard et al., 2012; European Parliament and Council, 1996). Importantly, carvacrol has shown promising results when used against various species of ticks, including *R. sanguineus* (Araújo et al., 2016; Novato et al., 2015; Tabari et al., 2017). Natural or synthetic chemical substances can be included in pesticide formulations in order to make their application feasible (York, 2016). Thus, effectiveness, stability, and ease of application are among the criteria that these formulations must satisfy (Díaz et al., 2019; Ferreira et al., 2017). For the development of an acaricide formulation containing carvacrol, its low solubility in water must be considered, and, for example, the use of colloidal distribution systems, may be required (Ryu et al., 2018). Because acaricides are typically applied topically (Dantas-Torres, 2008), shampoo formulations emerge as viable alternatives, having already shown good efficiency in the delivery of other acaricides (Franc and Cadiergues, 1999; Heukelbach et al., 2006; Schuele et al., 2008).

The current study aimed to formulate a shampoo containing carvacrol against the brown dog tick *R. sanguineus* and the cattle tick *Rhipicephalus microplus*, used here as model organism. The cattle tick *R. microplus* is an excellent model for testing acaricide formulations, due to its ease of maintenance in experimental animals and the large numbers of ticks that can be obtained for testing, when compared to other tick species. Even though it is uncommon to use shampoos in cattle, we tested the acaricide formulation on these two species as a proof of concept, and because it is of the utmost importance to test whether the acaricide formulation is tick species-specific.

2. Materials and methods

2.1. Shampoo formulation

The carvacrol shampoo was formulated at room temperature with the following raw materials: 14.0 g sodium lauryl ether sulfate, 1.0 g 30% cocamidopropyl betaine, and 0.5 g lauryl glucoside as surfactants; 0.75 g carboxymethyl cellulose as gelling agent; 0.2 g methylparaben as preservative; 10% citric acid (q.s.) as acidulant; 3.0 g carvacrol (W224502, Sigma-Aldrich, St. Louis, MO, USA), and ultrapure water (q. s. 100 g) as base solution. The surfactants and carvacrol were mixed until completely homogenized. Next, citric acid was added to adjust the pH to 5.0. The carboxymethyl cellulose and methylparaben (previously solubilized in water at 100 °C) were then added. Finally, the mass of formulation was adjusted to 100 g with ultrapure water. At each addition, the formulation was completely homogenized with a glass rod. A shampoo formulation without carvacrol was also prepared.

2.1. Sensory evaluation and pH stability

Sensory evaluation and pH stability tests were performed in triplicate, at 37–2 °C, 25–2 °C or 5–2 °C, without direct light exposure, using 100 mL of shampoo sealed hermetically in amber glass flasks. The results were recorded after 1, 7, and 15 days of preparation. The sensory evaluation tests consisted of judging any changes in color, odor, and appearance and results were ranked in three categories: normal, without alteration; slightly modified; and intensely modified (ANVISA, 2004). The pH was measured with universal pH indicator strips (gradation 1.0, range 0–14). The accuracy of the indicator strips was confirmed by testing them against buffer solutions of known pH.

2.2. Tick collection and maintenance

Naturally detached engorged *R. sanguineus* and *R. microplus* females were obtained from artificially-infested New Zealand rabbits and calves, respectively, and maintained at 27–1 °C, 80% humidity (Biological Oxygen Demand) until oviposition was completed. Eggs were collected and incubated for hatching. Larvae aged 14–21 days were used in larval immersion tests. The experimental procedures were approved by the Federal University of Maranhão (UFMA) ethics committee under protocol number 23115.005443/2017–51.

2.3. Larval immersion test

The larval immersion test was performed according to Klafke et al. (2006). Carvacrol shampoo and the shampoo without carvacrol were prepared and immediately diluted in water to be used in the assay. A carvacrol hydroalcoholic solution (3.0 g carvacrol diluted in 50% ethanol solution, q.s. 100 g) was also used. Water, 50% ethanol, and the non-carvacrol

shampoo were considered as negative controls. The formulation samples were diluted at a ratio of 1:16, 1:19, 1:23, and 1:47 in water. Approximately 500 larvae were immersed for 10 min in each treatment solution and then transferred to a filter paper to dry. Subsamples of approximately 100 larvae were transferred to a clean dry filter paper (8.5×7.5 cm) that was folded and closed with plastic clips. The packets were incubated at 27 ± 1 °C and relative humidity $\geq 80\%$ for 24 h. After incubation, dead and live larvae were counted: immobile ticks were considered dead. Three independent repetitions of the experiment were conducted for each experimental group. The statistical analysis of mortality data from the larval immersion test was performed using GraphPad Prism 8.0 software (version 8, GraphPad Inc., San Diego, CA, USA). The mean values for each Treatment were compared by analysis of variance (ANOVA), followed by Tukey's test ($p < 0.05$) to compare differences between specific groups.

3. Results and discussion

In this study, the shampoo containing carvacrol exhibited a pearl-like color, the characteristic carvacrol odor, pH 5.0, and the same color at 5, 25 and 37 °C. After 1, 7, and 15 days of preparation, at 5 and 25 °C, it remained homogeneous, and was rated normal, without alteration. However, a phase separation occurred at 37 °C after 7 days and the shampoo was rated as slightly modified after 7 days and intensely modified after 15 days. The incorporation of carvacrol in formulations is important for the stability of the product, by preventing degradation and microbiological contamination, facilitating its application, and enhancing the effect of the active compound (Díaz et al., 2019; Ferreira et al., 2017).

The carvacrol-containing shampoo was highly effective against both tick species, leading to 100% mortality in *R. microplus* and *R. sanguineus* after treatment with 0.125% of carvacrol shampoo (1:23 dilution of the shampoo in water) and with 0.15% of carvacrol shampoo (1:19 dilution of the shampoo in water), respectively (Table 1). The carvacrol hydro-alcoholic solution had no efficacy against *R. sanguineus* at the tested concentrations and showed only low efficacy (21.6 2.5% mortality at 0.175%, 1:16 dilution ratio) against *R. microplus* at the highest concentration used (Table 1).

Table 1. Mortality (mean ± SD) of *Rhipicephalus sanguineus* and *Rhipicephalus microplus* larvae, treated with different amounts of the carvacrol shampoo and with the experimental controls.

Treatment	Dilution ratio	Carvacrol concentration (%)	Mortality (%)	
			<i>R. sanguineus</i>	<i>R. microplus</i>
Water	-	-	0.0 ± 0.0 ^a 0.0 ± 0.0 ^a	0.0 ± 0.0 ^a 0.0 ± 0.0 ^a
50% Ethanol	-	-	0.0 ± 0.0 ^a 0.0 ± 0.0 ^a	0.0 ± 0.0 ^a 0.0 ± 0.0 ^a
Non-carvacrol shampoo	1:16	-	0.0 ± 0.0 ^a 0.0 ± 0.0 ^a	0.0 ± 0.0 ^a 0.0 ± 0.0 ^a
	1:19	-	0.0 ± 0.0 ^a 0.0 ± 0.0 ^a	0.0 ± 0.0 ^a 0.0 ± 0.0 ^a
	1:23	-	0.0 ± 0.0 ^a 0.0 ± 0.0 ^a	0.0 ± 0.0 ^a 0.0 ± 0.0 ^a
	1:47	-	0.0 ± 0.0 ^a 0.0 ± 0.0 ^a	0.0 ± 0.0 ^a 0.0 ± 0.0 ^a
Carvacrol hydroalcoholic solution	1:16	0.175	0.0 ± 0.0 ^a 0.0 ± 0.0 ^a	21.6 ± 2.5 ^b
	1:19	0.15	0.0 ± 0.0 ^a 0.0 ± 0.0 ^a	3.4 ± 1.5 ^a
	1:23	0.125	0.0 ± 0.0 ^a 0.0 ± 0.0 ^a	16.6 ± 5.4 ^b
	1:47	0.0625	0.0 ± 0.0 ^a 0.0 ± 0.0 ^a	0.0 ± 0.0 ^a 0.0 ± 0.0 ^a
Carvacrol shampoo	1:16	0.175	100.0 ± 0.0 ^c	100.0 ± 0.0 ^d
	1:19	0.15	100.0 ± 0.0 ^c	100.0 ± 0.0 ^d
	1:23	0.125	97.3 ± 1.4 ^b	100.0 ± 0.0 ^d
	1:47	0.0625	97.3 ± 1.8 ^b	39.4 ± 5.7 ^c

Mean values, followed by different letters in the same column, are significantly different ($p < 0.05$).

Carvacrol has been studied for several years as a bactericide, fungicide, acaricide, and insecticide because of the growing interest for active compounds that are safe for human and animal health and the environment and that exert weak selection pressure leading to resistance (Abbaszadeh et al., 2014; Araújo et al., 2016; Park et al., 2017; Ryu et al., 2018). Even though the acaricidal activity of carvacrol against ticks including *R. sanguineus* has already been demonstrated (Araújo et al., 2016; Costa-Júnior et al., 2016; Cruz et al., 2013; Novato et al., 2018; Senra et al., 2013a, 2013b), carvacrol-containing formulations have been poorly explored against ticks, especially *R. sanguineus* (Lima et al., 2017, 2019; Novato et al., 2019). The bioactivity of carvacrol, which is frequently diluted in ethanol for the larval immersion test against unengorged *Rhipicephalus* larvae (Coelho et al., 2020; Daemon et al., 2012; Scoralik et al., 2012), can be partly explained based on its interaction with the tick's surface. The cuticle, the outermost part of the integument covering ticks, is composed mainly of lipids,

polyphenols, proteins, and chitin (Hackman and Filshie, 1982; Lees, 1947). Because of its liposolubility, carvacrol has been suggested to interact strongly with the cuticle.

The mortality rates found in the tests comparing the carvacrol-containing shampoo and the carvacrol hydroalcoholic solution (Table 1) suggest a possible synergistic effects of the components of the shampoo formulation. The intrinsic properties of some compounds may have acaricidal effect or contribute to it. For instance, carboxymethyl cellulose (at 0.1%) alone is capable of inhibiting larval hatching in engorged *R. microplus* females (de Mendonça et al., 2019). Similarly, high mortality rates of *R. sanguineus* were observed when glycerin, which also forms a film on the surfaces where it is applied, was used in formulations associated with the terpene thymol (Delmonte et al., 2017). The authors suggested that this effect may be attributable to the increased water loss through the cuticle or to the film produced, which causes occlusion of the gas exchange channels. Most water loss in ticks occurs through the cuticle and spiracles (Lees, 1947), and the lipid layer in the cuticle plays an important role in regulating this occurrence. The higher efficacy of carvacrol shampoo against ticks compared to hydroalcoholic solution may be explained by the use of surfactants in the former increasing the terpene efficacy.

This is a proof-of-concept study, in which only the in vitro efficacy of one shampoo formulation was tested. Nonetheless, given the high efficiency of the carvacrol shampoo, even at low concentrations, the data presented here provide considerable support for new studies including, the assessment of *in vivo* effectiveness, allergenicity, and potential to cause dermatitis, contributing to the validation of integrated tick management strategies. In conclusion, the carvacrol shampoo developed was effective against *R. sanguineus* and *R. microplus* larvae under an in vitro study.

Credit author statement

Anildes Iran Pereira Sousa, carried out the experiment, collected samples, and data, and performed laboratory analyses.

Glayane de Jesus Soares Castro, collected samples and data, performed laboratory analyses, and prepared the first draft of the manuscript.

Caio Pavão Tavares and Tássia Lopes do Vale, carried out the experiment, collected samples, and data, and performed laboratory analyses.

Livio Costa-Junior was responsible for conceptualization, funding acquisition, visualization, and writing - review & editing.

Alexandra Martins dos Santos Soares, was responsible for project administration, conceptualization, funding acquisition, supervision, visualization, and writing - review & editing.

Declaration of competing interest

The authors declare no conflict of interest related to this work.

Acknowledgments

The authors thank FINEP (Funding Authority for Studies and Projects) and FAPEMA (Maranhão Research Foundation) for supporting the IECT (Science and Technology Institute of Maranhão) Biotechnology. This study was financed in part by the Coordination for the Improvement of Higher Education Personnel (CAPES/Brazil) – Finance Code 001.

4. References

- Abbaszadeh, S., Sharifzadeh, A., Shokri, H., Khosravi, A.R., Abbaszadeh, A., 2014. Antifungal efficacy of thymol, carvacrol, eugenol and menthol as alternative agents to control the growth of food-relevant fungi. *J. Med. Mycol.* 24 (2), e51–e56.
- ANVISA, 2004. Guia de estabilidade de produtos cosméticos (Brasília, Agência Nacional de Vigilância Sanitária (ANVISA)).
- Araújo, L.X., Novato, T.P.L., Zeringota, V., Maturano, R., Melo, D., Da Silva, B., Daemon, E., De Carvalho, M.G., Monteiro, C.M.O., 2016. Synergism of thymol, carvacrol and eugenol in larvae of the cattle tick, *Rhipicephalus microplus*, and brown dog tick, *Rhipicephalus sanguineus*. *Med. Vet. Entomol.* 30, 377–382.
- Becker, S., Webster, A., Doyle, R.L., Martins, J.R., Reck, J., Klafke, G.M., 2019. Resistance to deltamethrin, fipronil and ivermectin in the brown dog tick, *Rhipicephalus sanguineus* sensu stricto, Latreille (Acari: ixodidae). *Ticks Tick Borne Dis* 10 (5), 1046–1050.
- Borges, L.M.F., Soares, S.F., Fonseca, I.N., Chaves, V.V., Louly, C.C.B., 2007. Resistência acaricida em larvas de *Rhipicephalus sanguineus* (Acari: ixodidae) de Goiania-GO. Brasil. *Rev. Patol. Trop.* 36 (1), 87–95.
- Coelho, L., Paula, L.G.F., Alves, S.G.A., Sampaio, A.L.N., Bezerra, G.P., Vilela, F.M.P., Matos, R.S., Zeringota, V., Borges, L.M.F., Monteiro, C., 2020. Combination of thymol and eugenol for the control of *Rhipicephalus sanguineus* sensu lato: evaluation of synergism on immature stages and formulation development. *Vet. Parasitol.* 277, 108989.
- Costa-Júnior, L.M., Miller, R.J., Alves, P.B., Blank, A.F., Li, A.Y., Pérez de León, A.A., 2016. Acaricidal efficacies of *Lippia gracilis* essential oil and its phytochemicals against organophosphate-resistant and susceptible strains of *Rhipicephalus (Boophilus) microplus*. *Vet. Parasitol.* 228, 60–64.

- Cruz, E.M.O., Costa-Junior, L.M., Pinto, J.A.O., Santos, D.A., de Araujo, S.A., Arrigoni-Blank, M.F., Bacci, L., Alves, P.B., Cavalcanti, S.C.H., Blank, A.F., 2013. Acaricidal activity of *Lippia gracilis* essential oil and its major constituents on the tick *Rhipicephalus (Boophilus) microplus*. *Vet. Parasitol.* 195, 198–202.
- Daemon, E., Monteiro, C.M.O., Maturano, R., Senra, T.O.S., Calmon, F., Faza, A., Prata, M.C.A., Georgopoulos, S.L., de Oliveira, L.F.C., 2012. Spectroscopic evaluation of thymol dissolved by different methods and influence on Acaricidal activity against larvae of *Rhipicephalus microplus* (Acari: iXodidae). *Parasitol. Res.* 111, 1901–1906.
- Dantas-Torres, F., 2008. The brown dog tick, *Rhipicephalus sanguineus* (Latreille, 1806) (Acari: iXodidae): from taxonomy to control. *Vet. Parasitol.* 152, 173–185.
- de Mendonça, A.E., Moreira, R.G., do Amaral, M.P.H., Monteiro, C.M.O., de Mello, V., Vilela, F.M.P., Mendonça Homem, F.C., Furlong, J., Dolinski, C., Prata, M.C.A., das Chagas, E.F., 2019. Entomopathogenic nematodes in pharmaceutical formulations for *Rhipicephalus microplus* (Acari: iXodidae) control: *in vitro* evaluation of compatibility, thermotolerance, and efficiency. *Ticks Tick Borne Dis* 10, 781–786.
- Delmonte, C., Cruz, P.B., Zeringóta, V., de Mello, V., Ferreira, F., Amaral, M.P.H., Daemon, E., 2017. Evaluation of the acaricidal activity of thymol incorporated in two formulations for topical use against immature stages of *Rhipicephalus sanguineus* sensu lato (Latreille, 1806) (Acari: iXodidae). *Parasitol. Res.* 116, 2957–2964.
- Díaz, E.L., Camberos, E.P., Herrera, G.A.C., Espinosa, M.E., Andrews, H.E., Buelnas, N.A.P., Ortega, A.G., Velázquez, M.M., 2019. Development of essential oil-based phytoformulations to control the cattle tick *Rhipicephalus microplus* using a miXture design approach. *EXp. Parasitol.* 201, 26–33.
- EAFUS, 2006. A Food Additive Database. Centre for Food Safety, and Applied Nutrition (US Food and Drug Administration, Washington, DC, USA).
- Ellse, L., Wall, R., 2013. The use of essential oils in veterinary ectoparasite control: a review. *Med. Vet. Entomol.* 28 (3), 233–243.
- European Parliament and Council, 1996. Regulation (EC) No 2232/96 the European parliament, and of the Council on 28 october 1996, commission decision of 23 february 1999 adopting a register of flavouring substances used in or on foodstuffs. *Off. J. Eur. Commun* 1–37. L84:1999/217/EC.
- Ferreira, F.M., Delmonte, C.C., Novato, T.L.P., Monteiro, C.M.O., Daemon, E., Vilela, F.M.P., Amaral, M.P.H., 2017. Acaricidal activity of essential oil of *Syzygium aromaticum*, hydrolate and eugenol formulated or free on larvae and engorged females of *Rhipicephalus microplus*. *Med. Vet. Entomol.* 32 (1), 41–47.
- Franc, M., Cadiergues, M.C., 1999. Activity of a deltamethrin shampoo Against *Ctenocephalides felis* and *Rhipicephalus sanguineus* in dogs. *Vet. Parasitol.* 81, 341–346.

- Hackman, R.H., Filshie, B.K., 1982. The tick Cuticle. In: Obenchain, F.D., Galun, R. (Eds.), *Physiology of Ticks*. Pergamon Press, Oxford, New York, Toronto, Sydney, Paris, Frankfurt, pp. 1–42.
- Heukelbach, J., Oliveira, F.A.S., Speare, R., 2006. A new shampoo based on nem (*Azadirachta indica*) is highly effective against head lice *in vitro*. *Parasitol. Res.* 99, 353–356.
- Hyldgaard, M., Mygind, T., Meyer, R.L., 2012. Essential oils in food preservation: mode of action, synergies, and interactions with food matriX components. *Front. Microbiol.* 3 (12).
- Klafke, G.M., Sabatini, G.A., de Albuquerque, T.A., Martins, J.R., Kemp, D.H., Miller, R. J., Schumaker, T.T., 2006. Larval immersion tests with ivermectin in populations of the cattle tick *Rhipicephalus (Boophilus) microplus* (Acari: iXodidae) from State of São Paulo, Brazil. *Vet. Parasitol.* 142, 386–390.
- Lees, A.D., 1947. Transpiration and the structure of the epicuticle in ticks. *J. Expt. Biol.* 23 (3–4), 379–410.
- Lima, A.S., Landulfo, G.A., Costa-Júnior, L.M., 2019. Repellent effects of encapsulated carvacrol on the *Rhipicephalus (Boophilus) microplus* (Acari: iXodidae). *J. Med. Entomol.* 56, 881–885.
- Lima, A.S., Maciel, A.P., Mendonça, C.J.S., Costa-Junior, L.M., 2017. Use of encapsulated carvacrol with yeast cell walls to control resistant strains of *Rhipicephalus microplus* (Acari: iXodidae). *Ind. Crop. Prod.* 108, 190–194.
- Novato, T., Gomes, G.A., Zeringóta, V., Franco, C.T., de Oliveira, D.R., Melo, D., de Carvalho, M.G., Daemon, E., de Oliveira Monteiro, C.M., 2018. *In vitro* assessment of the acaricidal activity of carvacrol, thymol, eugenol and their acetylated derivatives on *Rhipicephalus microplus* (Acari: iXodidae). *Vet. Parasitol.* 260, 1–4.
- Novato, T.L.P., Marchesini, P., Muniz, N., Prata, M.C.A., Furlong, J., Vilela, F.M.P., Daemon, E., Maturano, R., Monteiro, C., 2019. Evaluation of synergism and development of a formulation with thymol, carvacrol and eugenol for *Rhipicephalus microplus* control. *Expt. Parasitol.* 207, 107774.
- Novato, T.P.L., Araújo, L.X., de Monteiro, C.M.O., Maturano, R., Senra, T.O.S., Matos, R. S., Gomes, G.A., de Carvalho, M.G., Daemon, E., 2015. Evaluation of the combined effect of thymol, carvacrol and (E)-cinnamaldehyde on *Amblyomma sculptum* (Acari: iXodidae) and *Dermacentor nitens* (Acari: iXodidae) larvae. *Vet. Parasitol.* 212, 331–335.
- Park, J.H., Jeon, Y.J., Lee, C.H., Chung, N., Lee, H.S., 2017. Insecticidal toxicities of carvacrol and thymol derived from *Thymus vulgaris* Lin. against *Pochazia shantungensis* Chou & Lu., newly recorded pest. *Sci. Rep.* 7, 40902.
- Rodriguez-Vivas, R.I., Ojeda-Chi, M.M., Trinidad-Martinez, I., Perez de León, A.A., 2017. First documentation of ivermectin resistance in *Rhipicephalus sanguineus* sensu lato (Acari: iXodidae). *Vet. Parasitol.* 233, 9–13.

- Ryu, V., McClements, D.J., Corradini, M.G., Yang, J.S., McLandsborough, L., 2018. Natural antimicrobial delivery systems: formulation, antimicrobial activity, and mechanism of action of quillaja saponin-stabilized carvacrol nanoemulsions. *Food Hydrocolloids* 82, 442–450.
- Schuele, G., Barnett, S., Bapst, B., Cavaliero, T., Luempert, L., Strehlau, G., Young, D., Moran, C., Junquera, P., 2008. The effect of water and shampooing on the efficacy of a pyriproxyfen 12.5% topical solution against brown dog tick (*Rhipicephalus sanguineus*) and cat flea (*Ctenocephalides felis*) infestations on dogs. *Vet. Parasitol.* 151, 300–311.
- Scoralik, M.G., Daemon, E., Monteiro, C.M.O., Maturano, R., 2012. Enhancing the acaricide effect of thymol on larvae of the cattle tick *Rhipicephalus microplus* (Acari: ixodidae) by solubilization in ethanol. *Parasitol. Res.* 110, 645–648.
- Senra, T.O.S., Calmon, F., Zeringóta, V., Monteiro, C.M.O., Maturano, R., Matos, R.S., Melo, D., Gomes, G.A., de Carvalho, M.G., Daemon, E., 2013a. Investigation of activity of monoterpenes and phenylpropanoids against immature stages of *Amblyomma cajennense* and *Rhipicephalus sanguineus* (Acari: ixodidae). *Parasitol. Res.* 112, 3471–3476.
- Senra, T.O.S., Zeringóta, V., Monteiro, C.M.O., Calmon, F., Maturano, R., Gomes, G.A., Faza, A., de Carvalho, M.G., Daemon, E., 2013b. Assessment of the acaricidal activity of carvacrol, (E)-cinnamaldehyde, trans-anethole, and linalool on larvae of *Rhipicephalus microplus* and *Dermacentor nitens* (Acari: ixodidae). *Parasitol. Res.* 112, 1461–1466.
- Tabari, M.A., Youssefi, M.R., Maggi, F., Benelli, G., 2017. Toxic and repellent activity of selected monoterpenoids (thymol, carvacrol and linalool) against the castor bean tick, *Ixodes ricinus* (Acari: ixodidae). *Vet. Parasitol.* 245, 86–91.
- York, P., 2016. Delineamento de formas farmacêuticas. In: Aulton, M.E., Taylor, K.M.G. (Eds.), *Aulton - Delineamento de formas farmacêuticas*. Elsevier, Rio de Janeiro, pp. 17–28.

5. CONSIDERAÇÕES FINAIS

Concluímos que os óleos essenciais de diferentes cultivares de *Ocimum basilicum* apresentam potencial anti-helmíntico, sendo as cultivares que apresentam linalol e metil chavicol as mais promissoras, e também, concluímos que o shampoo contendo carvacrol apresenta eficácia acaricida.

6. REFERÊNCIAS GERAIS

- ZAKARIA, Z. et al. Antioxidant activity of *Coleus blumei*, *Orthosiphon stamineus*, *Ocimum basilicum* and *Mentha arvensis* from Lamiaceae family. *Int J Nat Eng Sci*, v. 2, n. 1, p. 93-95, 2008.
- ABBASZADEH, S.; SHARIFZADEH, A.; SHOKRI, H.; KHOSRAVI, A.R.; ABBASZADEH, A. Antifungal efficacy of thymol, carvacrol, eugenol and menthol as alternative agents to control the growth of food-relevant fungi. *Journal de mycologie medicale*, v. 24, n. 2, p. 51-56, 2014.
- ABOSHADY, H. M.; MANDONNET, N.; FÉLICITÉ, Y.; HIRA, J.; FOURCOT, A.; BARBIER, C.; BAMBOU, J. C. Dynamic transcriptomic changes of goat abomasal mucosa in response to *Haemonchus contortus* infection. *Veterinary research*, v. 51, n. 1, p. 1-12, 2020.
- ABOSSE, J. S.; TEREFE, G.; TESHALE, B. M. Comparative study on pathological changes in sheep and goats experimentally infected with *Haemonchus contortus*. *Surgical and Experimental Pathology*, v. 5, n. 1, p. 1-12, 2022.
- ABOU EL-SOUD, N. H.; DEABES, M.; ABOU EL-KASSEM, L.; KHALIL, M. Chemical composition and antifungal activity of *Ocimum basilicum* L. essential oil. *Open access Macedonian journal of medical sciences*, v. 3, n. 3, p. 374-379, 2015.
- ABOU EL-SOUD, N. H.; DEABES, M.; ABOU EL-KASSEM, L.; KHALIL, M. Chemical composition and antifungal activity of *Ocimum basilicum* L. essential oil. *Open access Macedonian journal of medical sciences*, v. 3, n. 3, p. 374, 2015.
- ABOSHADY, H. M., MANDONNET, N., FÉLICITÉ, Y., HIRA, J., FOURCOT, A., BARBIER, C., BAMBOU, J. C. Dynamic transcriptomic changes of goat abomasal mucosa in response to *Haemonchus contortus* infection. *Veterinary research*, v. 51, n. 1, p. 1-12, 2020.
- ABRAHAM, L. S.; MOREIRA, A. M.; MOURA, L. H.; DIAS, M. F. R. G.; ADDOR, F. A. S. A. Tratamentos estéticos e cuidados dos cabelos: uma visão médica (parte 2). *Surgical & Cosmetic Dermatology*, v. 1, n. 4, p. 178-185, 2009.
- ADAMS, H. R. *Farmacologia e terapêutica veterinária*. 8. ed. Rio de Janeiro: Guanabara Koogan, 2003.
- AESCHBACH, R.; LÖLIGER, J.; SCOTT, B. C.; MURCIA, A.; BUTLER, J.; HALLIWELL, B.; ARUOMA, O. I. Antioxidant actions of thymol, carvacrol, 6-gingerol, zingerone and hydroxytyrosol. *Food and Chemical Toxicology*, v. 32, n. 1, p. 31-36, 1994.
- ALAM, R. T.; HASSANEN, E. A.; EL-MANDRAWY, S. A. *Heamonchus contortus* infection in Sheep and Goats: alterations in haematological, biochemical, immunological, trace element and oxidative stress markers. *Journal of Applied Animal Research*, v. 48, n. 1, p. 357-364, 2020.

- ALBUQUERQUE, A. C. A.; BASSETTO, C. C.; ALMEIDA, F. A.; AMARANTE, A. F. Development of *Haemonchus contortus* resistance in sheep under suppressive or targeted selective treatment with monepantel. *Veterinary Parasitology*, v. 246, p. 112-117, 2017.
- ALMEIDA, I.; ALVIANO, D. S.; VIEIRA, D. P.; ALVES, P. B.; BLANK, A. F.; LOPES, A. H.; ROSA, M. D. S. S. Antigiardial activity of *Ocimum basilicum* essential oil. *Parasitology Research*, v. 101, n. 2, p. 443-452, 2007.
- ALMEIDA, N. E. C.; AGUIAR, I.; CARDOSO, D. R. Mechanism of hop-derived terpenes oxidation in beer. *Journal of the Brazilian Chemical Society*, v. 26, p. 2362-2368, 2015.
- ALMEIDA, R. R. **Mecanismos de ação dos monoterpenos aromáticos: timol e carvacrol.** 26 f. Moografia (Graduação em Química) - Universidade Federal de São João del-Rei, São João del-Rei, 2015.
- ALVES, K. C. S.; SANTOS, N. J. A.; MAIA, M. C. M.; OLIVEIRA, J. P. G.; VIEIRA, R. S. L.; MAIA, J. T. L. S. Métodos alternativos para o controle de carrapatos: uma análise bibliométrica. *Brazilian Journal of Development*, v. 7, n. 4, p. 37905-37920, 2021.
- AMARANTE, A. F. T.; BAGNOLA JR, J.; AMARANTE, M. R. V.; BARBOSA, M. A. Host specificity of sheep and cattle nematodes in São Paulo state, Brazil. *Veterinary parasitology*, v. 73, n. 1-2, p. 89-104, 1997.
- AMARANTE, A. F. T.; BRICARELLO, P. A.; HUNTLEY, J. F.; MAZZOLIN, L. P.; GOMES, J. C. Relationship of abomasal histology and parasite-specific immunoglobulin A with the resistance to *Haemonchus contortus* infection in three breeds of sheep. *Veterinary Parasitology*, v. 128, n. 1-2, p. 99-107, 2005.
- AMARANTE, A. F. T.; RAGOZO, Alessandra.; SILVA, Bruna Fernanda. *Os parasitas de ovinos*. São Paulo: Editora UNESP, 2014.
- ANDERSON, J. A.; COATS, J. R. Acetylcholinesterase inhibition by nootkatone and carvacrol in arthropods. *Pesticide Biochemistry and Physiology*, v. 102, n. 2, p. 124-128, 2012.
- ANDRÉ, W. P. P.; RIBEIRO, W. L. C.; OLIVEIRA, L. M. B.; MACEDO, I. T. F.; RONDON, F. C. M.; BEVILAQUA, C. M. L. Óleos essenciais e seus compostos bioativos no controle de nematóides gastrintestinais de pequenos ruminantes. *Acta Scientiae Veterinariae*, v. 46, p. 1-14, 2018.
- ANDRE, W. P.; RIBEIRO, W. L.; CAVALCANTE, G. S.; SANTOS, J. M.; MACEDO, I. T.; PAULA, H. C., BEVILAQUA, C. M. Comparative efficacy and toxic effects of carvacryl acetate and carvacrol on sheep gastrointestinal nematodes and mice. *Veterinary Parasitology*, v. 218, p. 52-58, 2016.
- ANDREOTTI, R. Situação atual da resistência do carrapato-do-boi (*Boophilus microplus*) aos acaricidas no Brasil. Campo Grande: Embrapa Gado de Corte, 2010.

ANVISA. *Agência Nacional de Vigilância Sanitária. Resolução nº 47, de 8 de setembro de 2009. Estabelece regras para elaboração, harmonização, atualização, publicação e disponibilização de bulas de medicamentos para pacientes e para profissionais de saúde.* Disponível em: https://bvsms.saude.gov.br/bvs/saudelegis/anvisa/2009/res0047_08_09_2009_rep.html. Acesso em: 10 jun. 2020.

ANVISA. *Guia de estabilidade de produtos cosméticos* (Brasília, Agência Nacional. 2004.

ARAÚJO, L. X.; NOVATO, T. P. L.; ZERINGOTA, V.; MATURANO, R.; MELO, D.; SILVA, B. C.; MONTEIRO, C. M. O. Synergism of thymol, carvacrol and eugenol in larvae of the cattle tick, *Rhipicephalus microplus*, and brown dog tick, *Rhipicephalus sanguineus*. *Medical and veterinary entomology*, v. 30, n. 4, p. 377-382, 2016.

ARENA, J. P.; LIU, K. K.; PARESS, P. S.; SCHAEFFER, J. M.; CULLY, D. F. Expression of a glutamate-activated chloride current in *Xenopus oocytes* injected with *Caenorhabditis elegans* RNA: evidence for modulation by avermectin. *Molecular Brain Research*, v. 15, n. 3-4, p. 339-348, 1992.

ATHANASIADOU, S.; KYRIAZAKIS, I. Plant secondary metabolites: antiparasitic effects and their role in ruminant production systems. *Proceedings of the Nutrition Society*, v. 63, n. 4, p. 631-639, 2004.

AZERÊDO, G. A. **Potencial de aplicação dos óleos essenciais de orégano (*Origanum vulgare* L.) e alecrim (*Rosmarinus officinalis* L.) como sanitizantes naturais em hortaliças minimamente processadas.** 2011. 135f. Tese (Doutorado) - Universidade Federal de Pernambuco, Recife, 2011.

BACKUS, L. H.; PÉREZ, A. M. L.; FOLEY, J. E. Effect of Temperature on Host Preference in Two Lineages of the Brown Dog Tick, *Rhipicephalus sanguineus*. *The American Journal of Tropical Medicine and Hygiene*, v. 104, n. 6, p. 2305-2311, 2021.

BAHIENSE, T. C.; FERNANDES, É. K.; ANGELO, I. D. C.; PERINOTTO, W. M. D. S., BITTENCOURT, V. R. Evaluation of the biological control potential of Metarhizium anisopliae toward *Boophilus microplus* in pen trials. *Revista Brasileira de Parasitologia Veterinária*, v. 16, n. 4, p. 243-245, 2007.

BAIRD, C.; CANN, M. *Química ambiental*. 4. ed. Porto Alegre: Bookman, 2011.

BAKKALI, F.; AVERBECK, S.; AVERBECK, D.; IDAOMAR, M. Biological effects of essential oils—a review. *Food and chemical toxicology*, v. 46, n. 2, p. 446-475, 2008.

BARROS-BATTESTI D. M.; ARZUA, M.; BECHARA, G. H. *Carrapatos de importância médico-veterinária da região neotropical: um guia ilustrado para identificação de espécies*. São Paulo, Vox/ICTTD-3/Butantan, 2006.

BEALL, M. J.; ALLEMAN, A. R.; BREITSCHWERDT, E. B.; COHN, L. A.; COUTO, C. G.; DRYDEN, M. W.; YABSLEY, M. J. Seroprevalence of *Ehrlichia canis*,

Ehrlichia chaffeensis and *Ehrlichia ewingii* in dogs in North America. *Parasites & vectors*, v. 5, n. 1, p. 1-11, 2012.

BEATI, L.; KEIRANS, J. E. Analysis of the systematic relationships among ticks of the genera *Rhipicephalus* and *Boophilus* (Acari: Ixodidae) based on mitochondrial 12S ribosomal DNA gene sequences and morphological characters. *Journal of Parasitology*, v. 87, n. 1, p. 32-48, 2001.

BECHARA, G. H.; MORELLI, J.; SZABÓ, M. P. J. Skin test and tick immune status in susceptible and resistant cattle in Brazil. *Annals-New York Academy of Sciences*, v. 916, p. 570-575, 2000.

BECKER, S.; WEBSTER, A.; DOYLE, R.L.; MARTINS, J.R.; RECK, J.; KLAFFE, G.M. Resistance to deltamethrin, fipronil and ivermectin in the brown dog tick, *Rhipicephalus sanguineus* sensu stricto, Latreille (Acari: Ixodidae). *Ticks and tick-borne diseases*, v. 10, n. 5, p. 1046-1050, 2019.

BEN ARFA, A.; PREZIOSI-BELLOY, L.; CHALIER, P.; GONTARD, N. Antimicrobial paper based on a soy protein isolate or modified starch coating including carvacrol and cinnamaldehyde. *Journal of agricultural and food chemistry*, v. 55, n. 6, p. 2155-2162, 2007.

BEN-ERIK, V. W.; WINK, M. *Medicinal plants of the world*. Pretoria: Briza Publications, 2017.

BESIER, R. B.; KAHN, L. P.; SARGISON, N. D.; VAN WYK, J. A. The pathophysiology, ecology and epidemiology of *Haemonchus contortus* infection in small ruminants. *Advances in parasitology*, v. 93, p. 95-143, 2016.

BISSINGER, B. W.; ROE, R. M. Tick repellents: past, present, and future. *Pesticide biochemistry and physiology*, v. 96, n. 2, p. 63-79, 2010.

BLANK, A. F.; SOUZA, E. M. D.; ARRIGONI-BLANK, M. D. F.; PAULA, J. W. A. D.; ALVES, P. B. Maria Bonita: a linalool type basil cultivar. *Pesquisa Agropecuaria Brasileira*, v. 42, p. 1811-1813, 2007.

BLENAU, W.; RADEMACHER, E.; BAUMANN, A. Plant essential oils and formamidines as insecticides/acaricides: what are the molecular targets?. *Apidologie*, v. 43, n. 3, p. 334-347, 2012.

BOGNI, S.; CHAGAS, A. C. S.; SILVA, I. C.; BOSCHINI, L.; VIEIRA, L. C. C.; CORRÊA, A. G. Ação acaricida de terpenos sobre o carrapato bovino *Rhipicephalus* (*Boophilus*) *microplus*. In: Anais XVI Congresso Brasileiro de Parasitologia Veterinária, Campo Grande, Mato Grosso do Sul, p 16, 2010.

BOMBARDI, L. M. *Geografia do uso de agrotóxicos no Brasil e conexões com a União Europeia*. São Paulo: FFLCH-USP, 2017.

BORGES, L.M.F.; SOARES, S.F.; FONSECA, I.N.; CHAVES, V.V.; LOULY, C.C.B. Resistência acaricida em larvas de *Rhipicephalus sanguineus* (Acari: Ixodidae) de

Goiânia-GO. Brasil. *Rev. Patol. Trop.*, v. 36, n. 1, p. 87-95, 2007.

BORGES, L.M.F.; SOUSA, L.A.D.; BARBOSA, C. S. Perspectives for the useof plant extracts to control the cattle tick *Rhipicephalus (Boophilus) microplus*. *Rev. Bras. Parasitol. Vet.*, v. 20, p. 89–96, 2011.

BORTOLUZZI, B. B.; BUZATTI, A.; DESCHAMPS, C.; BERTOLDI, F. C.; CHaabán, A.; PERRUCCI, S.; MOLENTO, M. B. Fitoterapia no controle de parasitos gastrintestinais de ruminantes: ênfase no gênero *Mentha* e seus componentes bioativos. *Ars Veterinaria*, v. 36, n. 4, p. 253-270, 2020.

BRASIL. Ministério da Agricultura, Pecuária e Abastecimento. Portaria nº 56, de 07/12/99. Diário Oficial da União, Brasília, 08 dez. 1999. Seção I, p. 34-45.

BRITES-NETO, J.; DUARTE, K. M. R.; MARTINS, T. F. Tick-borne infections in human and animal population worldwide. *Veterinary world*, v. 8, n. 3, p. 301, 2015.

BRITO, D. R. B.; SANTOS, A.; GUERRA, C. G. V. Ectoparasitos em rebanhos de caprinos e ovinos na microrregião do Alto Mearim e Grajau, Estado do Maranhão. *Brasil J Vet Parasitol*, v. 14, p. 59-63, 2005.

BRITO, L. G.; NETTO, F. G. S.; OLIVEIRA, M. C. S.; BARBIERI, F. S. Bio-ecologia, importância medico veterinária e controle do carrapato, com ênfase no carrapato dos bovinos, *Rhipicephalus (Boophilus) microplus*. *Embrapa Rondônia-Documentos (INFOTECA-E)*, p. 21. 2006.

BROGLIO-MICHELETTI, S. M. F.; VALENTE, E. C. N.; SOUZA, L. A. D.; DIAS, N. D. S.; ARAÚJO, A. M. N. D. Extratos de plantas no controle de *Rhipicephalus (Boophilus) microplus* (Canestrini, 1887) (Acari: Ixodidae) em laboratório. *Revista Brasileira de Parasitologia Veterinária*, v. 18, p. 44-48, 2009.

BRUIN, W.; VAN DER MERWE, C.; KRITZINGER, Q.; BORNMAN, R.; KORSTEN, L. Ultrastructural and developmental evidence of phytotoxicity on cos lettuce (*Lactuca sativa*) associated with nonylphenol exposure. *Chemosphere*, v. 169, p. 428-436, 2017.

BURG, R. W.; STAPLEY, E. O. Isolation and characterization of the producing organism. In: CAMPBELL, W. C., editor. *Ivermectin and abamectin*. New York: Springer; 1989.

BURKE, J. M.; MILLER, J. E. Evaluation of multiple low doses of copper oxide wire particles compared with levamisole for control of *Haemonchus contortus* in lambs. *Veterinary parasitology*, v. 139, n. 1-3, p. 145-149, 2006.

BURT, S. A.; FLEDDERMAN, M. J.; HAAGSMAN, H. P.; VAN KNAPEN, F.; VELDHUIZEN, E. J. Inhibition of *Salmonella enterica* serotype Enteritidis on agar and raw chicken by carvacrol vapour. *International Journal of Food Microbiology*, v. 119, n. 3, p. 346-350, 2007.

CAMICAS, J. L.; HERVY, J. P.; ADAM, F.; MOREL, P. C. The ticks of the world (Acarida, Ixodida): nomenclature, described stages, hosts, distribution. France: Orstom Editions, 1998.

CAMILLO, G.; VOGEL, F. F.; SANGIONI, L. A.; CADORE, G. C.; FERRARI, R. Eficiência *in vitro* de acaricidas sobre carapatos de bovinos no Estado do Rio Grande do Sul, Brasil. *Ciência Rural*, v. 39, p. 490-495, 2009.

CAMILO, C. J.; ALVES NONATO, C. D. F.; GALVÃO-RODRIGUES, F. F.; COSTA, W. D.; CLEMENTE, G. G.; SOBREIRA MACEDO, M. A. C.; COSTA, J. G. M. Acaricidal activity of essential oils: a review. *Trends in Phytochemical Research*, v. 1, n. 4, p. 183-198, 2017.

CAMPOS, R. N. S.; BACCI, L.; ARAÚJO, A. P. A.; BLANK, A. F.; ARRIGONI-BLANK, M. F.; SANTOS, G. R. A.; RONER, M. N. Essential oils of medicinal and aromatic plants in the control of tick *Rhipicephalus microplus*. *Archivos de Zootecnia*, v. 61, n. R, p. 67-78, 2012.

CARDOSO, A.S. ; SANTOS, E. G. G.; SILVA LIMA, A.; TEMEYER, K. B.; LEON, A. A. P.; JUNIOR, L. M. C.; SANTOS SOARES, A. M. Terpenes on *Rhipicephalus (Boophilus) microplus*: Acaricidal activity and acetylcholinesterase inhibition. *Veterinary parasitology*, v. 280, p. 1-5, 2020.

CARVALHO, A. C. B. **Plantas medicinais e fitoterápicos: regulamentação sanitária e proposta de modelo de monografia para espécies vegetais oficializadas no Brasil.** 2011. 318 f. Tese (Doutorado em Ciências da Saúde) -Universidade de Brasília, Brasília, 2011.

CARVALHO, C. O.; CHAGAS, A. C. S.; COTINGUIBA, F.; FURLAN, M.; BRITO, L. G.; CHAVES, F. C.; AMARANTE, A. F. The anthelmintic effect of plant extracts on *Haemonchus contortus* and *Strongyloides venezuelensis*. *Veterinary parasitology*, v. 183, n. 3-4, p. 260-268, 2012.

CASTILHO, C. V.; FANTATTO, R. R.; GAÍNZA, Y. A.; BIZZO, H. R.; BARBI, N. S.; LEITÃO, S. G.; CHAGAS, A. C. S. *In vitro* activity of the essential oil from *Hesperozygis myrtoides* on *Rhipicephalus (Boophilus) microplus* and *Haemonchus contortus*. *Revista Brasileira de Farmacognosia*, v. 27, p. 70-76, 2017.

CASTRO, K. N. D. C.; CHAGAS, A. C. D. S.; COSTA-JÚNIOR, L. M.; CANUTO, K. M.; BRITO, E. S. D.; RODRIGUES, T. H. S.; ANDRADE, I. M. Acaricidal potential of volatile oils from *Croton species* on *Rhipicephalus microplus*. *Revista Brasileira de Farmacognosia*, v. 29, p. 811-815, 2020.

CASTRO, L. M.; PINTO, N. B.; CASTRO, L. L. D.; MOURA, M. Q.; MOTA, T. O.; MADRID, I. M.; BERNE, M. E. A. Atividade ovicida do óleo essencial e do extrato hidroalcoólico de *Ocimum basilicum* sobre nematódeos gastrintestinais de ovinos. *Science and Animal Health*, v. 5, n. 2, p. 138-150, 2017.

ÇETIN, H.; CILEK, J. E.; OZ, E.; AYDIN, L. E. V. E. N. T.; DEVECI, O.; YANIKOGLU, A. Acaricidal activity of *Satureja thymbra* L. essential oil and its major

components, carvacrol and γ -terpinene against adult *Hyalomma marginatum* (Acari: Ixodidae). *Veterinary Parasitology*, v. 170, n. 3-4, p. 287-290, 2010.

CEZAR, A. S.; CATTO, J. B.; BIANCHIN, I. Controle alternativo de nematoides gastrintestinais dos ruminantes: atualidade e perspectivas. *Ciência Rural*, v. 38, n.7, p. 2083-2091, 2008.

CHAGAS, A. C.S.; BARROS, L. D.; COTINGUIBA, F.; FURLAN, M.; GIGLIOTTI, R.; SENA OLIVEIRA, M. C.; BIZZO, H. R. *In vitro* efficacy of plant extracts and synthesized substances on *Rhipicephalus (Boophilus) microplus* (Acari: Ixodidae). *Parasitology Research*, v. 110, n. 1, p. 295-303, 2012.

CHEESEMAN, C. L.; DELANY, N. S., WOODS, D. J.; WOLSTENHOLME, A. J. High-affinity ivermectin binding to recombinant subunits of the *Haemonchus contortus* glutamate-gated chloride channel. *Mol Biochem Parasitol*, v. 114, n. 2, p. 161-168, 2001.

Coelho, L.; Paula, L.G.F.; Alves, S.G.A.; Sampaio, A.L.N.; Bezerra, G.P.; Vilela, F.M.P.; Matos, R.S.; Zeringota, V.; Borges, L.M.F.; Monteiro, C. Combination of thymol and eugenol for the control of *Rhipicephalus sanguineus* sensu lato: evaluation of synergism on immature stages and formulation development. *Vet. Parasitol*, v. 277, p. 1-37, 2020.

COLES, G. C.; BAUER, C.; BORGSTEED, F. H. M.; GEERTS, S.; KLEI, T. R.; TAYLOR, M. A.; WALLER, P. J. World Association for the Advancement of Veterinary Parasitology (WAAVP) methods for the detection of anthelmintic resistance in nematodes of veterinary importance. *Veterinary parasitology*, v. 44, n. 1-2, p. 35-44, 1992.

CORREIA, S. D. J.; DAVIDI, J. M.; SILVA, E. P. D.; DAVID, J. P.; LOPES, L. M.; GUEDES, M. L. S. Flavonóides, norisoprenóides e outros terpenos das Folhas de *Tapirira guianensis*. *Química Nova*. v. 31, n. 8, p. 2056-2059, 2008.

COSTA, C. T. C.; BEVILAQUA, C. M. L.; CAMURÇA-VASCONCELOS, A. L. F.; MACIEL, M. V.; MORAIS, S. M.; CASTRO, C. M. S.; OLIVEIRA, L. M. B. *In vitro* ovicidal and larvicidal activity of *Azadirachta indica* extracts on *Haemonchus contortus*. *Small Ruminant Research*, v. 74, n. 1-3, p. 284-287, 2008.

COSTA, V. M. M.; SIMÕES, S. V. D.; RIET-CORREA, F. Doenças parasitárias em ruminantes no semiárido brasileiro. *Pesquisa Veterinária Brasileira*, v. 29, p. 563-568, 2009.

COSTA-JÚNIOR, L. M.; MILLER, R. J.; ALVES, P. B.; BLANK, A. F.; LI, A. Y.; LEON, A. A. P. Acaricidal efficacies of *Lippia gracilis* essential oil and its phytochemicals against organophosphate-resistant and susceptible strains of *Rhipicephalus (Boophilus) microplus*. *Veterinary parasitology*, v. 228, p. 60-64, 2016.

COSTA-JUNIOR, L. M.; SILVA, C. R.; SOARES, A. M.; MENEZES, A. S.; SILVA, M. R.; AMARANTE, A. F.; ALENCAR, L. M. Assessment of biophysical properties of

Haemonchus contortus from different life cycle stages with atomic force microscopy. *Ultramicroscopy*, v. 209, p. 1-21, 2020.

COX-GEORGIAN, D.; RAMADOUSS, N.; DONA, C.; BASU, C. Therapeutic and medicinal uses of terpenes. In: *Medicinal Plants*. Springer, Cham, p. 333-359, 2019.

CRUZ, E. M. O. **Época de colheita, irrigação, fitoquímica e atividades carrapaticida e fungicida do óleo essencial de genótipos de Lippia gracilis Schauer.** 2013. 77 f. Tese (Doutorado em Biotecnologia) – Rede Nordeste de Biotecnologia – RENORBIO, Universidade Federal de Sergipe. São Cristovão, 2013.

CRUZ, E. M. O.; COSTA-JUNIOR, L. M.; PINTO, J. A. O.; ALEXANDRIA SANTOS, D.; ARAUJO, S. A.; FÁTIMA ARRIGONI-BLANK, M.; BLANK, A. F. Acaricidal activity of *Lippia gracilis* essential oil and its major constituents on the tick *Rhipicephalus (Boophilus) microplus*. *Veterinary Parasitology*, v. 195, n. 1-2, p. 198-202, 2013.

DAEMON, E.; MONTEIRO, C.M.O.; MATURANO, R.; SENRA, T.O.S.; CALMON, F.; FAZA, A.; PRATA, M.C.A.; GEORGOPoulos, S.L.; OLIVEIRA, L.F.C. Spectroscopic evaluation of thymol dissolved by different methods and influence on acaricidal activity against larvae of *Rhipicephalus microplus* (Acari: Ixodidae). *Parasitology research*, v. 111, n. 5, p. 1901-1906, 2012.

DAMASCENO, S. R.; OLIVEIRA, F. R. A.; CARVALHO, N. S.; BRITO, C. F.; SILVA, I. S.; SOUSA, F. B. M.; MEDEIROS, J. V. R. Carvacryl acetate, a derivative of carvacrol, reduces nociceptive and inflammatory response in mice. *Life Sciences*, v. 94, n. 1, p. 58-66, 2014.

DANTAS-TORRES, F. Biology and ecology of the brown dog tick, *Rhipicephalus sanguineus*. *Parasites & vectors*, v. 3, n. 1, p. 1-11, 2010.

DANTAS-TORRES, F. The brown dog tick, *Rhipicephalus sanguineus* (Latreille, 1806) (Acari: Ixodidae): from taxonomy to control. *Vet Parasitol.*, v. 152, p. 173-185, 2008.

DANTAS-TORRES, F; FIGUEREDO, L. A.; BRANDÃO-FILHO, S. P. *Rhipicephalus sanguineus* (Acari: Ixodidae), the brown dog tick, parasitizing humans in Brazil. *Revista da Sociedade Brasileira de Medicina Tropical*, v. 39, p. 64-67, 2006.

DAVEY, R. B.; MILLER, R. J.; GEORGE, J. E. Efficacy of amitraz applied as a dip against an amitraz-resistant strain of *Rhipicephalus (Boophilus) microplus* (Acari: Ixodidae) infested on cattle. *Veterinary Parasitology*, v. 152, n. 1-2, p. 127-135, 2008.

DELMONTE, C.; CRUZ, P.B.; ZERINGO'TA, V.; MELLO, V.; FERREIRA, F.; AMARAL, M.P.H.; DAEMON, E. Evaluation of the acaricidal activity of thymol incorporated in two formulations for topical use against immature stages of *Rhipicephalus sanguineus* sensu lato (Latreille, 1806) (Acari: Ixodidae). *Parasitology research*, v. 116, n. 11, p. 2957-2964, 2017.

DEWICK, Paul M. *Medicinal natural products: a biosynthetic approach*. Toronto, Ontario: John Wiley & Sons, 2002.

DIAS, M. F. R. G. Hair cosmetics: an overview. *International Journal of Trichology*, v. 7, n. 1, p. 2-15, 2015.

DÍAZ, E.L.; CAMBEROS, E.P.; HERRERA, G.A.C.; ESPINOSA, M.E.; ANDREWS, H.E.; BUELNAS, N.A. P.; ORTEGA, A.G.; VELA ZQUEZ, M.M. Development of essential oil-based phyto-formulations to control the cattle tick *Rhipicephalus microplus* using a mixture design approach. *Experimental parasitology*, v. 201, p. 26-33, 2019.

DIETRICH, M.; GOMEZ-DIAZ, E.; MCCOY, K. D. Worldwide distribution and diversity of seabird ticks: implications for the ecology and epidemiology of tick-borne pathogens. *Vector-Borne and Zoonotic Diseases*, v. 11, n. 5, p. 453-470, 2011.

DOLAN, M. C.; JORDAN, R. A.; SCHULZE, T. L.; SCHULZE, C. J.; CORNELL MANNING, M.; RUFFOLO, D.; KARCHESY, J. J. Ability of two natural products, nootkatone and carvacrol, to suppress *Ixodes scapularis* and *Amblyomma americanum* (Acari: Ixodidae) in a Lyme disease endemic area of New Jersey. *Journal of economic entomology*, v. 102, n. 6, p. 2316-2324, 2009.

DOMINGUES, R.; WOHLRES-VIANA, S.; REIS, D. R. L.; TEIXEIRA, H. C.; FERREIRA, A. P.; GUIMARÃES, S. E. F.; MACHADO, M. A. Expression of immune response genes in peripheral blood of cattle infested with *Rhipicephalus microplus*. *Genet Mol Res*, v. 13, n. 2, p. 4013-4021. 2014.

DRAELOS, Z. D.; MURRELL, D. F.; HUGHES, M. H.; ZANE, L. T. Post Hoc Analyses of the Effect of Crisaborole Topical Ointment, 2% on Atopic Dermatitis: Associated Pruritus from Phase 1 and 2 Clinical Studies. *Journal of drugs in dermatology: JDD*, v. 15, n. 2, p. 172-176, 2016.

DUBEY, V. S.; BHALLA, R.; LUTHRA, R. An overview of the non-mevalonate pathway for terpenoid biosynthesis in plants. *J. Biosci*, v. 28, n. 5, p. 637-646, 2003.

EAFUS. *A Food Additive Database. Centre for Food Safety, and Applied Nutrition (US Food and Drug Administration*, Washington, DC, USA, 2006.

ECHEVARRIA, F.; BORBA, M. F. S.; PINHEIRO, A. C.; WALLER, P. J.; HANSEN, J. W. The prevalence of anthelmintic resistance in nematode parasites of sheep in Southern Latin America: Brazil. *Veterinary Parasitology*, v. 62, n. 3-4, p. 199-206, 1996.

EHSAN, M.; HU, R. S.; LIANG, Q. L.; HOU, J. L.; SONG, X.; YAN, R.; LI, X. Advances in the development of anti-*Haemonchus contortus* vaccines: challenges, opportunities, and perspectives. *Vaccines*, v. 8, n. 3, p. 1-18, 2020.

EL-ASHRAM, S.; AL NASR, I.; MEHMOOD, R.; HU, M.; HE, L.; SUO, X. *Haemonchus contortus* and ovine host: a retrospective review. *Int J Adv Res*, v. 5, p. 972-999, 2017.

ELLSE, L.; WALL, R. The use of essential oils in veterinary ectoparasite control: a review. *Medical and Veterinary Entomology*, v. 28, n. 3, p. 233-243, 2014.

EMERY, D. L.; HUNT, P. W.; LE JAMBRE, L. F. *Haemonchus contortus*: the then and now, and where to from here?. *International Journal for Parasitology*, v. 46, n. 12, p. 755-769, 2016.

ENCINOSA GUZMAN, P. E.; BELLO SOTO, Y.; RODRÍGUEZ-MALLON, A. Genetic and biological characterization of a Cuban tick strain from *Rhipicephalus sanguineus* complex and its sensitivity to different chemical acaricides. *International Journal of Acarology*, v. 42, n. 1, p. 18-25, 2016.

ENGSTRÖM, M. *Understanding the bioactivity of plant tannins: developments in analysis methods and structure-activity studies*. 2016.

ESCÁMEZ, J. C.; RUBÍ, J. C. M.; RODRIGUEZ; F. Y. Intoxicación por Organoclorados, carbamatos y herbicidas. In: CEBRIÁN, J. G. et al. (eds.). *Principios de Urgencias, Emergencias y Cuidados Críticos*. 1996. Disponível em: <http://umeet.uninet.edu/tratado/c1006i.html>. Acesso em: 20 abr. 2021.

European Parliament and Council, 1996. *Regulation (EC) No 2232/96 the European parliament, and of the Council on 28 october 1996, commission decision of 23 february 1999 adopting a register of flavouring substances used in or on foodstuffs*. Off. J. Eur. Commun 1–37. L84:1999/217/EC.

FAJIMI, A. K.; TAIWO, A. A. Herbal remedies in animal parasitic diseases in Nigeria: a review. *African journal of biotechnology*, v. 4, n. 4, p. 303-307, 2005.

FAO. Resistance management and integrated parasite control in ruminants, Guidelines. Rome, *Animal Production and Health Division*, pp. 78–118, 2004.

FARIAS, M. P. O.; SOUSA, D. P. D.; ARRUDA, A. C.; WANDERLEY, A. G.; TEIXEIRA, W. C.; ALVES, L. C.; FAUSTINO, M. A. D. G. Potencial acaricida do óleo de andiroba *Carapa guianensis* Aubl. sobre fêmeas adultas ingurgitadas de *Anocentor nitens* Neumann, 1897 e *Rhipicephalus sanguineus* Latreille, 1806. *Arq. Bras. Med. Zootec.*, v. 61, n. 4, p. 877-882, 2009.

FARIAS, N. A. R.; CUNHA FILHO, N. A.; VAZ JUNIOR, I. S. Uso de acaricidas em *Rhipicephalus (B.) microplus* de duas regiões fisiográficas do Rio Grande do Sul. *Acta scientiae veterinariae*, v. 36, n. 1, p. 25-30, 2008.

FELIPE, L. O.; BICAS, J. L. Terpenos, aromas e a química dos compostos naturais. *Química Nova na Escola*, v. 39, n. 2, p. 120-130, 2017.

FERREIRA, F. M.; DELMONTE, C. C.; NOVATO, T. L. P.; MONTEIRO, C. M. O.; DAEMON, E.; VILELA, F. M. P.; AMARAL, M. P. H. Acaricidal activity of essential oil of *Syzygium aromaticum*, hydrolate and eugenol formulated or free on larvae and engorged females of *Rhipicephalus microplus*. *Medical and veterinary entomology*, v. 32, n. 1, p. 41-47, 2018.

FERREIRA, F.M.; DELMONTE, C.C.; NOVATO, T.L.P.; MONTEIRO, C.M.O.; DAEMON, E.; VILELA, F. M.P.; AMARAL, M.P.H. Acaricidal activity of essential oil of *Syzygium aromaticum*, hydrolate and eugenol formulated or free on larvae and

engorged females of *Rhipicephalus microplus*. *Medical and veterinary entomology*, v. 32, n. 1, p. 41-47, 2018.

FERREIRA, L. E.; BENINCASA, B. I.; FACHIN, A. L.; CONTINI, S. H. T.; FRANÇA, S. C.; CHAGAS, A. C. S.; BELEBONI, R. O. Essential oils of *Citrus aurantifolia*, *Anthemis nobile* and *Lavandula officinalis*: *in vitro* anthelmintic activities against *Haemonchus contortus*. *Parasites & vectors*, v. 11, n. 1, p. 1-9, 2018.

FERREIRA, L. E.; BENINCASA, B. I.; FACHIN, A. L.; FRANCA, S. C.; CONTINI, S. S.; CHAGAS, A. C.; BELEBONI, R. O. *Thymus vulgaris* L. essential oil and its main component thymol: Anthelmintic effects against *Haemonchus contortus* from sheep. *Veterinary Parasitology*, v. 228, p. 70-76, 2016.

FERREIRA, T. P. *In vitro* Acaricidal Activity of *Ocimum gratissimum* Essential Oil on *Rhipicephalus sanguineus*, *Amblyomma sculptum* and *Rhipicephalus microplus* Larvae. *Revista Virtual de Química*, v. 11 n. 5, p. 1604-1613, 2019.

FRAGA, A. B.; ALENCAR, M. M. D.; FIGUEIREDO, L. A. D.; RAZOOK, A. G.; CYRILLO, J. N. D. S. G. Análise de fatores genéticos e ambientais que afetam a infestação de fêmeas bovinas da raça Caracu por carrapatos (*Boophilus microplus*). *Revista Brasileira de Zootecnia*, v. 32, p. 1578-1586, 2003.

Franc, M.; Cadiergues, M.C. 1999. Activity of a deltamethrin shampoo against *Ctenocephalides felis* and *Rhipicephalus sanguineus* in dogs. *Veterinary parasitology*, v. 81, n. 4, p. 341-346, 1999.

FREITAS FERNANDES, F.; FREITAS, E. P. S. Acaricidal activity of an oleoresinous extract from *Copaifera reticulata* (Leguminosae: Caesalpinoideae) against larvae of the southern cattle tick, *Rhipicephalus (Boophilus) microplus* (Acari: Ixodidae). *Veterinary parasitology*, v. 147, n. 1-2, p. 150-154, 2007.

FURLONG, J. *Carrapato: problemas e soluções*. Juiz de Fora: Embrapa Gado de Leite, 2005.

FURLONG, J. Controle do carrapato dos bovinos na Região Sudeste do Brasil. *Cadernos Técnicos da Escola de Veterinária da UFMG*, v. 8, p. 46-61, 1993.

FURLONG, J.; MARTINS, J. R. Resistência a carrapaticidas. *Circular Técnica – CNPGL*, n. 59, p. 1-25, 2000.

FURLONG, J.; SALES, R. O. Controle estratégico de carrapatos no bovino de leite: uma revisão. *Rev. Brás. Hig. San. Anim.*, v. 1. n. 2, p. 44-72, 2007.

FUTSE, J. E.; UETI, M. W.; KNOWLES JR, D. P.; PALMER, G. H. Transmissão de *Anaplasma marginale* por *Boophilus microplus*: retenção da competência do vetor na ausência de interação vetor-patógeno. *J Clin Microbiol*, v. 41, p. 3829-3834, 2003.

GALBREATH, K. E.; HOBERG, E. P.; COOK, J. A.; ARMIÉN, B.; BELL, K. C.; CAMPBELL, M. L.; HOPE, A. G. Building an integrated infrastructure for exploring

biodiversity: field collections and archives of mammals and parasites. *Journal of Mammalogy*, v. 100, n. 2, p. 382-393, 2019.

GARCIA, A. A.; CARRIL, E. P. U. Metabolismo secundário de plantas. *Reduca (biología)*, v. 2, n. 3, p. 1-5, 2011.

GARCIA, M. V.; MONTEIRO, A. C.; SZABÓ, M. P. J.; PRETTE, N. Eventos externos e internos da infecção de larvas e ninhas de *Rhipicephalus sanguineus* por *Metarhizium anisopliae*. *Arquivo Brasileiro de Medicina Veterinária e Zootecnia*, v. 60, p. 855-863, 2008.

GARCIA-BUSTOS, J. F.; SLEEB, B. E.; GASSER, R. B. An appraisal of natural products active against parasitic nematodes of animals. *Parasites & Vectors*, v. 12, n. 1, p. 1-22, 2019.

GEARY, T. G.; SIMS, S. M.; THOMAS, E. M.; VANOVER, L.; DAVIS, J. P.; WINTERROWD, C. A.; THOMPSON, D. P. *Haemonchus contortus*: ivermectin-induced paralysis of the pharynx. *Experimental parasitology*, v. 77, n. 1, p. 88-96, 1993.

GERSHENZON, J.; DUDAREVA, N. The function of terpene natural products in the natural world. *Nature chemical biology*, v. 3, n. 7, p. 408-414, 2007.

GIACHINO, R.; SÖNMEZ, Ç.; TONK, F. A.; BAYRAM, E.; YÜCE, S.; TELCI, I.; FURAN, M. A. RAPD and essential oil characterization of *Turkish basil (Ocimum basilicum L.)*. *Plant Systematics and Evolution*, v. 300, n. 8, p. 1779-1791, 2014.

GIGLIOTTI, R.; OLIVEIRA, H. N.; BILHASSI, T. B.; PORTILHO, A. I.; OKINO, C. H.; MARCONDES, C. R.; SENA OLIVEIRA, M. C. Estimates of repeatability and correlations of hemoparasites infection levels for cattle reared in endemic areas for *Rhipicephalus microplus*. *Veterinary parasitology*, v. 250, p. 78-84, 2018.

GILLEARD, J. S. *Haemonchus contortus* as a paradigm and model to study anthelmintic drug resistance. *Parasitology*, v. 140, n. 12, p. 1506-1522, 2013.

GINNSBERG, H. S.; GLAZER, I. *Anti-tick biological control agents: assessment and future perspectives*. Cambridge, England: Cambridge University Press, p. 447-469, 2008.

GODARA, R.; KATOCH, R.; RAFIQI, S. I.; YADAV, A.; NAZIM, K.; SHARMA, R.; KATOCH, M. Synthetic pyrethroid resistance in *Rhipicephalus (Boophilus) microplus* ticks from north-western Himalayas, India. *Tropical animal health and production*, v. 51, n. 5, p. 1203-1208, 2019.

GODDARD, J. Focus of human parasitism by the brown dog tick, *Rhipicephalus sanguineus* (Acari: Ixodidae). *Journal of medical entomology*, v. 26, n. 6, p. 628-631, 1989.

GODOI, C. R.; SILVA, E. F. P. Tick *Boophilus microplus* and impact on animal production-literature review. *PUBVET*, v. 3, n. 22, 2009.

GOMES, A. Berne: um pequeno parasito, porem um grande problema. *Embrapa Gado de Corte Divulga*, n. 27, p. 1-5, 1998.

GONZALES, J. C. *O controle do carrapato bovino*. Porto Alegre: Editora Sulina, 1975.

GOVINDARAJAN, M.; SIVAKUMAR, R. Mosquito adulticidal and repellent activities of botanical extracts against malarial vector, *Anopheles stephensi* Liston (Diptera: Culicidae). *Asian Pac. J. Trop. Med*, v. 4, p. 941–947, 2011.

GOVINDARAJAN, M.; SIVAKUMAR, R.; RAJESWARY, M.; YOGALAKSHMI, K. Chemical composition and larvicidal activity of essential oil from *Ocimum basilicum* (L.) against *Culex tritaeniorhynchus*, *Aedes albopictus* and *Anopheles subpictus* (Diptera: Culicidae). *Experimental parasitology*, v. 134, n. 1, p. 7-11, 2013.

GRAY, J.; DANTAS-TORRES, F.; ESTRADA-PEÑA, A.; LEVIN, M. Systematics and ecology of the brown dog tick, *Rhipicephalus sanguineus*. *Ticks and tick-borne diseases*, v. 4, n. 3, p. 171-180, 2013.

GRISI, L.; LEITE, R. C.; MARTINS, J. R. D. S.; BARROS, A. T. M. D.; ANDREOTTI, R.; CANÇADO, P. H. D.; VILLELA, H. S. Reavaliação do impacto econômico potencial de parasitas bovinos no Brasil. *Rev Bras Parasitol Vet*, v. 23, p. 150-156, 2014.

GROSS, A. D.; TEMEYER, K. B.; DAY, T. A.; DE LEÓN, A. A. P.; KIMBER, M. J.; COATS, J. R. Interaction of plant essential oil terpenoids with the southern cattle tick tyramine receptor: A potential biopesticide target. *Chemico-biological interactions*, v. 263, p. 1-6, 2017.

GÜEZ, C. M.; SOUZA, R. O. D.; FISCHER, P.; LEÃO, M. F. D. M.; DUARTE, J. A.; BOLIGON, A. A.; MACHADO, M. M. Evaluation of basil extract (*Ocimum basilicum* L.) on oxidative, anti-genotoxic and anti-inflammatory effects in human leukocytes cell cultures exposed to challenging agents. *Brazilian Journal of Pharmaceutical Sciences*, v. 53, n. 1, p. 1-12, 2017.

GUGLIELMONE, A. A.; BEATI, L.; BARROS-BATTESTI, D. M.; LABRUNA, M. B.; NAVA, S.; VENZAL, J. M.; ESTRADA-PEÑA, A. Ticks (Ixodidae) on humans in south america. *Experimental & applied acarology*, v. 40, n. 2, p. 83-100, 2006.

GUIMARÃES, A. G.; OLIVEIRA, M. A.; SANTOS ALVES, R.; PASSOS MENEZES, P.; SERAFINI, M. R.; SOUZA ARAÚJO, A. A.; JÚNIOR, L. J. Q. Encapsulation of carvacrol, a monoterpane present in the essential oil of oregano, with β -cyclodextrin, improves the pharmacological response on cancer pain experimental protocols. *Chemico-biological interactions*, v. 227, p. 69-76, 2015.

GÜLÇİN, I.; ELMASTAŞ, M.; ABOUL-ENEIN, H. Y. Determination of antioxidant and radical scavenging activity of Basil (*Ocimum basilicum* L. Family Lamiaceae) assayed by different methodologies. *Phytother Res*, v. 21, n. 4, p. 354-361, 2007.

GUPTA, P.; ROBIN, V. V.; DHARMARAJAN, G. Towards a more healthy conservation paradigm: integrating disease and molecular ecology to aid biological conservation. *Journal of Genetics*, v. 99, n. 1, p. 1-26, 2020.

HACKMAN, R.H.; FILSHIE, B.K. *The tick Cuticle*. In: OBENCHAIN, F.D., GALUN, R. (Eds.), *Physiology of Ticks*. Pergamon Press, Oxford, New York, Toronto, Sydney, Paris, Frankfurt, pp. 1-42, 1982.

HASSANPOURAGHDAM, M. B.; GOHARI, G. R.; TABATABAEI, S. J.; DADPOUR, M. R. Inflorescence and leaves essential oil composition of hydroponically grown *Ocimum basilicum* L. *Journal of the Serbian Chemical Society*, v. 75, n. 10, p. 1361-1368, 2010.

HEIMERDINGER, A.; OLIVO, C. J.; MOLENTO, M. B.; AGNOLIN, C. A.; ZIECH, M. F.; SCARAVELLI, L. F. B.; CHARÃO, P. S. Extrato alcoólico de Capim-cidreira (*Cymbopogon citratus*) no controle do *Boophilus microplus* em bovinos. *Revista Brasileira de Parasitologia Veterinária*, v. 15, n. 1, p. 37-39, 2006.

HEUKELBACH, J.; OLIVEIRA, F.A.S.; SPEARE, R. A new shampoo based on neem (*Azadirachta indica*) is highly effective against head lice *in vitro*. *Parasitology Research*, v. 99, n. 4, p. 353-356, 2006.

HIERRO, I.; VALERO, A.; NAVARRO, M. C. *In vivo* larvicidal activity of monoterpenic derivatives from aromatic plants against L3 larvae of *Anisakis simplex* sl. *Phytomedicine*, v. 13, n. 7, p. 527-531, 2006.

HIGA, L. D. O. S.; GARCIA, M. V.; BARROS, J. C.; KOLLER, W. W.; ANDREOTTI, R. Evaluation of *Rhipicephalus* (*Boophilus*) *microplus* (Acari: Ixodidae) resistance to different acaricide formulations using samples from Brazilian properties. *Revista Brasileira de Parasitologia Veterinária*, v. 25, p. 163-171, 2016.

HONER, M. R.; GOMES, A. *O manejo integrado de mosca dos chifres, berne e carrapato em gado de corte*. Campo Grande: EMBRAPA-CNPGL, 1990.

HYLDGAARD, M.; MYGIND, T.; MEYER, R.L. Essential oils in food preservation: mode of action, synergies, and interactions with food matrix components. *Frontiers in microbiology*, v. 3, p. 12, 2012.

JACQUIET, P.; CABARET, J.; THIAM, E.; CHEIKH, D. Host range and the maintenance of *Haemonchus* spp. in an adverse arid climate. *International journal for parasitology*, v. 28, n. 2, p. 253-261, 1998.

JANER, E. C.; KLAFKE, G. M.; CAPURRO, M. L.; SCHUMAKER, T. T. S. Crossresistance between fipronil and lindane in *Rhipicephalus* (*Boophilus*) *microplus*. *Vet. Parasitol.*, v. 210, p. 77-83, 2015.

JANER, E. C.; KLAFKE, G. M.; CAPURRO, M. L.; SCHUMAKER, T. T. S. Cross-resistance between fipronil and lindane in *Rhipicephalus* (*Boophilus*) *microplus*. *Veterinary parasitology*, v. 210, n. 1-2, p. 77-83, 2015.

JAYAKUMAR, S.; MADANKUMAR, A.; ASOKKUMAR, S.; RAGHUNANDHAKUMAR, S.; KAMARAJ, S.; JOSEPHINE DIVYA; M. G.; DEVAKI, T. Potential preventive effect of carvacrol against diethylnitrosamine-induced hepatocellular carcinoma in rats. *Molecular and cellular biochemistry*, v. 360, n. 1, p. 51-60, 2012.

JITTAPALAPONG, S.; STICH, R. W.; GORDON, J. C.; WITTUM, T. E.; BARRIGA, O. O. Performance of female *Rhipicephalus sanguineus* (Acari: Ixodidae) fed on dogs exposed to multiple infestations or immunization with tick salivary gland or midgut tissues. *J Med Entomol*, v. 37, p. 601-611, 2000.

JONGEJAN, F.; UILENBERG, G. The global importance of ticks. *Parasitology*, v. 129, n. 1, p. 3-14, 2004.

JONSSON, N. N.; MILLER, R. J.; KEMP, D. H.; KNOWLES, A.; ARDILA, A. E.; VERRALL, R. G.; ROTHWELL, J. T. Rotation of treatments between spinosad and amitraz for the control of *Rhipicephalus (Boophilus) microplus* populations with amitraz resistance. *Veterinary parasitology*, v. 169, n. 1-2, p. 157-164, 2010.

JORDAN, ROBERT A.; SCHULZE, TERRY L.; DOLAN, MARC C. Efficacy of plant-derived and synthetic compounds on clothing as repellents against *Ixodes scapularis* and *Amblyomma americanum* (Acari: Ixodidae). *Journal of medical entomology*, v. 49, n. 1, p. 101-106, 2012.

KABARA, J. J. Phenols and Chelators. In: RUSSELL, N. J.; GOULD, G. W. (eds). *Food preservatives*. Blackie; London, UK, v. 200, p. 214, 1991.

KATIKI, L. M.; BARBIERI, A. M. E.; ARAUJO, R. C.; VERÍSSIMO, C. J.; LOUVANDINI, H.; FERREIRA, J. F. S. Synergistic interaction of ten essential oils against *Haemonchus contortus* *in vitro*. *Veterinary Parasitology*, v. 243, p. 47-51, 2017.

KATIKI, L. M.; CHAGAS, A. C. S.; TAKAHIRA, R. K.; JULIANI, H. R.; FERREIRA, J. F. S.; AMARANTE, A. F. T. D. Evaluation of *Cymbopogon schoenanthus* essential oil in lambs experimentally infected with *Haemonchus contortus*. *Veterinary parasitology*, v. 186, n. 3-4, p. 312-318, 2012.

KESSLER, R. H. Considerações sobre a transmissão de *Anaplasma marginale*. *Pesquisa Veterinária Brasileira*, v. 21, p. 177-179, 2001.

KHAIR-BARIYAH, S.; AHMED, D.; IKRAM, M. *Ocimum basilicum*: a review on phytochemical and pharmacological studies. *J Chemistry*, v. 2, n. 2, p. 78-85, 2012.

KHAIR-UL-BARIYAH, S.; AHMED, D.; AUJLA, M. Ikram. Comparative Analysis of *Ocimum basilicum* and *Ocimum sanctum*: Extraction Techniques and Urease and alpha-Amylase inhibition. *Pak. J. Chem*, v. 2, n. 3, p. 134-141, 2012.

KHALILI, M.; SAKHAEE, E.; AFLATOONIAN, M. R.; SHAHABI-NEJAD, N. Herd-prevalence of *Coxiella burnetii* (Q fever) antibodies in dairy cattle farms based on bulk tank milk analysis. *Asian Pacific journal of tropical medicine*, v. 4, n. 1, p. 58-60, 2011.

KIFERLE, C.; ASCRIZZI, R.; MARTINELLI, M.; GONZALI, S.; MARIOTTI, L.; PISTELLI, L.; PERATA, P. Effect of Iodine treatments on *Ocimum basilicum* L.: Biofortification, phenolics production and essential oil composition. *PLoS One*, v. 14, n. 12, p. e0226559, 2019.

KIRIMER, N.; BASER, K.; TÜMEN, G. Carvacrol-rich plants in Turkey. *Chemistry of Natural Compounds*, v. 31, n. 1, p. 37-41, 1995.

KLAFKE, G.M.; SABATINI, G.A.; ALBUQUERQUE, T.A.; MARTINS, J.R.; KEMP, D.H.; MILLER, R. J.; SCHUMAKER, T.T Larval immersion tests with ivermectin in populations of the cattle tick *Rhipicephalus (Boophilus) microplus* (Acari: Ixodidae) from State of São Paulo, Brazil. *Veterinary parasitology*, v. 142, n. 3-4, p. 386-390, 2006.

KLAFKE; G. M. Resistência de *R. (B.) microplus* contra os carrapaticidas. In: PEREIRA, M. C. et al. (eds.). *Rhipicephalus (Boophilus) microplus: Biologia, controle e resistência*. São Paulo: MedVet Livros, 2008.

KOC, S.; OZ, E.; CINBILGEL, I.; AYDIN, L.; CETIN, H. Acaricidal activity of *Origanum bilgeri* PH Davis (Lamiaceae) essential oil and its major component, carvacrol against adults *Rhipicephalus turanicus* (Acari: Ixodidae). *Veterinary Parasitology*, v. 193, n. 1-3, p. 316-319, 2013.

KOTZE, A. C.; HUNT, P. W.; SKUCE, P.; VON SAMSON-HIMMELSTJERNA, G.; MARTIN, R. J.; SAGER, H.; PRICHARD, R. K. Recent advances in candidate-gene and whole-genome approaches to the discovery of anthelmintic resistance markers and the description of drug/receptor interactions. *International Journal for Parasitology: Drugs and Drug Resistance*, v. 4, n. 3, p. 164-184, 2014.

LABRUNA, M. B.; KAMAKURA, O.; MORAES-FILHO, J.; HORTA, M. C.; PACHECO, R. C. Rocky Mountain spotted fever in dogs, Brazil. *Emerging infectious diseases*, v. 15, n. 3, p. 458, 2009.

LABRUNA, M. B.; MCBRIDE, J. W.; BOUYER, D. H.; CAMARGO, L. M. A.; CAMARGO, E. P.; WALKER, D. H. Molecular evidence for a spotted fever group *Rickettsia species* in the tick *Amblyomma longirostre* in Brazil. *Journal of medical entomology*, v. 41, n. 3, p. 533-537, 2004.

LACEY, E.; BRADY, R. L.; PRICHARD, R. K.; WATSON, T. R. Comparison of inhibition of polymerisation of mammalian tubulin and helminth ovicidal activity by benzimidazole carbamates. *Veterinary parasitology*, v. 23, n. 1-2, p. 105-119, 1987.

LAMBERT, R. J. W.; SKANDAMIS, P. N.; COOTE, P. J.; NYCHAS, G. J. A study of the minimum inhibitory concentration and mode of action of oregano essential oil, thymol and carvacrol. *Journal of applied microbiology*, v. 91, n. 3, p. 453-462, 2001.

LANE, J.; JUBB, T.; SHEPHARD, R.; WEBB-WARE, J.; FORDYCE, G. Priority list of endemic diseases for the red meat industries. pp. 282. Sydney, Australia, *Meat and Livestock Australia*. 2015.

LANG, G.; BUCHBAUER, G. A review on recent research results (2008–2010) on essential oils as antimicrobials and antifungals. A review. *Flavour and Fragrance Journal*, v. 27, n. 1, p. 13-39, 2012.

LARA, D. M. *Resistencia a los antihelminticos en nematodos de rumiantes y estrategias para su control*. Bogotá: Corpoca, 2007.

LARSEN, M. Biological control in a global perspective - a review with emphasis on *Duddingtonia flagrans*. In: FAO. Animal Production and Health Division. Biological control of nematode parasites of small ruminants in Asia. Final proceedings... Rome, Italy: FAO, 2002.

LATREILLE, P.A. *Genera crustaceorum et insectorum secundum ordinem naturalem in familia disposita, iconibus exemplisque plurimis explicata*, vol. 1. Parisiis et Argentorati: Amand Koenig, bibliopolam, 1806.

LAWRENCE, B. M. Chemical components of *Labiatae* oils and their exploitation. *Advances in Labiatae science*, p. 399-436, 1992.

LE JAMBRE, L.F. Relationship of blood loss to worm numbers, biomass and egg production in *Haemonchus contortus* infected sheep. *International Journal for Parasitology*, v. 25, p. 269-273, 1995.

LEES, A.D. Transpiration and the structure of the epicuticle in ticks. *The Journal of Experimental Biology*, v. 23, n. 3-4, p. 379-410, 1947.

LEI, J.; LESER, M.; ENAN, E. Nematicidal activity of two monoterpenoids and SER-2 tyramine receptor of *Caenorhabditis elegans*. *Biochemica pharmacology*, v. 79, n. 7, p. 1062-1071, 2010.

LEVINE, N. D. Weather and the ecology of bursate nematodes. *International Journal of Biometeorology*, v. 24, n. 4, p. 341-346, 1980.

LI, A. Y.; DAVEY, R. B.; MILLER, R. J.; GEORGE, J. E. Detection and characterization of amitraz resistance in the southern cattle tick, *Boophilus microplus* (Acari: Ixodidae). *Journal of medical entomology*, v. 41, n. 2, p. 193-200, 2004.

LIBER, Z.; CAROVIĆ-STANKO, K.; POLITEO, O.; STRIKIĆ, F.; KOLAK, I.; MILOS, M.; SATOVIC, Z. Chemical characterization and genetic relationships among *Ocimum basilicum* L. cultivars. *Chemistry & Biodiversity*, v. 8, n. 11, p. 1978-1989, 2011.

LIMA, A.S.; LANDULFO, G.A.; COSTA-JÚNIOR, L.M. Repellent effects of encapsulated carvacrol on the *Rhipicephalus (Boophilus) microplus* (Acari: Ixodidae). *Journal of medical entomology*, v. 56, n. 3, p. 881-885, 2019.

LIMA, A.S.; MACIEL, A.P.; MENDONÇA, C.J.S.; COSTA-JUNIOR, L.M. Use of encapsulated carvacrol with yeast cell walls to control resistant strains of *Rhipicephalus microplus* (Acari: Ixodidae). *Industrial Crops and Products*, v. 108, p. 190-194, 2017.

LOPES, A. J. O. **Estudo computacional da interação de terpenos com acetilcolinesterase de *Rhipicephalus microplus* e potenciais novos candidatos a carrapaticidas.** 2015. 74 f. Dissertação (Programa de Pós-Graduação em Ciências da Saúde) - Universidade Federal do Maranhão, São Luís, 2015.

LOPES, W. D. Z.; CRUZ, B. C.; TEIXEIRA, W. F. P.; FELIPPELLI, G.; MACIEL, W. G.; BUZZULINI, C.; COSTA, A. J. Efficacy of fipronil (1.0 mg/kg) against *Rhipicephalus (Boophilus) microplus* strains resistant to ivermectin (0.63 mg/kg). *Preventive veterinary medicine*, v. 115, n. 3-4, p. 88-93, 2014.

LOSADA, I. J.; SILVA, R.; LOSADA, M. A. Interaction of non-breaking directional random waves with submerged breakwaters. *Coastal Engineering*, v. 28, n. 1-4, p. 249-266, 1996.

LOULY, C. C. B.; FONSECA, I. N.; OLIVEIRA, V. F. D.; LINHARES, G. F. C.; MENEZES, L. B. D.; BORGES, L. M. F. Seasonal dynamics of *Rhipicephalus sanguineus* (Acari: Ixodidae) in dogs from a police unit in Goiania, Goias, Brazil. *Ciência Rural*, v. 37, p. 464-469, 2007.

LOVIS, L.; MENDES, M. C.; PERRET, J. L.; MARTINS, J. R.; BOUVIER, J.; BETSCHART, B.; SAGER, H. Use of the Larval Tarsal Test to determine acaricide resistance in *Rhipicephalus (Boophilus) microplus* Brazilian field populations. *Vet. Parasitol.*, v. 191, p. 323–331, 2013.

LYNAGH, T.; LAUBE, B. Opposing effects of the anesthetic propofol at pentameric ligand-gated ion channels mediated by a common site. *Journal of Neuroscience*, v. 34, n. 6, p. 2155-2159, 2014.

MACEDO, I. T. F.; OLIVEIRA, L. M. B. D.; ANDRÉ, W. P. P.; ARAÚJO, J. V. D.; SANTOS, J. M. L. D.; RONDON, F. C. M.; BEVILAQUA, C. M. L. Anthelmintic effect of *Cymbopogon citratus* essential oil and its nanoemulsion on sheep gastrointestinal nematodes. *Revista Brasileira de Parasitologia Veterinária*, v. 28, p. 522-527, 2019.

MACIEL, M. V.; MORAIS, S. M.; BEVILAQUA, C. M. L.; AMÓRA, S. S. A. Extratos vegetais usados no controle de dípteros vetores de zoonoses. *Rev. Bras. Plantas Med.* v. 12, n. 1, p. 105-112, 2010.

MACIEL, M. V.; MORAIS, S. M.; BEVILAQUA, C. M. L.; CAMURÇA-VASCONCELOS, A. L. F.; COSTA, C. T. C.; CASTRO, C. M. S. Ovicidal and larvicidal activity of *Melia azedarach* extracts on *Haemonchus contortus*. *Veterinary parasitology*, v. 140, n. 1-2, p. 98-104, 2006.

MAJUMDER, S. P.; DAS, A. C. Phosphate-solubility and phosphatase activity in *Gangetic alluvial* soil as influenced by organophosphate insecticide residues. *Ecotoxicology and Environmental Safety*, v. 126, p. 56-61, 2016.

MALLATOU, H.; PAPPAS, C. P.; KONDYLI, E.; ALBANIS, T. A. Pesticide residues in milk and cheeses from Greece. *Science of the total environment*, v. 196, n. 2, p. 111-117, 1997.

MANDELKOW, E.; MANDELKOW, E. M. Microtubular structure and tubulin polymerization. *Current Opinion in Cell Biology*, v. 1, n. 1, p. 5-9, 1989.

MARTIN, R. J. Modes of action of anthelmintic drugs. *The Veterinary Journal*, v. 154, n. 1, p. 11-34, 1997.

MARTIN, R. J.; ROBERTSON, A. P.; BJORN, H. Target sites of anthelmintics. *Parasitology*, v. 114, n. 7, p. 111-124, 1997.

MARTINEZ-TORRES, D.; DEVONSHIRE, A. L.; WILLIAMSON, M. S. Molecular studies of knockdown resistance to pyrethroids: cloning of domain II sodium channel gene sequences from insects. *Pesticide Science*, v. 51, n. 3, p. 265-270, 1997.

MARTINEZ-VELAZQUEZ, M.; CASTILLO-HERRERA, G. A.; ROSARIO-CRUZ, R.; FLORES-FERNANDEZ, J. M.; LOPEZ-RAMIREZ, J.; HERNANDEZ-GUTIERREZ, R.; DEL CARMEN LUGO-CERVANTES, E. Acaricidal effect and chemical composition of essential oils extracted from *Cuminum cyminum*, *Pimenta dioica* and *Ocimum basilicum* against the cattle tick *Rhipicephalus (Boophilus) microplus* (Acari: Ixodidae). *Parasitology research*, v. 108, n. 2, p. 481-487, 2011.

MARTINS, A. G. L. D. A.; NASCIMENTO, A. R.; MOUCHREK FILHO, J. E.; MENDES FILHO, N. E.; SOUZA, A. G.; ARAGÃO, N. E.; SILVA, D. S. V. D. Atividade antibacteriana do óleo essencial do manjericão frente a sorogrupos de *Escherichia coli* enteropatogênica isolados de alfaces. *Ciência Rural*, v. 40, p. 1791-1796, 2010.

MARTINS, A. G. L. D. A.; NASCIMENTO, A. R.; MOUCHREK FILHO, J. E.; MENDES FILHO, N. E.; SOUZA, A. G.; ARAGÃO, N. E.; SILVA, D. S. V. D. Atividade antibacteriana do óleo essencial do manjericão frente a sorogrupos de *Escherichia coli* enteropatogênica isolados de alfaces. *Ciência Rural*, v. 40, p. 1791-1796, 2010.

MARTINS, A. J.; VALLE, D. The pyrethroid knockdown resistance. *Insecticides-Basic and Other Applications*, v. 17, p. 38, 2012.

MCMURRY, J. *Química orgânica - Combo*. 7. ed. São Paulo: Cengage Learning, 2011.

MEEUSEN, E. N.; WALKER, J.; PETERS, A.; PASTORET, P. P.; JUNGERSEN, G. Current status of veterinary vaccines. *Clinical microbiology reviews*, v. 20, n. 3, p. 489-510, 2007.

MELO, L. M.; BEVILAQUA, C. M. L.; ARAÚJO, J. V. D.; MELO, A. C. F. L. Atividade predatória do fungo *Monacrosporium thaumasium* contra o nematóide *Haemonchus contortus*, após passagem pelo trato gastrintestinal de caprinos. *Ciência Rural*, v. 33, p. 169-171, 2003.

MENDONÇA, A.E.; MOREIRA, R.G.; AMARAL, M.P.H.; MONTEIRO, C.M.O.; MELLO, V.; VILELA, F.M.P.; MENDONÇA HOMEM, F.C.; FURLONG, J.; DOLINSKI, C.; PRATA, M.C.A.; CHAGAS, E.F. Entomopathogenic nematodes in pharmaceutical formulations for *Rhipicephalus microplus* (Acari: Ixodidae) control: In

vitro evaluation of compatibility, thermotolerance, and efficiency. *Ticks and tick-borne diseases*, v. 10, n. 4, p. 781-786, 2019.

MILLER, R. J.; GEORGE, J. E.; GUERRERO, F.; CARPENTER, L.; WELCH, J. B. Characterization of acaricide resistance in *Rhipicephalus sanguineus* (Latreille) (Acari: Ixodidae) collected from the Corozal army veterinary quarantine center, Panama. *Journal of medical entomology*, v. 38, n. 2, p. 298-302, 2001.

MOLENTO, M. B.; BRANDÃO, Y. O. Macrocyclic lactone resistance in nematodes of cattle in Brazil: Blame it to the ticks!. *Parasitology International*, p. 102588, 2022.

MOLENTO, M. B.; NIELSEN, M. K.; KAPLAN, R. M. Resistance to avermectin/milbemycin, anthelmintics in equine cyathostomins—current situation. *Veterinary Parasitology*, v. 185, n. 1, p. 16-24, 2012.

MONTEIRO, C. M. O.; DAEMON, E.; APARECIDO CLEMENTE, M.; SANTOS ROSA, L.; MATURANO, R. Acaricidal efficacy of thymol on engorged nymphs and females of *Rhipicephalus sanguineus* (Latreille, 1808) (Acari: Ixodidae). *Parasitology research*, v. 105, n. 4, p. 1093-1097, 2009.

MORAIS, S. M.; VILA-NOVA, N. S.; BEVILAQUA, C. M. L.; RONDON, F. C.; LOBO, C. H.; MOURA, A. D. A. A. N.; ANDRADE JR, H. F. Thymol and eugenol derivatives as potential antileishmanial agents. *Bioorganic & medicinal chemistry*, v. 22, n. 21, p. 6250-6255, 2014.

MOTA, M. A.; CAMPOS, A. K.; ARAÚJO, J. V. Controle biológico de helmintos parasitos de animais: estágio atual e perspectivas futuras. *Pesquisa Veterinária Brasileira*, v. 23, n. 3, p. 93-100, 2003.

MUELLER, R. S. Treatment protocols for demodicosis: an evidence-based review. *Veterinary Dermatology*, v. 15, n. 2, p. 75–89, 2004.

MUNIZ, V. M. Desenvolvimento de micropartícula contendo clorexidina e timol. 2018. 139 f. Dissertação (Mestrado) - Universidade Federal de Pernambuco. Centro de Ciências da Saúde. Programa de pós-graduação em Ciências Farmacêuticas, Recife, 2018.

NEUMANN, L. G. *Ixodidae in “Das Tierreich”, im Auftrage der K. preuss. Akad. d. Wiss. zu Berlin herausg.* v. F. E. Schulze. Berlin: Friedlander, 1911.

NEVES, J.; PINTO, E.; AMARAL, M. H.; BAHIA, M. F. Antifungal activity of a gel containing *Thymus vulgaris* essential oil against *Candida* species commonly involved in vulvovaginal candidosis. *Pharmaceutical Biology*, v. 47, n. 2, p. 151-153, 2009.

NIEUWHOF, G. J.; BISHOP, S. C. Costs of the major endemic diseases of sheep in Great Britain and the potential benefits of reduction in disease impact. *Animal Science*, v. 81, n. 1, p. 23-29, 2005.

NOGUEIRA PRISTA, L.; ALVES, A. C.; MORGADO, R. Tecnologia Farmacêutica. Fundação Calouste Gulbenkian, v. 7, p. 150-152, 2008.

NOGUEIRA PRISTA, L.; CORREIA ALVES, A.; MORGADO, R.; SOUSA LOBO, J. *Tecnologia Farmacêutica, I Volume.* Lisboa: Fundação Calouste Gulbenkian, 2003.

NOGUEIRA, B. C. F.; CAMPOS, A. K.; MUÑOZ-LEAL, S.; PINTER, A.; MARTINS, T. F. Oft and hard ticks (Parasitiformes: Ixodida) on humans: A review of Brazilian biomes and the impact of environmental change. *Acta Tropica*, p. 106598, 2022.

NOVATO, T. L. P.; MARCHEZINI, P.; MUNIZ, N.; AZEVEDO PRATA, M. C. FURLONG, J.; VILELA, F. M. P.; MONTEIRO, C. Evaluation of synergism and development of a formulation with thymol, carvacrol and eugenol for *Rhipicephalus microplus* control. *Experimental parasitology*, v. 207, p. 107774, 2019.

NOVATO, T. P. L.; ARAÚJO, L. X.; MONTEIRO, C. M. O.; MATURANO, R.; SENRA, T. D. O. S.; SILVA MATOS, R.; DAEMON, E. Evaluation of the combined effect of thymol, carvacrol and (E)-cinnamaldehyde on *Amblyomma sculptum* (Acari: Ixodidae) and *Dermacentor nitens* (Acari: Ixodidae) larvae. *Veterinary parasitology*, v. 212, n. 3-4, p. 331-335, 2015.

NOVATO, T.; GOMES, G. A.; ZERINGÓTA, V.; FRANCO, C. T.; OLIVEIRA, D. R.; MELO, D.; OLIVEIRA MONTEIRO, C. M. *In vitro* assessment of the acaricidal activity of carvacrol, thymol, eugenol and their acetylated derivatives on *Rhipicephalus microplus* (Acari: Ixodidae). *Veterinary parasitology*, v. 260, p. 1-4, 2018.

O'CONNOR, L. J.; KAHN, L. P.; WALKDEN-BROWN, S. W. The effects of amount, timing and distribution of simulated rainfall on the development of *Haemonchus contortus* to the infective larval stage. *Veterinary parasitology*, v. 146, n. 1-2, p. 90-101, 2007.

OLIVEIRA SOUZA HIGA, L.; GARCIA, M. V.; BARROS, J. C.; KOLLER, W. W.; ANDREOTTI, R. Acaricide resistance status of the *Rhipicephalus microplus* in Brazil: a literature overview. *Med chem*, v. 5, p. 326-333, 2015.

OLIVEIRA, T. M.; RIBEIRO, F. W.; SOUSA, C. P.; SALAZAR-BANDA, G. R.; LIMA-NETO, P.; CORREIA, A. N.; MORAIS, S. Current overview and perspectives on carbon-based (bio) sensors for carbamate pesticides electroanalysis. *Trends in Analytical Chemistry*, v. 124, p. 115779, 2020.

OSTERROHT, M. V.; WITZLER, L.; PIMENTA, S.; BRUM, C. "Passos para conversão à pecuária orgânica" e "Pecuária orgânica no Grupo Independência". *Agroecologia Hoje*, v. 2, n. 13, p. 15-20, 2002.

OTTAI, M. E. S.; AHMED, S. S.; EL-DIN, M. M. Genetic variability among some quantitative characters, insecticidal activity and essential oil composition of two Egyptian and French sweet basil varieties. *Australian Journal of Basic and Applied Sciences*, v. 6, n. 3, p. 185-192, 2012.

PADUCH, R.; KANDEFER-SZERSZEŃ, M.; TRYTEK, M.; FIEDUREK, J. Terpenes: substances useful in human healthcare. *Archivum immunologiae et therapiae experimentalis*, v. 55, n. 5, p. 315-327, 2007.

PARK, J. H.; JEON, Y. J.; LEE, C. H.; CHUNG, N.; LEE, H. S. Insecticidal toxicities of carvacrol and thymol derived from *Thymus vulgaris* Lin. against *Pochazia shantungensis* Chou & Lu., newly recorded pest. *Scientific reports*, v. 7, n. 1, p. 1-7, 2017.

PAROLA, P.; SOCOLOVSCHI, C.; JEANJEAN, L.; BITAM, I.; FOURNIER, P. E.; SOTTO, A.; RAOULT, D. Warmer weather linked to tick attack and emergence of severe rickettsioses. *PLoS neglected tropical diseases*, v. 2, n. 11, p. e338, 2008.

PASCUAL-VILLALOBOS, M. J.; BALLESTA-ACOSTA, M. C. Chemical variation in an *Ocimum basilicum* germplasm collection and activity of the essential oils on *Callosobruchus maculatus*. *Biochemical Systematics and Ecology*, v. 31, n. 7, p. 673-679, 2003.

PEIXOTO, M. G.; COSTA-JÚNIOR, L. M.; BLANK, A. F.; SILVA LIMA, A.; MENEZES, T. S. A.; ALEXANDRIA SANTOS, D.; FÁTIMA ARRIGONI-BLANK, M. Acaricidal activity of essential oils from *Lippia alba* genotypes and its major components carvone, limonene, and citral against *Rhipicephalus microplus*. *Veterinary parasitology*, v. 210, n. 1-2, p. 118-122, 2015.

PEIXOTO-NEVES, D.; SILVA-ALVES, K. S.; GOMES, M. D. M.; LIMA, F. C.; LAHLOU, S.; MAGALHÃES, P. J. C.; LEAL-CARDOSO, J. H. Vasorelaxant effects of the monoterpenic phenol isomers, carvacrol and thymol, on rat isolated aorta. *Fundamental & clinical pharmacology*, v. 24, n. 3, p. 341-350, 2010.

PEREIRA, M.; MATIAS, D.; PEREIRA, F.; REIS, C. P.; SIMÕES, M. F.; RIJO, P. Antimicrobial screening of *Plectranthus madagascariensis* and *P. neochilus* extracts. *Biomed Biopharm Res*, v. 12, p. 127-38, 2015.

PEREIRA-JUNIOR, R. A.; SOUSA, S. A. P.; VELOSO, F. P. F. D. S.; SILVA, L. A. D.; ALMEIDA, K. D. S. Eficácia de ivermectina e albendazol contra nematódeos gastrintestinais em rebanho ovino naturalmente infectado no município de Palmas-TO, Brasil. *Revista Científica de Medicina Veterinária*, v. 28, p. 1-10, 2017.

PERRY, R. N.; MOENS, M. *Introduction to plant-parasitic nematodes; modes of parasitism*. In: JONES, J.; GHEYSEN, G.; FENOLL, C. (eds.). Genomics and molecular genetics of plant-nematode interactions. New York: Springer, 2011.

PINTO, J. A. O.; FITZGERALD BLANK, A.; LIMA NOGUEIRA, P. C.; ARRIGONI-BLANK, M. D. F.; MATOS ANDRADE, T.; SANTOS SAMPAIO, T.; GARCIA PEREIRA, K. L. Chemical characterization of the essential oil from leaves of basil genotypes cultivated in different seasons. *Boletín Latinoamericano y del Caribe de Plantas Medicinales y Aromáticas*, v. 18, n. 1, 2019.

PONTES NETTO, D. Resíduos químicos no leite: risco à saúde pública. *I Seminário Estadual de Resíduos Químicos em Alimentos*, Foz do Iguaçu, 22 a 24 set. 2004.

POUND, J. M.; GEORGE, J. E.; KAMMLAH, D. M.; LOHMEYER, K. H.; DAVEY, R. B. Evidence for role of white-tailed deer (Artiodactyla: Cervidae) in epizootiology of bovina carrapatos e sul bovinos carrapatos (Acari: Ixodidae) em reinfestações ao longo

do Texas/Fronteira do México no sul do Texas: uma revisão e atualização. *J Econ Entomol.*, v. 103, p. 211–218, 2010.

PRATES, H. T.; OLIVEIRA, A. B.; LEITE, R. C.; CRAVEIRO, A. A. Atividade carrapaticida e composição química do óleo essencial do capim-gordura. *Pesquisa Agropecuária Brasileira*, v. 28, n. 5, p. 621-625, 1993.

PRETTE, N.; MONTEIRO, A. C.; GARCIA, M. V.; SOARES, V. E. Patogenicidade de isolados de *Beauveria bassiana* para ovos, larvas e ninfas ingurgitadas de *Rhipicephalus sanguineus*. *Ciência Rural*, v. 35, p. 855-861, 2005.

PUSHPANGADAN, P.; GEORGE, V. Basil. In: PETER, K. V. (ed). *Handbook of Herbs and Spices*. Cambridge England: Woodhead Publishing, 2012.

QAMAR, M. F.; MAQBOOL, A. Biochemical studies and serodiagnosis of haemonchosis in sheep and goats. *J. Anim. Plant Sci*, v. 22, n. 1, p. 32-38, 2012.

RAOULT, D.; LAURENT, J. C.; MUTILLOD, M. Monoclonal antibodies to *Coxiella burnetii* for antigenic detection in cell cultures and in paraffin-embedded tissues. *American journal of clinical pathology*, v. 101, n. 3, p. 318-320, 1994.

RATTAN, R. S. Mechanism of action of insecticidal secondary metabolites of plant origin. *Crop protection*, v. 29, n. 9, p. 913-920, 2010.

RAUH, J.; LUMMIS, S.; SATTELLE, D. Pharmacological and biochemical properties of insect GABA receptors. *Trends Pharmacol. Sci*, v. 11, p. 325-329, 1990.

Ravid, U.; Putievsky, E.; Katzir, I.; Lewinsohn, E. Enantiomeric composition of linalol in the essential oils of *Ocimum species* and in commercial basil oils. *Flavour and fragrance journal*, v. 12, n. 4, p. 293-296, 1997.

RAYNAL, J. T.; SILVA, A. A. B. D.; SOUSA, T. D. J.; BAHIENSE, T. C.; MEYER, R.; PORTELA, R. W. Acaricides efficiency on *Rhipicephalus (Boophilus) microplus* from Bahia state North-Central region. *Revista Brasileira de Parasitologia Veterinária*, v. 22, p. 71-77, 2013.

RECK, J.; SOUZA, U.; SOUZA, G.; KIELING, E.; DALL'AGNOL, B.; WEBSTER, A.; MARTINS, J. R. Records of ticks on humans in Rio Grande do Sul state, Brazil. *Ticks and tick-borne diseases*, v. 9, n. 5, p. 1296-1301, 2018.

REGITANO, L. C. A.; PRAYAGA, K. *Ticks and tick-borne diseases in cattle*. In: BISOP, S. C. et al. (eds.). *Breeding for disease resistance in farm animals*. Oxfordshire: CAB International, 2010.

RENÉ-MARTELLET, M.; MORO, C. V.; CHÈNE, J.; BOURDOISEAU, G.; CHABANNE, L.; MAVINGUI, P. Update on epidemiology of canine babesiosis in Southern France. *BMC Vet Res*, v. 25, n. 11, p. 1-11, 2015.

ROBERTS, F. H. S.; O'SULLIVAN, P. J. Methods for egg counts and larval cultures for strongyles infesting the gastro-intestinal tract of cattle. *Australian Journal of*

Agricultural Research, v. 1, n. 1, p. 99-102, 1950.

ROBERTS, J. A. Resistance of cattle to the tick *Boophilus microplus* (Canestrini). II. Stages of the life cycle of the parasite against which resistance is manifest. *The Journal of Parasitology*, v. 54, n. 4, p. 667-673, 1968.

RODRIGUES, L. **Toxicidade do odor de óleos essenciais de *Eucalyptus globulus* e *Corymbia citriodora* sobre o carrapato *Rhipicephalus microplus***. 2018. 79f. Dissertação (mestrado) – Instituto de Zootecnia. APTA/SAA, Nova Odessa, SP, 2018.

RODRÍGUEZ, A. V.; GOLDBERG, V.; VIOTTI, H.; CIAPPESONI, G. Early detection of *Haemonchus contortus* infection in sheep using three different faecal occult blood tests. *Open Veterinary Journal*, v. 5, n. 2, p. 90-97, 2015.

RODRÍGUEZ-GONZÁLEZ, Á.; ÁLVAREZ-GARCÍA, S.; GONZÁLEZ-LÓPEZ, Ó.; SILVA, F.; CASQUERO, P. A. Insecticidal properties of *Ocimum basilicum* and *Cymbopogon winterianus* against *Acanthoscelides obtectus*, insect pest of the common bean (*Phaseolus vulgaris*, L.). *Insects*, v. 10, n. 5, p. 151, 2019.

RODRIGUEZ-VIVAS, R. I.; JONSSON, N. N.; BHUSHAN, C. Strategies for the control of *Rhipicephalus microplus* ticks in a world of conventional acaricide and macrocyclic lactone resistance. *Parasitology research*, v. 117, n. 1, p. 3-29, 2018.

RODRIGUEZ-VIVAS, R. I.; OJEDA-CHI, M. M.; TRINIDAD-MARTINEZ, I.; LEÓN, A. P. First documentation of ivermectin resistance in *Rhipicephalus sanguineus* sensu lato (Acari: Ixodidae). *Veterinary parasitology*, v. 233, p. 9-13, 2017.

RODRIGUEZ-VIVAS, R. I.; TREES, A. J.; ROSADO-AGUILAR, J. A.; VILLEGAS-PEREZ, S. L.; HODGKINSON, J. E. Evolution of acaricide resistance: phenotypic and genotypic changes in field populations of *Rhipicephalus* (*Boophilus*) *microplus* in response to pyrethroid selection pressure. *International journal for parasitology*, v. 41, n. 8, p. 895-903, 2011.

ROEBER, F.; JEX, A. R.; GASSER, R. B. Impact of gastrointestinal parasitic nematodes of sheep, and the role of advanced molecular tools for exploring epidemiology and drug resistance-an Australian perspective. *Parasites & vectors*, v. 6, n. 1, p. 1-13, 2013.

ROEL, A. R.; VENDRAMIM, J. D. Residual effect of ethyl acetate extract of *Trichilia pallida* Swartz (Meliaceae) for *Spodoptera frugiperda* (JE Smith, 1797) (Lepidoptera: Noctuidae) larvae of different ages. *Ciência Rural*, v. 36, n. 4, p. 1049-1054, 2006.

ROMA, G. C.; OLIVEIRA, P. R.; PIZANO, M. A.; MATHIAS, M. I. C. Determination of LC₅₀ of permethrin acaricide in semi-engorged females of the tick *Rhipicephalus sanguineus* (Latreille, 1806) (Acari: Ixodidae). *Experimental Parasitology*, v. 123, n. 3, p. 269-272, 2009.

ROTHWELL, J. *Modern chemical treatments for sheep infested with external parasites*. In: HOLDSWORTH, P. A. (ed.). Ectoparasiticide use in contemporary Australian livestock production. Camberra: Avicare Limited, 2005.

- RYU, V.; MCCLEMENTS, D. J.; CORRADINI, M. G.; YANG, J. S.; MCLANDSBOROUGH, L. Natural antimicrobial delivery systems: Formulation, antimicrobial activity, and mechanism of action of quillaja saponin-stabilized carvacrol nanoemulsions. *Food Hydrocolloids*, v. 82, p. 442-450, 2018.
- SAJJADI, S.E. Analysis of the essential oils of two cultivated basil (*Ocimum basilicum* L.) from Iran. *Daru- Journal of Faculty of Pharmacy*, v. 14, p. 128–130, 2006.
- SALES, H. J. S. P. Lavandula L. Aplicação da cultura in vitro à produção de óleos essenciais e seu potencial económico em Portugal. *Revista Brasileira de Plantas Medicinais*, v. 17, p. 992-999, 2015.
- SANGSTER, N. C. Managing parasiticide resistance. *Veterinary Parasitology*, v. 98, n. 1-3, p. 89-109, 2001.
- SANTORO, G. F.; CARDOSO, M. G.; GUIMARÃES, L. G. L.; MENDONÇA, L. Z.; SOARES, M. J. *Trypanosoma cruzi*: activity of essential oils from *Achillea millefolium* L., *Syzygium aromaticum* L. and *Ocimum basilicum* L. on epimastigotes and trypomastigotes. *Experimental parasitology*, v. 116, n. 3, p. 283-290, 2007.
- SANTOS, E. G. G. D.; BEZERRA, W. A. D. S.; TEMEYER, K. B.; LEÓN, A. A.; COSTA-JUNIOR, L. M.; SOARES, A. M. D. S. Effects of essential oils on native and recombinant acetylcholinesterases of *Rhipicephalus microplus*. *Braz J Vet Parasitol*, v. 30, n. 2, p. 1-8, 2021.
- SANTOS, F. C. C.; VOGEL, F. S. F.; MONTEIRO, S. G. Efeito do óleo essencial de manjericão (*Ocimum basilicum* L.) sobre o carapato bovino *Rhipicephalus (Boophilus) microplus* em ensaios *in vitro*. *Semina: Ciências Agrárias*, v. 33, n. 3, p. 1133-1139, 2012.
- SAXENA, V. K.; MAHESHWARI, U. K. Seasonal incidence of *Rhipicephalus sanguineus* (Lat) (Acarina) on a wild host *Hemiechinus auritus* collaris (Gray) (Insectivora). *J Commun Dis.*, v. 17, n. 3, p. 227-229.1985.
- SCHAEFFER, J. M.; HAINES, H. W. Avermectin binding in *Caenorhabditis elegans*: a two-state model for the avermectin binding site. *Biochemical pharmacology*, v. 38, n. 14, p. 2329-2338, 1989.
- SCHUELE, G.; BARNETT, S.; BAPST, B.; CAVALIERO, T.; LUEMPERT, L.; STREHLAU, G.; JUNQUERA, P. The effect of water and shampooing on the efficacy of a pyriproxyfen 12.5% topical solution against brown dog tick (*Rhipicephalus sanguineus*) and cat flea (*Ctenocephalides felis*) infestations on dogs. *Veterinary parasitology*, v. 151, n. 2-4, p. 300-311, 2008.
- SCORALIK, M. G.; DAEMON, E.; OLIVEIRA MONTEIRO, C. M.; MATURANO, R. Enhancing the acaricide effect of thymol on larvae of the cattle tick *Rhipicephalus microplus* (Acari: Ixodidae) by solubilization in ethanol. *Parasitology research*, v. 110, n. 2, p. 645-648, 2012.

SENRA, T. O. S.; CALMON, F.; ZERINGÓTA, V.; MONTEIRO, C. M. O.; MATURANO, R.; SILVA MATOS, R.; DAEMON, E. Investigation of activity of monoterpenes and phenylpropanoids against immature stages of *Amblyomma cajennense* and *Rhipicephalus sanguineus* (Acari: Ixodidae). *Parasitology research*, v. 112, n. 10, p. 3471-3476, 2013.

SENRA, T. O. S.; ZERINGÓTA, V.; OLIVEIRA MONTEIRO, C. M.; CALMON, F.; MATURANO, R.; GOMES, G. A.; DAEMON, E. Assessment of the acaricidal activity of carvacrol, (E)-cinnamaldehyde, trans-anethole, and linalool on larvae of *Rhipicephalus microplus* and *Dermacentor nitens* (Acari: Ixodidae). *Parasitology research*, v. 112, n. 4, p. 1461-1466, 2013.

SHAROPOV, F. S.; SATYAL, P.; ALI, N. A. A.; POKHAREL, S.; ZHANG, H.; WINK, M.; SETZER, W. N. The essential oil compositions of *Ocimum basilicum* from three different regions: Nepal, Tajikistan, and Yemen. *Chemistry & biodiversity*, v. 13, n. 2, p. 241-248, 2016.

SHOOP, W. L.; MROZIK, H.; FISHER, M. H. Structure and activity of avermectins and milbemycins in animal health. *Veterinary parasitology*, v. 59, n. 2, p. 139-156, 1995.

SIANI, A. C.; SAMPAIO, A.; SOUSA, M.; HENRIQUES, M.; RAMOS, M. Óleos essenciais–potencial anti-inflamatório. *Biotecnologia Ciência & Desenvolvimento*, v. 23, p. 38-43, 2006.

SILVA LIMA, A.; CARVALHO, J. F.; PEIXOTO, M. G.; BLANK, A. F.; BORGES, L. M. F.; COSTA JUNIOR, L. M. Assessment of the repellent effect of *Lippia alba* essential oil and major monoterpenes on the cattle tick *Rhipicephalus microplus*. *Medical and Veterinary Entomology*, v. 30, n. 1, p. 73-77, 2016.

SILVA, C. R.; LIFSCHITZ, A. L.; MACEDO, S. R.; CAMPOS, N. R.; VIANA-FILHO, M.; ALCÂNTARA, A. C.; COSTA-JUNIOR, L. M. Combination of synthetic anthelmintics and monoterpenes: Assessment of efficacy, and ultrastructural and biophysical properties of *Haemonchus contortus* using atomic force microscopy. *Veterinary Parasitology*, v. 290, p. 109345, 2021.

SILVA, J.; ABEBE, W.; SOUSA, S. M.; DUARTE, V. G.; MACHADO, M. I. L.; MATOS, F. J. A. Analgesic and anti-inflammatory effects of essential oils of *Eucalyptus*. *Journal of ethnopharmacology*, v. 89, n. 2-3, p. 277-283, 2003.

SILVA, M. E.; ARAÚJO, J. V.; SILVEIRA, W. F.; CARVALHO, L. M.; RIBEIRO, R. R. Effectiveness of *Cratylia argentea* as an animal feed supplement in the control of gastrointestinal nematodes in sheep. *Semina: Ciências Agrárias*, v. 39, n. 2, p. 657-665, 2018.

SILVA, N. B.; TAUS, N. S.; JOHNSON, W. C.; MIRA, A.; SCHNITTGER, L., VALENTE, J. D.; VIEIRA, R. F. First report of *Anaplasma marginale* infection in goats, Brazil. *PLoS One.*, v. 13, n. 8, p. 1-6, 2018.

SILVA, S. T.; BERTOLUCCI, S. K. V.; CUNHA, S. H. B.; LAZZARINI, L. E. S.; TAVARES, M. C.; PINTO, J. E. B. P. Effect of light and natural ventilation systems on the growth parameters and carvacrol content in the *in vitro* cultures of *Plectranthus amboinicus* (Lour.) Spreng. *Plant Cell, Tissue and Organ Culture (PCTOC)*, v. 129, n. 3, p. 501-510, 2017.

SILVA, V. A.; SOUSA, J. P.; GUERRA, F. Q. S.; PESSÔA, H. L. F.; FREITAS, A. F. R.; ALVES, L. B. N.; LIMA, E. O. Antibacterial activity of *Ocimum basilicum* essential oil and linalool on bacterial isolates of clinical importance. *International Journal of Pharmacognosy and Phytochemical Research*, v. 7, n. 6, p. 1066-1071, 2015.

SILVA, V. A.; SOUSA, J. P.; GUERRA, F. Q. S.; PESSÔA, H. L. F.; FREITAS, A. F. R.; ALVES, L. B. N.; LIMA, E. O. Antibacterial activity of *Ocimum basilicum* essential oil and linalool on bacterial isolates of clinical importance. *International Journal of Pharmacognosy and Phytochemical Research*, v. 7, n. 6, p. 1066-1071, 2015.

SILVEIRA, W. H.; CARVALHO, GABRIEL DOMINGOS.; PECONICK, ANA PAULA. Medidas de controle do carrapato *Rhipicephalus microplus*: uma breve revisão. *PUBVET*, v. 8, p. 1136-1282, 2014.

SIQUEIRA, V. L. *Cuidados microbiológicos em cosméticos e produtos de higiene pessoal*. 2005. Disponível em: https://crq4.org.br/informativomat_394. Acesso: 15 abr. 2021.

SMYTH, J. D., WAKELIN, D. *Introduction to animal parasitology*. Cambridge: Cambridge University Press, 1994.

SOARES, A. M. D. S.; PENHA, T. A.; ARAÚJO, S. A. D.; CRUZ, E. M. O.; BLANK, A. F.; COSTA-JUNIOR, L. M. Assessment of different *Lippia sidoides* genotypes regarding their acaricidal activity against *Rhipicephalus (Boophilus) microplus*. *Revista Brasileira de Parasitologia Veterinária*, v. 25, p. 401-406, 2016.

SOCOLOVSCHI, C.; KERNIF, T.; RAOULT, D.; PAROLA, P. *Borrelia, Rickettsia, and Ehrlichia* species in bat ticks, France, 2010. *Emerging infectious diseases*, v. 18, n. 12, p. 1966, 2012.

SOLANO-GALLEG, L.; BANETH, G. Babesiosis in dogs and cats-expanding parasitological and clinical spectra. *Vet Parasitol. Vet Parasitol*, v. 181, p. 48–60, 2011.

SONENSHINE, D. E. *Biology of ticks. vol. 1*. New York: Oxford University Press, 1991.

SOTOMAIOR, C. S.; CARLI, L. M.; TANGLEICA, L.; KAIBER, B. K.; SOUZA, F. P. Identificação de ovinos e caprinos resistentes e susceptíveis aos helmintos gastrintestinais. *Revista Acadêmica*. v. 5, n. 4, p. 397-412, 2009.

SOUSA, J. P.; AZERÊDO, G. A. ARAÚJO TORRES, R.; SILVA VASCONCELOS, M. A.; CONCEIÇÃO, M. L.; SOUZA, E. L. Synergies of carvacrol and 1, 8-cineole to inhibit bacteria associated with minimally processed vegetables. *International journal of food microbiology*, v. 154, n. 3, p. 145-151, 2012.

SOUZA, A. C. C.; CERQUEIRA, R. L.; FURLONG, J.; TEIXEIRA, H. P.; MASCARENHAS, W. P. Sensibilidade do carapato *Boophilus microplus* a solventes. *Ciência Rural*, v. 33, p. 109-114, 2003.

SPINOSA, H. S.; GÓRHIAK, S. L.; BERNARDI, M. M. *Farmacologia aplicada à medicina veterinária*. 4 ed. Rio de Janeiro: Guanabara Koogan, 2006.

SPINOSA, H. S.; GÓRNIAK, S. L.; BERNARDI, M. M. *Farmacolocia aplicada à medicina Veterinária*. 5ed (Reimpr). Rio de Janeiro, RJ: Guanabara Koogan. 2014.

SPIRO, T. G.; STIGLIANI, W. M. *Química ambiental*. Pearson Prentice-Hall, 2009.

STEPEK, G.; BUTTLE, D. J.; DUCE, I. R.; LOWE, A.; BEHNKE, J. M. Natural plant cysteine proteinases as anthelmintics? *Trends. Parasitol*, v. 20, n. 7, p. 322-7, 2004.

STOTZER, E. S.; LOPES, L. B.; ECKSTEIN, C.; MORAES, M. C. M. M.; RODRIGUES, D. S.; BASTIANETTO, E. Impacto econômico das doenças parasitárias na pecuária. *Revista Brasileira de Higiene e Sanidade Animal*, v. 8, n. 3, p. 198-221, 2014.

SUNTRES, Z. E.; COCCIMIGLIO, J.; ALIPOUR, M. The bioactivity and toxicological actions of carvacrol. *Critical reviews in food science and nutrition*, v. 55, n. 3, p. 304-318, 2015.

TABARI, M. A.; YOUSSEFI, M. R.; MAGGI, F.; BENELLI, G. Toxic and repellent activity of selected monoterpenoids (thymol, carvacrol and linalool) against the castor bean tick, *Ixodes ricinus* (Acari: Ixodidae). *Veterinary parasitology*, v. 245, p. 86-91, 2017.

TAIZ, L., ZEIGER, E. *Fisiologia vegetal*. 5. ed. Porto Alegre: Artmed, 2013.

TAVARES, J. S. Biotratamento de *Rhipicephalus sanguineus* (LATREILLE, 1806) com extratos obtidos de fungos basidiomicetos do baixo Amazonas. 2015. 44f. Dissertação (Mestrado) - Universidade do Estado do Amazonas (UEA), Parintins, 2015.

TAYLOR, M. A. Recent developments in ectoparasiticides. *The Veterinary Journal*, v. 161, n. 3, p. 253-268, 2001.

TELCI, I.; BAYRAM, E.; YILMAZ, G.; AVCI, B. Variability in essential oil composition of Turkish basil (*Ocimum basilicum* L.). *Biochemical Systematics and Ecology*, v. 34, n. 6, p. 489-497, 2006.

THI, Q. V.; LAM, A. N. L. Formulation and effectiveness of Neem oil shampoo on companion animals. In: *AIP Conference Proceedings*. AIP Publishing LLC, p. 040003, 2021.

TIWARI, S.; PANDEY, S.; CHAUHAN, P. S.; PANDEY, R. Biocontrol agents in co-inoculation manages root knot nematode [*Meloidogyne incognita* (Kofoid & White) Chitwood] and enhances essential oil content in *Ocimum basilicum* L. *Industrial Crops and Products*, v.97, p. 292-301, 2017.

TORRES, F. C. Avaliação da atividade carrapaticida das frações dos óleos essenciais de citronela (*Cymbopogon winterianus*), alecrim (*Rosmarinus officinalis*) e aroeira (*Schinus molle*). 2010. 69 f. Dissertação (Mestrado em Engenharia e Tecnologia de Materiais) - Pontifícia Universidade Católica do Rio Grande do Sul, Porto Alegre, 2010.

TSUKAHARA, Y.; GIPSON, T. A.; HART, S. P.; DAWSON, L.; WANG, Z.; PUCHALA, R.; GOETSCH, A. L. Genetic selection for resistance to gastrointestinal parasitism in meat goats and hair sheep through a performance test with artificial infection of *Haemonchus contortus*. *Animals*, v. 11, n. 7, p. 1-19, 2021.

VALENTE, P. P.; AMORIM, J. M.; CASTILHO, R. O.; LEITE, R. C.; RIBEIRO, M. F. B. *In vitro* acaricidal efficacy of plant extracts from Brazilian flora and isolated substances against *Rhipicephalus microplus* (Acari: Ixodidae). *Parasitology Research*, v. 113, n. 1, p. 417-423, 2014.

VALERO, M.; SALMERON, M. C. Antibacterial activity of 11 essential oils against *Bacillus cereus* in tyndallized carrot broth. *International journal of food microbiology*, v. 85, n. 1-2, p. 73-81, 2003.

VAN DEN BROM, R.; MOLL, L.; KAPPERT, C.; VELLEMA, P. *Haemonchus contortus* resistance to monepantel in sheep. *Veterinary Parasitology*, v. 209, n. 3-4, p. 278-280, 2015

VAN WYK, J. A.; MAYHEW, E. Morphological identification of parasitic nematode infective larvae of small ruminants and cattle: A practical lab guide. *Onderstepoort Journal of Veterinary Research*, v. 80, n. 1, p. 1-14, 2013.

VAN WYK, RD.; BARON, S.; MARITZ-OLIVIER, C. An integrative approach to understanding pyrethroid resistance in *Rhipicephalus microplus* and *R. decoloratus* ticks. *Ticks and Tick-borne Diseases*, v. 7, n. 4, p. 586-594, 2016.

VANDE VELDE, F.; CHARLIER, J.; CLAEREBOUT, E. Farmer behavior and gastrointestinal nematodes in ruminant livestock-uptake of sustainable control approaches. *Front Vet Sci*, v. 16, n. 5, p. 1-12, 2018.

VERCRUYSSSE, J.; KNOX, D. P.; SCHETTERS, T. P.; WILLADSEN, P. Veterinary parasitic vaccines: pitfalls and future directions. *Trends in parasitology*, v. 20, n. 10, p. 488-492, 2004.

VIDOTTO, O.; PREVENTIVA, D. D. M. V. Complexo Carrapato-Tristeza Parasitária e outras parasitoses de bovinos. 2005. Disponível em: publication at: <https://www.researchgate.net/publication/237832290>. Acesso em: 20 jun. 2021.

VIEGAS JÚNIOR, C. Terpenos com atividade inseticida: uma alternativa para o controle químico de insetos. *Química Nova*, v. 26, n. 3, p. 390-400, 2003.

VIEIRA, L. S. Métodos alternativos de controle de nematóides gastrintestinais em caprinos e ovinos. *Tecnologia & Ciência Agropecuária*, v. 2, n. 2, p. 49-56, 2008.

VIEIRA, R. F.; SIMON, J. E. Chemical characterization of basil (*Ocimum spp.*) found in the markets and used in traditional medicine in Brazil. *Economic botany*, v. 54, n. 2, p. 207-216, 2000.

VINCENZI, M.; STAMMATI, A.; VINCENZI, A.; SILANO, M. Constituents of aromatic plants: carvacrol. *Fitoterapia*, v. 75, n. 7-8, p. 801-804, 2004.

VIVEIROS, C. T. Parasitoses gastrintestinais em bovinos na ilha de S. Miguel, Açores – Inquéritos de exploração, resultados laboratoriais e métodos de controlo. 2009. 104 f. Dissertação (Mestrado) - Universidade Técnica de Lisboa - Faculdade de Medicina Veterinária. 2009.

WALKER, A. The Genus *Rhipicephalus* (Acari, Ixodidae): A Guide to the Brown Ticks of the World; Jane B. Walker, James E. Keirans and Ivan G. Horak. *Tropical Animal Health and Production*, v. 32, n. 6, p. 417, 2000.

WALLER, P. J. Global perspectives on nematode parasite control in ruminant livestock: the need to adopt alternatives to chemotherapy, with emphasis on biological control. *Animal Health Research Reviews*, v. 4, n. 1, p. 35-44, 2003.

WATANABE, M.; NAKAO, R.; AMIN-BABJEE, S. M.; MAIZATUL, A. M.; YOUN, J. H.; QIU, Y.; WATANABE, M. Molecular screening for *Rickettsia*, Anaplasmataceae and *Coxiella burnetii* in *Rhipicephalus sanguineus* ticks from Malaysia. *Trop Biomed*, v. 32, p. 390-398, 2015.

WOLSTENHOLME, A. J.; ROGERS, A. T. Glutamate-gated chloride channels and the mode of action of the avermectin/milbemycin anthelmintics. *Parasitology*, v. 131, n. 1, p. 85-95, 2005.

WOOSTER, M. J.; WOODGATE, R. G.; CHICK, B. F. Reduced efficacy of ivermectin, abamectin and moxidectin against field isolates of *Haemonchus contortus*. *Australian veterinary journal*, v. 79, n. 12, p. 840-842, 2001.

YANG, L.; WEN, K. S.; RUAN, X.; ZHAO, Y. X.; WEI, F.; WANG, Q. Response of plant secondary metabolites to environmental factors. *Molecules*, v. 23, n. 4, p. 762, 2018.

YORK, P. *Delineamento de formas farmacêuticas*. In: Aulton, M.E., Taylor, K.M.G. (Eds.), Aulton - Delineamento de formas farmacêuticas. Elsevier, Rio de Janeiro, pp. 17–28, 2016.

ZAKARIA, Z.; AZIZ, R.; LACHIMANAN, Y. L.; SREENIVASAN, S.; RATHINAM, X. Antioxidant activity of *Coleus blumei*, *Orthosiphon stamineus*, *Ocimum basilicum* and *Mentha arvensis* from Lamiaceae family. *Int J Nat Eng Sci*, v. 2, n. 1, p. 93-95, 2008.